

*Self-assembly of a peptide amphiphile
containing l-carnosine and its mixtures
with a multilamellar vesicle forming lipid*

Article

Accepted Version

Castelletto, V., Cheng, G., Stain, C., Connon, C. and Hamley, I. ORCID: <https://orcid.org/0000-0002-4549-0926> (2012) Self-assembly of a peptide amphiphile containing l-carnosine and its mixtures with a multilamellar vesicle forming lipid. *Langmuir*, 28 (31). pp. 11599-11608. ISSN 0743-7463 doi: 10.1021/la302210b Available at <https://centaur.reading.ac.uk/28942/>

It is advisable to refer to the publisher's version if you intend to cite from the work. See [Guidance on citing](#).

To link to this article DOI: <http://dx.doi.org/10.1021/la302210b>

Publisher: American Chemical Society

All outputs in CentAUR are protected by Intellectual Property Rights law, including copyright law. Copyright and IPR is retained by the creators or other copyright holders. Terms and conditions for use of this material are defined in the [End User Agreement](#).

www.reading.ac.uk/centaur

CentAUR

Central Archive at the University of Reading

Reading's research outputs online

**Self-Assembly of a Peptide Amphiphile Containing L-Carnosine and its
Mixtures with a Multilamellar Vesicle Forming Lipid.**

V. Castelletto^{†,*}, G. Cheng^{†,‡}, C. Stain[#], C. J. Connon[†], I. W. Hamley^{†,*}

[†]*School of Chemistry, Food Science and Pharmacy, University of Reading,
Whiteknights, Reading RG6 6AD, UK*

[‡]*Currently at Department of Chemistry, University of Liverpool, Crown Street,
Liverpool L69 3BX, UK*

[#]*Centre for Advanced Microscopy, University of Reading, Whiteknights
Reading, RG6 6AF, UK*

* Authors for Correspondence.

Abstract

The self-assembly of the peptide amphiphile (PA) hexadecyl-(beta alanine-histidine) is examined in aqueous solution, along with its mixtures with multilamellar vesicles formed by DPPC (dipalmitoyl phosphatidylcholine). This PA, denoted C₁₆-βAH contains a dipeptide headgroup corresponding to the bioactive molecule L-carnosine. It is found to self-assemble into nanotapes based on stacked layers of molecules. Bilayers are found to coexist with monolayers in which the PA molecules pack with alternating up-down arrangement so that the headgroups decorate both surfaces. The bilayers become dehydrated as PA concentration increases and the number of layers in the stack decreases to produce ultrathin nanotapes comprising 2-3 bilayers. Addition of the PA to DPPC multi-lamellar vesicles leads to a transition to well-defined unilamellar vesicles. The unique ability to modulate the stacking of this PA as a function of concentration, combined with its ability to induce a multi-lamellar to uni-lamellar thinning of DPPC vesicles may be useful in biomaterials applications where the presentation of the peptide function at the surface of self-assembled nanostructures is crucial.

Introduction

L-carnosine (β -alanine-histidine, β AH) is a naturally occurring dipeptide, first isolated from a meat extract,¹⁻² present in the muscle and brain tissues of humans and other vertebrates in relatively high concentrations.

Dipeptide β AH has a range of biological activities.¹⁻² The β -alanine β A residue in β AH can react directly with oxidized carbohydrates and lipids,³ and is therefore implicated in antioxidant properties for these species.⁴⁻⁶ The antioxidant properties of the dipeptide⁵ are also able to delay senescence of cultured cells.⁷ The histidine (H) residue in β AH has the ability to bind to transition metal ions⁸⁻¹¹ or to inhibit glycation-induced protein cross-linking.¹² In particular, the prevention of cross-linking has a protective effect in inhibiting fibrillisation of α -crystallin during the formation of cataracts.¹³⁻¹⁴ This property may also be important in the proposed application of β AH to treat Alzheimer's disease.¹²

Peptide β AH is highly water soluble, but it does not self assemble in water.¹⁵ Therefore, in view of the wide range of biological activities of β AH, the construction of novel β AH supramolecular self assemblies is a crucial step to expand the potential biochemical activities of this peptide.

We have recently¹⁵ explored the use of the bulky aromatic Fmoc [Fmoc = *N*-(fluorenyl-9-methoxycarbonyl)] unit as a structure-directing agent to induce fibrillisation of β AH. We showed that Fmoc- β AH forms well defined amyloid fibrils containing β -sheets above a critical aggregation concentration.

An alternative strategy to drive β AH self assembly involves turning the dipeptide into a peptide amphiphile (PA) through the lipidation of β AH. PA-based materials are attracting immense attention due to diverse range of demonstrated potential applications in regenerative medicine.¹⁶⁻²¹ It is expected that the self assembly of the lipid- β AH will be driven by the hydrophobicity of the lipidated chain.

There is only one report in the literature about lipidated β AH.²² In that work, potential cosmetic applications of C_{16} - β AH were demonstrated since the PA is able to diffuse into the stratum corneum and the epidermal and dermal skin layers.

Here, we explore the use of the C_{16} palmitoyl lipid chain as a structure-directing agent to induce fibrillisation of carnosine. We investigate the self-assembly of C_{16} - β AH using a variety of spectroscopic and microscopic techniques, and compare it to the self assembly of Fmoc- β AH or the behaviour of β AH in water. A PA with a palmitoyl chain is chosen for study since it is compatible with the lipid chain length in DPPC for our study on incorporation in vesicles, and also because C_{16} chains are capable to drive self-assembly due to amphiphilicity, whereas shorter lipid chains may not be, and longer lipid chains can show restricted dynamics and/or crystallization.

Formatted: Subscript

Comment [IW1]: Add reference to my Soft Matter review on PAs

In addition, we describe the utility of multilamellar vesicles (MLVs) for C_{16} - β AH delivery and stabilization. We examine dipalmitoylphosphatidylcholine (DPPC) MLVs as a model system to incorporate C_{16} - β AH, with the potential for applications in delivery to cells. In particular, we investigate the incorporation of C_{16} - β AH into DPPC vesicles at C_{16} - β AH concentrations higher than the PA self assembly concentration.

Experimental

Materials.

Palmitic acid was purchased from TCI Europe. L-Carnosine, *N*-hydroxysuccinimide (HOSu), 1,3-dicyclohexylcarbodiimide (DCC), triethylamine and DPPC were purchased from Sigma-Aldrich (UK); acetonitrile (HPLC grade), tetrahydrofuran (THF) and water (HPLC) were purchased from Fisher Scientific (UK).

Synthesis of C_{16} - β AH. The C_{16} - β AH (Scheme 1) was synthesized by reaction of L-carnosine in the co-solvent of acetonitrile and water (1:2) in the presence of triethylamine with the NHS ester of palmitic acid, which was prepared by treatment of

palmitic acid with *N*-hydroxysuccinimide (HOSu) in the presence of dicyclohexylcarbodiimide (DCC) in THF at room temperature. The obtained crude product was purified by semi-preparative RP-HPLC on a Discovery® BIO wide pore C18 SUPELCO HPLC Column (10 µm particle size, L × I.D. 15 cm × 10 mm) using a mobile phase of HPLC grade water and acetonitrile with 0.1% TFA with the proportion of acetonitrile increasing linearly from 0% to 90% over 20 min and then decreasing linearly to 0% over a further 10 min. The product was white powder after lyophilization (retention time, t_R = 13.24 min): ^1H NMR (400MHz, MeOH- d_4) δ (ppm): 8.80 (d, J = 1.6 Hz, 1H), 7.35 (d, J = 1.2 Hz, 1H), 4.77 (AB_q, J = 5.2 Hz, 1H), 3.40 (td, J_1 = 6.8 Hz, J_2 = 2.0 Hz, 2H), 3.30 (m, 1H), 3.13 (dd, J_1 = 15.4 Hz, J_2 = 8.0 Hz, 1H), 2.43 (t, J = 7.2 Hz, 2H), 2.18 (t, J = 7.2 Hz, 2H), 1.60 (m, 2H), 1.30 (brs, 24H), 0.92 (t, J = 6.8 Hz, 3H). ESI-MS m/z for $\text{C}_{25}\text{H}_{44}\text{N}_4\text{O}_4$ $[\text{M}+1]^+$ calc. 465.3443, found 465.3443.

Solution and gel formation. C_{16} - β AH solutions and gels were dissolved in Milli-Q water to the desired concentration and mixed using a vortex mixer, followed by sonication in an ultrasonic bath at $\sim 50^\circ\text{C}$ for 5-10 minutes.

Preparation of Vesicles. DPPC MLVs were produced using the thin layer hydration method.²³ A weighted amount of DPPC was dissolved in ethanol in a 100 ml round bottom flask. The ethanol was removed to form a thin DPPC film on the walls of the flask under reduced pressure using a rotary evaporator. Any residual solvent was removed under a stream of N_2 . A weighed amount of water was then added to the thin DPPC film. The flask was then returned to the rotoevaporator, immersed in a water bath, and vigorously rotated for ~ 5 minutes, to reconstitute the lipid film at 50°C . The solution was then put in a flask and vortexed at 50°C for five more minutes. Vesicles of DPPC+ C_{16} - β AH were prepared following a protocol similar to that used for DPPC MLVs, but mixing a convenient amount of DPPC+ C_{16} - β AH in ethanol.

Pyrene (Pyr) Fluorescence Spectroscopy. Spectra were recorded with a Varian Cary Eclipse Fluorescence Spectrometer with samples in 4 mm inner width Quartz

cuvettes. Pyr fluorescence assays were made using a sample containing only Pyr (1.3×10^{-5} wt %), or a set of C₁₆- β AH solutions dissolved in 1.3×10^{-5} wt % Pyr. All spectra were measured from 366 to 460 nm, using $\lambda_{\text{ex}} = 339$ nm.

Circular Dichroism (CD). Spectra were recorded using a Chirascan spectropolarimeter (Applied Photophysics, UK). CD was performed on C₁₆- β AH, DPPC or C₁₆- β AH+DPPC solutions in water and placed in cover slip cuvettes (0.01 or 0.1 mm thick). Spectra are presented with absorbance $A < 2$ at any measured point with a 0.5 nm step, 1 nm bandwidth and 1 second collection time per step at 20 °C.

Fourier Transform Infra-red (FTIR) spectroscopy. Spectra were measured on a Nicolet Nexus spectrometer with DTGS detector. FTIR data was measured for C₁₆- β AH, DPPC or C₁₆- β AH+DPPC solutions in D₂O. Samples were sandwiched between two CaF₂ plate windows (spacer 0.0125 mm) Spectra were scanned 128 times over the range of 4000-900 cm⁻¹. Data was corrected by baseline subtraction.

Small-Angle X-ray Scattering (SAXS). Experiments were performed on beamline ID02 at the ESRF (Grenoble, France) and on beamline SWING at SOLEIL²⁴ (L'Orme des Merisiers, France). At ID02, 0.5 wt% C₁₆- β AH samples were placed in a glass capillary mounted in a brass block for temperature control. Micropumping was used to minimise beam damage, by displacing a drop of the sample by 0.01-0.1 mm for each exposure. The sample-to-detector distance was 1 m, and the X-ray energy was 12.46 keV. The $q = 4\pi\sin\theta/\lambda$ range was calibrated using silver behenate. Data processing (background subtraction, radial averaging) was performed using the software SAXSUtilities. At SWING, a few microlitres of sample (0.03 and 2 wt% C₁₆- β AH, 1.5 wt% DPPC or 0.03 wt% C₁₆- β AH + 1.5 wt% DPPC solutions) were injected at a slow and very reproducible flux into a quartz capillary, placed in front of the X-ray beam. After the sample was injected in the capillary and delivered in front of the X-ray beam, the flow was stopped during the SAXS data acquisition. The wavenumber q range was set to 0.004-0.5 Å⁻¹, with $\lambda = 1.03$ Å (12 keV). The images captured by the

AVIEX170170 CCD detector were radially averaged and corrected for transmitted intensity and water background using the software *Foxtrot*.

SAXS Theory. The SAXS intensity from a finite stack of unoriented bilayers can formally be written, within the monodisperse approximation, as:

$$I(q) \propto \langle F^2(q)S(q) \rangle \quad (1)$$

where $F^2(q)$ is the form factor, and $S(q)$ is the interference or structure factor, which tends to unity for weakly interacting systems.

The form factor was modelled as for a lipid bilayer, based on a sum of Gaussian functions to represent the electron density profile. The details of the model are given elsewhere.²⁵ The model assumes an electron density profile comprising Gaussians for the headgroups on either side of the bilayer and another Gaussian for the hydrocarbon chain. Scheme 2 shows a scheme of the electron density distribution along the lamellar normal and illustrates the parameters in the model. The total form factor, is taken as the contributions from the headgroup $F_H(q)$ and the hydrocarbon chain $F_C(q)$:

$$F(q) = 2F_H(q) + F_C(q) \quad (2)$$

where

$$F_H(q) = \sqrt{2\pi}\sigma_H\rho_H \exp\left(-\frac{\sigma_H^2 q^2}{2}\right) \cos(qz_H) \quad (3)$$

and

$$F_C(q) = \sqrt{2\pi}\sigma_C\rho_C \exp\left(-\frac{\sigma_C^2 q^2}{2}\right) \quad (4)$$

The fitting parameters of the model in Equations 3 and 4 are the electron densities of the headgroup (ρ_H), the thickness z_H , the electron density of the hydrocarbon chains (ρ_C), the standard deviation of the position of the Gaussian peak z_H (σ_H) and the standard deviation of the position of the Gaussian peak at z_C

(σ_c) (Scheme 2). The midpoint of the bilayer is defined as $z = 0$. In our model we assumed a Gaussian distribution of inter-headgroup thicknesses z_H , with an associated degree of polydispersity Δ_H .

To model the structure factor, $S(q)$ in Equation 1, we used the modified Caillé theory appropriate for lamellar systems. It corresponds to a multilayer structure influenced by thermal fluctuations. Details of the model are given elsewhere.²⁶⁻²⁷ Briefly, $S(q)$ is described by:

$$S(q) = N_{diff} + \sum_{N_k=N-2\sigma}^{N+2\sigma} x_K S_K \quad (5)$$

where N is the total number of layers within a scattering domain and N_{diff} accounts for a diffuse background, due to a number of uncorrelated bilayers in $S(q)$, x_k is the weight of the structure factor S_k ,²⁸ and S_k is given by:

$$S_K = N_K + 2 \sum_{m=1}^{N_K-1} (N_K - m) \cos(mqd) e^{-\left(\frac{2}{2\pi}\right)^2 q^2 \eta \gamma} \left(\frac{d}{2\pi} \right)^2 q^2 \eta \quad (6)$$

γ is the Euler constant in Equation 6, d is the layer spacing, and

$$\eta = \pi k_B T / 2d^2 (BK_c)^{1/2} \quad (7)$$

is the Caillé parameter which is a measure for the bilayer fluctuations and depends on the bilayer rigidity K_c and the bulk modulus of compression B .

The fitting parameters in Equations 5-7 are N , d , η and N_{diff} .

Congo red assay. A drop of C₁₆- β AH gel was placed onto a glass microscope slide, and partially stained using a freshly prepared and filtered 1 wt % Congo red solution in water. The partially stained gel was then placed under a cover slip and observed with the microscope through crossed polarizers, using an Olympus BX41 polarized microscope.

Transmission Electron Microscopy (TEM). Imaging was performed using a Philips CM20 TEM microscope operated at 200 kV. Droplets of C₁₆- β AH, DPPC or C₁₆- β AH+DPPC solutions solution were placed on Cu grids coated with a carbon film

(Agar Scientific, UK), stained with uranyl acetate (1 wt %) (Agar Scientific, UK) and dried.

X-ray Diffraction (XRD). X-ray diffraction was performed on stalks prepared by suspending drops of 0.8 wt% C₁₆-βAH solutions between the ends of wax-coated capillaries, and allowing them to dry. The stalk was mounted vertically onto the four axis goniometer of a RAXIS IV++ x-ray diffractometer (Rigaku) equipped with a rotating anode generator. The XRD data was collected using a Saturn 992 CCD camera.

DLS. Experiments were performed using an ALV CGS-3 system with 5003 multidigital correlator. The light source was a 20 mW He–Ne laser, linearly polarized, with $\lambda = 633$ nm. The scattering angle $\theta = 90^\circ$ was used for all the experiments.

Samples were loaded into standard 0.5 cm diameter cylindrical glass cells. DLS experiments measured the intensity correlation function of the ~~radiated-scattered~~ light

$g^{(2)}(q, t)$.²⁹

$$g^{(2)}(q, t) = 1 + A[g^{(1)}(q, t)]^2 \quad (8)$$

where A accounts for a correction factor depending on the alignment of the instrument, $q = [4\pi n \sin(\theta/2)]/\lambda$ is the scattering vector (λ = vacuum wavelength of the radiation and n = refractive index of the medium), t is the delay time, and $g^{(1)}(q, t)$ is the electric field correlation function. The program CONTIN can be used to determine the relaxation rate distribution of the system³⁰ through the modeling of the field correlation function according to:

$$g^{(1)}(t) = \int_0^\infty G(\Gamma) \exp(-\Gamma t) d\Gamma \quad (9)$$

where $G(\Gamma)$ is the relaxation rate distribution. CONTIN allows for the inverse Laplace transform in Equation 9 and provides a tool for calculating the size distribution of the system ($G(R_H)$; R_H = hydrodynamic radius). R_H is taken as the radius corresponding to the maximum in $G(R_H)$.

Comment [IW2]: Did you actually use CONTIN or the ALV software?

Cryogenic-Scanning Electron microscopy (cryo-SEM). Imaging was performed using an FEI Quanta 600F instrument. Samples were mounted onto aluminium stubs and frozen in liquid nitrogen slush at approximately -210 °C. Once frozen, samples were transferred under vacuum to a sample preparation chamber and allowed to equilibrate to the appropriate temperature prior to fracturing. C₁₆-βAH solutions were fractured at -140 °C and allowed to sublime at -90 °C for approximately 10 minutes. DPPC or C₁₆-βAH+DPPC solutions were fractured at -180 °C, to promote vitreous fracturing of individual vesicles, and then allowed to sublime at -90 °C for approximately 5 minutes. All samples were allowed to cool to -140 °C, and then coated with platinum, prior to imaging at 5 kV.

Cryogenic-Transmission Electron Microscopy (Cryo-TEM). Experiments were performed at Unilever Research, Colworth (Bedford, UK). Solutions of C₁₆-βAH, DPPC or C₁₆-βAH+DPPC, were blotted and vitrified using a Gatan Cp3 cryoplunge system. The samples were prepared at a controlled temperature of 22 °C and at a relative humidity around 90%. A 3-μl drop of each solution was placed on a 400-mesh copper TEM grid (Agar) covered with a perforated carbon film (plasma treated). The drop was automatically blotted and the sample was plunged into liquid ethane (-183 °C) to form a vitrified specimen, then transferred to liquid nitrogen (-196°C) for storage. The specimen was examined in a JEOL JEM-2100 electron microscope at 200 kV, at temperatures below -175 °C. Images were recorded digitally on a Gatan UltraScan 1000 cooled CCD camera using DigitalMicrograph (Gatan) in the low-dose imaging mode to minimize beam exposure and electron-beam radiation damage.

Results and Discussion

Self assembly of C₁₆-βAH in water

In the following we will describe the self-assembly of C₁₆-βAH in pure water. In order to highlight the secondary structure formation in C₁₆-βAH solutions, selected

results obtained for C₁₆-βAH solutions will be compared with those previously obtained by us for Fmoc-βAH and βAH solutions.¹⁵

Figure S1 (Supporting Information) shows that C₁₆-βAH dissolves in water as a solution for concentrations lower than 0.7 wt% peptide while it forms a self standing gel at higher concentration. The pH of the sample continuously drops from 6.5 to 1.5 when the concentration of the sample increases from 0.003 to 2 wt% C₁₆-βAH (inset Figure S1, Supporting Information). The isoelectric point for C₁₆-βAH is 7.85, calculated using web-based software.³⁷ Since solutions of the PA have pH~6.5-4, while self standing gels have pH ~ 1.5, the molecule will carry positive charge under these conditions.

C₁₆-βAH gel point is 0.7 wt% PA (inset Figure S1), lower than Fmoc-βAH gel point (1 wt% PA).¹⁵ The octanol-water partition coefficient, $\log P = 3.971$, calculated for C₁₆-βAH using web-based software,³¹ quantifies the hydrophobicity of C₁₆-βAH. Addition of C₁₆ leads to a substantial increase in hydrophobicity ($\log P = -3.7$ for βAH and $\log P = 0.483$ for Fmoc-βAH)¹⁵ and allows for gelation at concentrations lower than the Fmoc-βAH gel point.

Pyr fluorescence studies were performed in order to determine the critical aggregate concentration (*cac*) for C₁₆-βAH in water. This technique is routinely used to determine the critical micellar concentration for amphiphiles.³²⁻³⁵ It is based on the analysis of the fluorescence intensity of the 0-0 band at $\lambda \sim 373$ nm, denoted I_1 , and the fluorescence intensity of the ratio I_1/I_3 (I_3 : third principal vibronic band at $\lambda \sim 383$ nm).³⁶ We have used Pyr fluorescence in the past to determine the *cac* Fmoc-βAH.¹⁵

The inset in Figure 1 shows some representative examples of the fluorescence emission spectra measured for samples containing (0-6x10⁻³) wt % C₁₆-βAH and 1.3x10⁻⁵ wt % Pyr. The emission spectrum of Pyr ($\lambda_{ex} = 339$ nm) is

characterized by the strong fluorescence of the 0-0 band ($\lambda \sim 373$ nm), denoted as I_1 ³⁶ (inset Figure 1). The results show an increase of the fluorescence intensity upon increasing the C_{16} - β AH concentration with a fixed concentration of Pyr. This increase in intensity reflects the increase of the lifetime of the excited state of Pyr.³³ Pyr in water has only a very small absorption at 339 nm, which increases substantially upon transfer to a less polar domain.³³ Therefore, the enhancement of the intensity with increasing C_{16} - β AH concentration is correlated to the formation of aggregates and the insertion of the Pyr probe within the hydrophobic cores of those aggregates. This idea is supported by the measurement of the I_1/I_3 fraction discussed below.

Below the *cac*, Pyr is free in the solution and the quantum yield is high, leading to a high I_1/I_3 ratio (Figure 1). Above the *cac*, Pyr is selectively sequestered within the aggregates core and the I_1/I_3 ratio is reduced. Our results indicate a *cac* value of 2×10^{-3} wt% C_{16} - β AH. This value is significantly lower than *cac* = 7×10^{-2} wt % measured for Fmoc- β AH, proving that the C_{16} -unit drives self assembly more efficiently than the Fmoc-unit.

~~The isoelectric point for C_{16} - β AH is 7.85, calculated using web-based software³⁷. According to the data in Figure S1, C_{16} - β AH dissolved in water presents one positive charge for concentrations higher or equal to the *cac*. Solutions of the PA have pH 6.5-4, while self-standing gels have pH 1.5. ADD HERE A PARAGRAPH ABOUT SPECULATION OF DEPENDENCE OF pH on SELF-ASSEMBLY.~~

The secondary structure in solution was investigated by CD. Figure S2 (Supporting Information) shows the CD spectra obtained for 0.05 wt% C_{16} - β AH. The CD spectrum for 0.05 wt % C_{16} - β AH is characterized by a wide positive band at ~ 222 nm (Figure S2). It is possible that the positive band at 222 nm arise from stacking interactions of the aromatic-histidine units, because CD spectra with peaks in this region have been reported for Fmoc-tyrosine derivatives,³⁸⁻³⁹ and Fmoc-peptides containing R, G and D residues.⁴⁰

Comment [IW3]: You have said you removed the spectrum for 0.8% but it's still in the figure

since the CD spectra for random coil poly-L-histidine is characterized by a maximum at 222 nm.⁴¹⁻⁴³ ~~The CD spectra do not provide information about the self-assembly of C₁₆-βAH into structures with highly hydrophobic cores capable of hosting the Pyr probe, as depicted in Figure 1. Therefore, the~~ secondary structure of C₁₆-βAH aggregates was further studied by FTIR ~~at for~~ 0.8 and 2 wt% C₁₆-βAH solutions.

Figure 2a corresponds to the amide I region of the FTIR spectra. The significant peaks are listed in Table 1 and compared to those previously measured by us for βAH and Fmoc-βAH.¹⁵ The data in Figure 2a shows that C₁₆-βAH appears to self assemble into β-sheets at 0.8 and 2 wt% (Table 1). In addition to the FTIR bands previously measured for Fmoc-βAH (Figure 2a, Table 1), the peak at 1718 cm⁻¹, probably associated to the C=O vibration of the C₁₆-chain,⁴¹ appears to be exclusively associated to C₁₆-βAH concentration, as it develops upon increasing PA content and it is not present for βAH or Fmoc-βAH (Table 1).

Other regions of the FTIR spectra provide information on the lipid chain ordering present in C₁₆-βAH but obviously absent for βAH or Fmoc-βAH. Figure 2b (CH₂ stretch region 2850-2950 cm⁻¹) shows peaks in the spectra at 2952, 2917 and 2849 cm⁻¹, which arise from stretching vibrations of the CH₂ groups. In the range 1400-1500 cm⁻¹, the FTIR spectra for all samples display a peak at 1462 cm⁻¹ corresponding to the CH₂/CH₃ scissoring band⁴²⁻⁴⁴ (results not shown).

The results from FTIR spectroscopy experiments show that C₁₆-βAH self assembles in solution forming β-sheet structures, held together by the amide bonds in the βAH block, while ~~the there is also stacking of~~ histidine rings ~~are stacked within some domains in the sample~~. The lipid chains presumably associate via hydrophobic interaction.

Further evidence for amyloid formation by C₁₆-βAH was provided by Congo red dye labeling experiments. Congo red staining leading to birefringence is a characteristic of amyloid formation⁴⁵ Figure S3 (Supporting Information) shows POM

images obtained for a pure 2 wt % C₁₆-βAH gel (Figure S3a) and the same gel partially stained with Congo red (Figure S3b). The bright grey birefringence in Figure S3a denotes a supramolecular ordered structure in the sample. Figure S3b shows the regions of light green birefringence caused by the Congo red binding to the amyloid fibrils.

SAXS and XRD were used to characterize the long range order of C₁₆-βAH in solution. Figure 3a shows the SAXS profile measured for 0.5 and 2 wt% C₁₆-βAH. The XRD profile measured for a stalk dried from a 0.8 % C₁₆-βAH solution is displayed in Figure 3b.

The SAXS for 0.5 wt% peptide (Figure 3a) shows three characteristic peaks at q radii 1:1.95:2.9 (principal peak wavenumber $q_0 = 0.1348 \text{ \AA}^{-1}$) which correspond to a layered structure with a period of $d = 46.6 \text{ \AA}$.⁴⁶ This indicates a bilayer structure of the PA when the length of a C₁₆ chain (16 Å) plus βAH headgroup (7 Å) is considered. In contrast to the data for the 0.5 wt% PA solution, the intensity profile for 2 wt% C₁₆-βAH lacks defined Bragg peaks. Instead, it is characterized by a broad peak centred at $q = 0.1696 \text{ \AA}^{-1}$, which is mainly due to the form factor of the layered nanotape structure, as discussed shortly. In addition, both SAXS for 0.5 and 2 wt% peptide show a peak at $q = 0.3034 \text{ \AA}^{-1}$ ($d = 20.7 \text{ \AA}$).

The XRD profile for a stalk dried from a solution of C₁₆-βAH (Figure 3b) is characterized by two peaks with ratios of peak position 1:2 ($q_0 = 0.1784 \text{ \AA}^{-1}$) which correspond to a layered structure with a period of $d = 35.2 \text{ \AA}$.⁴⁶ Figure 3b shows a peak at $q = 0.3049 \text{ \AA}^{-1}$ ($d = 20.6 \text{ \AA}$), similar to that displayed in Figure 3a. The peaks at 4.2 \AA in Figure 3b correspond to the lateral distance between C₁₆ chains.

The SAXS curves in Figure 3a were modelled according to Equations 1-7, using the software SASfit.⁴⁷ The fitted SAXS curves are displayed in Figure 4, while the obtained parameters are listed in Table 2.

According to the results in Table 2, the number of layers in the stack, N , within the nanotapes decreases when the PA concentration increases from 0.5 to 2 wt%, from $N=7$ to $N=2-3$ (the fit shown in Figure 4b is for $N=3$, but similar quality fits could be obtained for $N=2$) corresponding to the loss of Bragg peaks on increasing concentration. From the fit parameters, it is possible to estimate a total layer thickness $l \sim (2z_H + 2\sigma_H)$ with an uncertainty Δ_H . For 0.5 wt% C₁₆- β AH (Table 2), the value of l is less than $d = 46.6$ Å as expected for a hydrated multilayer stack (i.e. $d = l + l_w$, where l_w is the thickness of the water layer). For the 2 wt% C₁₆- β AH solution, the period is reduced to $d = 35.2$ Å. The estimated value of l is similar to d , within the experimental error, suggesting a dehydrated structure.

Based on the analysis of the SAXS data for the PA at the two concentrations, model layer structures shown in Figure 5 are put forward. The 0.5 wt% solution comprises mainly hydrated bilayers with a spacing of 46 Å (Figure 5a). It is proposed that these structures coexist with monolayers with a thickness of 21 Å. This thickness is consistent with the up-down arrangement of PA molecules shown in Figure 5b. A coexistence of bilayers and thicker structures with occasional monolayers was observed by us recently for the PA C₁₆-KTTKS.⁴⁸ At higher concentration, the C₁₆- β AH tapes undergo dehydration leading to a 35 Å bilayer thickness, which is consistent with the estimated length of the molecules stacked in a bilayer allowing for some disorder in the lipid chains as shown in Figure 5c.

It is proposed that the concentration-induced reduction in observed layer spacing for C₁₆- β AH results from dehydration of the bilayers. This may be facilitated by screening of electrostatic interactions on the peptide headgroups (due to increased counterion concentration) which reduces the charge on the surface of the layers, leading to a 'coagulation' or coalescence transition, due to the attractive van der Waals interactions, analogous to phenomena in colloidal dispersions described by the DLVO theory.

The self-assembled structure formed by C₁₆-βAH was further investigated at the nanoscale for 0.05 and 1 wt% C₁₆-βAH. TEM images for 0.05 wt% C₁₆-βAH revealed extended tape structures (Figure 6a) with a persistence length of (4.4±1.3) μm and (82.9±18)nm thick. Increasing the concentration to 1 wt% C₁₆-βAH, TEM shows the formation of twisted thin fibrils (16.4±3) nm thin (Figure 6b) or rigid tapes (101.4±16.9) nm thick (Figure 6c). Our previous TEM study on Fmoc-βAH solutions, showed long twisted fibres (6.9±0.8) nm thick.¹⁵ It is evident that the formation of C₁₆-βAH wide tapes is a consequence of the self assembly of the PA into layered structures (Figure 5).

Cryo-SEM images obtained for 1 wt% C₁₆-βAH sample (Figure S4, Supporting Information) confirm the results shown in Figure 6. The sample self assembles in two different species, i.e., (129±18) nm thick tapes and very thin twisted fibres. Because of the process involved in the cryo-SEM sample preparation, the thin fibres are clustered together (Figure S4). The cryo-SEM resolution did not allow for an accurate determination of the thin fibril thickness in Figure S4, however it could be estimated to be of the order of tens of nm.

Study of DPPC and DPPC/C₁₆-βAH solutions.

In the following we will discuss the formation of DPPC vesicles containing C₁₆-βAH. Vesicles were produced in solutions containing 1.5 wt% DPPC and 1.5 wt% DPPC+0.03 wt% C₁₆-βAH, following the protocol described in the Experimental Section. We chose to study solutions containing 0.03 wt% C₁₆βAH because although this concentration is above the cac, it is low enough for the PA to be potentially incorporated into the DPPC vesicles without changing its structural parameters.

Self-assembly was first examined by TEM for 0.03 wt% C₁₆-βAH solution, and then vesicles prepared with 1.5 wt% DPPC or 1.5 wt% DPPC+0.03 wt% C₁₆-βAH were imaged.

Figure 7a shows a TEM image of peptide tapes formed at 0.03 wt% C₁₆- β AH, similar to those already discussed in Figure 6.

TEM image in Figure 7b shows the formation of collapsed vesicles in a 1.5 wt% solution of DPPC, denoting that DPPC vesicles are not robust enough to be dried onto a TEM grid without collapsing. Indeed, it has already been reported that MLVs might be affected by the high vacuum and staining process required by TEM.⁴⁹ However, DPPC vesicles were nearly spherical in shape, with three different size distributions centred on 2.6 μ m, 215 nm and 72 nm (Figure 7b).

The preparation of DPPC/C₁₆- β AH mixed vesicles resulted in three size distributions, similarly to DPPC vesicles (Figure 7c). Figure 7c is characterized by the absence of PA tapes, but the formation of nearly spherical vesicles with size distributions centred on 2.3 μ m, 630 nm and 87 nm.

The formation of DPPC/PA vesicles, was further studied by cryo-SEM. Figure 7d, 7e and 7f respectively show representative images for 0.03 wt% C₁₆- β AH, 1.5 wt% DPPC and 1.5 wt% DPPC+0.03 wt% C₁₆- β AH.

SEM images in Figure 7d show a network structure for C₁₆- β AH, comprising (40 \pm 7) Å thick fibres. DPPC vesicles shown in Figure 7e are nearly spherical, with size distributions centred on 7.2 μ m and 551 nm. Samples containing DPPC+C₁₆- β AH mixtures (Figures 7f) did not show the formation of fibril networks similar to those shown in Figure 7d, but instead nearly spherical vesicles with size distributions centred on 5.4 μ m and 508 nm.

TEM and cryo-SEM only show the formation of vesicles for the DPPC+C₁₆- β AH system with sizes similar to pure DPPC vesicles, suggesting the incorporation of the PA within the DPPC vesicles. TEM vesicle sizes, affected by sample preparation protocol and high vacuum, are smaller than cryo-SEM vesicle sizes. In order to evaluate the effect of the sample preparation protocol, cryo-TEM was used as an alternative imaging method to study DPPC and DPPC+C₁₆- β AH solutions.

Figure 8a, 8b and 8c show cryo-TEM images obtained for 0.03 wt% C₁₆-βAH, 1.5 wt% DPPC and 1.5 wt% DPPC+ 0.03 wt% C₁₆-βAH respectively.

Figure 8a shows the presence of (146.3±30.2) nm thick tapes for 0.03 wt% C₁₆-βAH. 1.5 wt% DPPC solutions (Figure 8b) contained vesicles with two size distributions centred on 540 nm and 90 nm. Similarly, 1.5 wt% DPPC+ 0.03 wt% C₁₆-βAH solutions showed the formation of vesicles 550 nm and 80 nm in size (Figure 8c). Some of the vesicle shapes in Figures 8b-c are not smoothly curved but have segments of zero curvature. This effect has been previously observed in DPPC vesicles and was ascribed to the freezing process used in cryo-TEM.⁵⁰ Nevertheless, data in Figures 8b,c show a decrease in the vesicle wall thickness from 13 nm for DPPC to 7 nm for DPPC+C₁₆-βAH. The vesicles wall thickness reduction will be discussed below in relation to SAXS results.

Dynamic light scattering (DLS) experiments were performed on 1.5 wt% DPPC+0.03 wt% C₁₆-βAH sample and the corresponding control solutions. The resulting $G(R_H)$ distributions provide two main R_H values for each solution (Figure 9). These values are $R_{H1} = 109$ nm and $R_{H2} = 436$ nm for 0.03 wt% C₁₆-βAH. According to Figure 9, $R_{H1} = 352$ nm and $R_{H2} = 2.6$ μm for 1.5 wt% DPPC while $R_{H1} = 544$ nm and $R_{H2} = 3.9$ μm for 1.5 wt% DPPC+0.03 wt% C₁₆-βAH. The increase in vesicle size for the mixed system registered by DLS was probably hindered-obscured in TEM and SEM results due to the sample preparation methods.

Cumulative data obtained by results in Figure 7-9 show that both DPPC and DPPC+ C₁₆-βAH solutions only form vesicles, with wide size distributions ranging from ~ 0.07μm to 7.2 μm . The hydrodynamic radius of DPPC vesicles increases while the thickness of the DPPC vesicles wall is reduced upon addition of C₁₆-βAH. In addition. FTIR experiments on 1.5 wt% DPPC+0.03 wt% C₁₆-βAH and the corresponding controls, show that the β-sheet structure present for 0.03 wt% C₁₆-βAH is not present in the mixed system (Figure S5, Supporting Information), in good

agreement with the absence of tapes in Figures 7c,f and Figure 8c. Figure S5 shows that at low wave numbers, there is only one FTIR absorption band at 1737 cm^{-1} , associated to the C=O vibration of the C_{16} chain.⁴¹

The information related to the vesicles wall structure is constrained by the resolution of the microscopy techniques used in Figures 7-8. Therefore, SAXS was used to get an insight into the influence of C_{16} - βAH addition to the walls of DPPC vesicles. Figure 9 shows the SAXS measured for 1.5 wt% DPPC, 0.3 wt % C_{16} - βAH and 1.5 wt% DPPC+0.3 wt% C_{16} - βAH .

The SAXS data for DPPC has been collected at $20\text{ }^{\circ}\text{C}$ which corresponds to the $\text{L}_{\beta'}$ gel phase of the lipid.⁵¹ Peaks 1, 3 and 4 measured for 1.5 wt% DPPC (Figure 10) can be indexed as the 1st, 2nd and 3rd order respectively of a 64.3 \AA layer spacing, which is consistent with the values reported in the literature for the DPPC $\text{L}_{\beta'}$ gel phase.⁵²⁻⁵³ The SAXS measured for 0.03 wt% C_{16} - βAH shows a peak centred on 21.1 \AA , in agreement with the data in Figure 3a. While SAXS confirmed the multilamellar nature of pure DPPC vesicles, the absence of Bragg reflections in the SAXS curve for 1.5 wt% DPPC+0.3 wt% C_{16} - βAH denotes a low internal order at the vesicles wall.

The SAXS curves for 1.5 wt% DPPC and 1.5 wt% DPPC+0.3 wt% C_{16} - βAH were modelled using Equations 1-7. The results are displayed in Figure 10, while the extracted parameters are listed in Table 2.

The parameter $2z_H = 40.3\text{ \AA}$ obtained for 1.5 wt% DPPC (Figure 11a), is in good agreement with the 44.2 \AA reported by Nagle⁵¹ for the headgroup peak-peak distance in the electron density profile for DPPC at $20\text{ }^{\circ}\text{C}$.

The SAXS data shows that addition of 0.03 wt% C_{16} - βAH produces unilamellar vesicles, since the data in Figure 10b are fitted to a model comprising only the bilayer form factor. The SAXS curve for this system can be modelled with z_H

and σ_H values similar to the values for DPPC suggesting that the DPPC still forms the majority component in the unilamellar vesicle walls (Figure 11b).

A transition for MLVs to unilamellar vesicles was suggested by cryo-TEM results above, which denoted a reduction in the vesicle wall thickness upon addition of C₁₆- β AH to the DPPC system (Figures 8b-c).

It has already been reported that for dimyristoylphosphatidylcholine/dimyristoylphosphatidylserine mixtures, charge repulsion between headgroups tends to destabilize multilamellar vesicles resulting into unilamellar vesicles.⁵⁴ The unilamellar structure of DPPC/C₁₆- β AH vesicles might be a consequence of electrostatic interactions. While DPPC is a zwitterionic molecule in solution, the imidazole moiety of C₁₆- β AH has an effective charge of -1 in solution.⁸ It is proposed that addition of C₁₆- β AH decorates DPPC bilayers, allowing thinning into a unilamellar structure due to charge repulsion between the headgroups.

Conclusion

In a previous paper we proved that Fmoc- β AH self assembles into fibrils such that the fibril core contains the β AH moiety while a fraction of the Fmoc unit is exposed to the aqueous environment.¹⁵ It was argued that although the Fmoc unit is highly hydrophobic, the exposure of this unit to water probably drives Fmoc- β AH self assembly into fibrils.

Here, we prove that C₁₆ drives the self assembly of C₁₆- β AH into nanotapes because of excluded volume constraints imposed by the C₁₆ chain structure. C₁₆- β AH nanotapes with a bilayer internal structure coexist with C₁₆- β AH nanotapes with a monolayer internal structure in solution. C₁₆- β AH monolayer nanotapes consist of PA molecules packed with an alternating up-down arrangement. Within the bilayer nanotapes, C₁₆- β AH adopts a single orientation allowing for the formation of β -sheets

strands between neighbouring headgroups. Bilayer nanotapes become dehydrated as PA concentration increases.

We explored the influence of DPPC vesicles on the self-assembly of a model peptide amphiphile, and *vice-versa*. We proved that the addition of the PA to DPPC MLVs does not affect the overall size of the vesicle, but induces a transition to well-defined unilamellar vesicles. C₁₆-βAH does not self-assemble into fibres in the presence of DPPC. It is probable that the PA decorates either side of the DPPC bilayers, adding a net negative charge to the DPPC bilayers surface and driving a multilamellar to unilamellar transition *via* electrostatic repulsion of the headgroups.

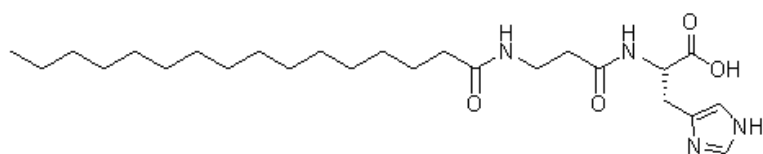
Acknowledgements This work was supported by EPSRC grants EP/F048114/1 and EP/G026203/1. We are grateful to T. Narayanan for support in the beamline ID02 (ESRF) during SAXS experiments (project code SC3235) and to J. Perez for his help during SAXS experiments at SOLEIL (project code 20110562)

Table 1: Wavenumbers (cm^{-1}) and assignments of the FTIR bands of C_{16} - βAH , βAH , and Fmoc- βAH .

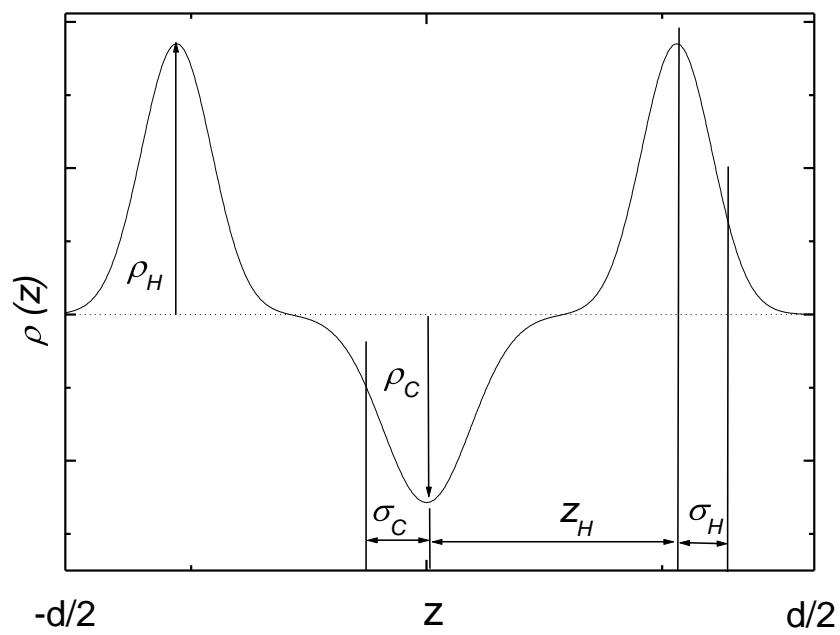
0.8 wt% C_{16} - βAH [cm^{-1}]	1 wt% βAH ¹⁵ [cm^{-1}]	1 wt% Fmoc- βAH ¹⁵ [cm^{-1}]	Assignments
1718			C=O vibration of the C_{16} chain ⁴¹
		1684	β -sheet ⁵⁵⁻⁵⁸
1675		1670	TFA counterions ⁵⁹⁻⁶⁰
1629		1636	β -sheet ⁵⁵⁻⁵⁸
1605	1594		NH_3^+ group ⁶¹⁻⁶²
1586		1591	NH_4^+ ions ¹⁵

Table 2. SAXS parameters extracted from the fittings of the experimental data shown in Figure 4 and Figure 10

Sample	$2z_H$ [Å]	Δ_H [Å]	ρ_H [rel. units]	σ_H [Å]	ρ_C [rel. units]	σ_C [Å]	N	d [Å]	η
0.5 wt% C ₁₆ -βAH	33.5	3.6	1.3e-6	3.6	-6.2e-8	3.3	6.6	46.5	0.12
2 wt% C ₁₆ -βAH	31.8	5.8	4.9e-8	4.9	-6.2e-9	2.2	3	35.3	0.2
1.5 wt% DPPC	40.3	13.4	2.5e-8	3	-2.4e-8	3	7.9	63.7	0.06
1.5 wt% DPPC+ 0.03wt% C ₁₆ -βAH	42.5	5	1.9e-7	3	-1.3e-7	3.7	-	-	-



Scheme 1 - Chemical structure of C₁₆-BAH



Scheme 2. Gaussian model of the electron density profile used to describe $F(q)$ in Equation 2.

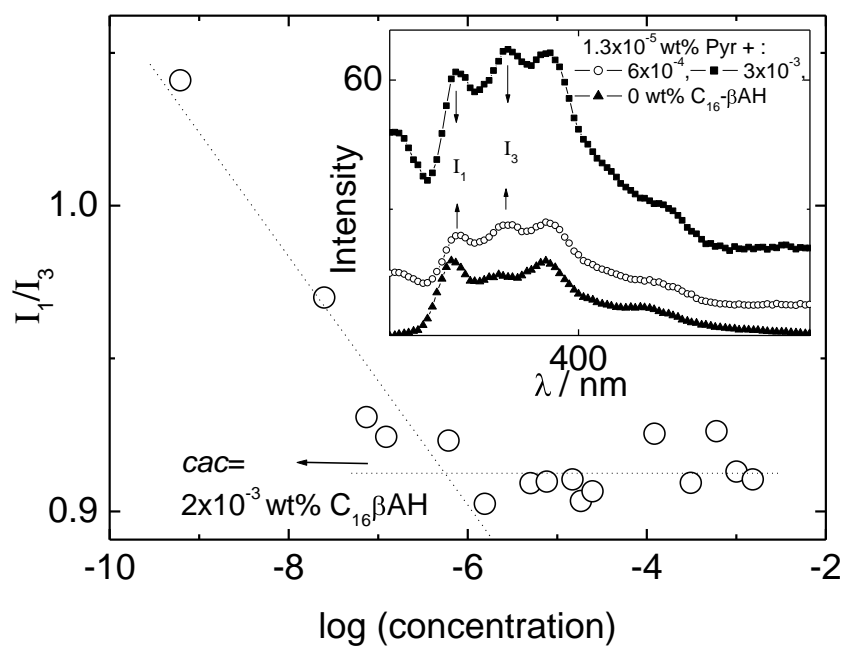


Figure 1. Determination of cac from the dependence of I_1/I_3 on $\text{C}_{16}\text{-}\beta\text{AH}$ concentration for samples containing $1.3 \times 10^{-3} \text{ wt\%}$ Pyrene.

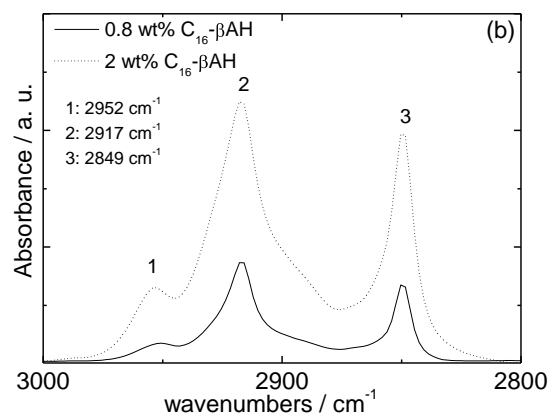
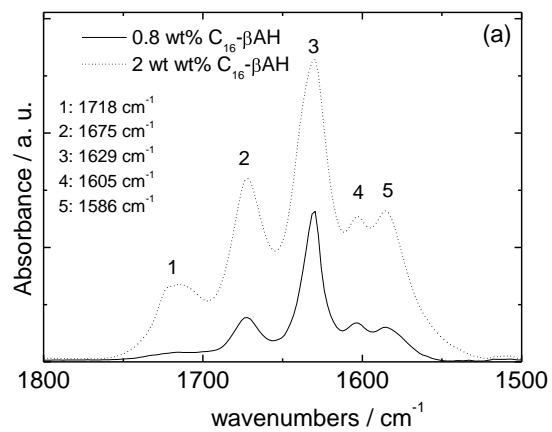


Figure 2. (a-b) FTIR data measured for 0.8 and 2 wt% $\text{C}_{16}\text{-}\beta\text{AH}$.

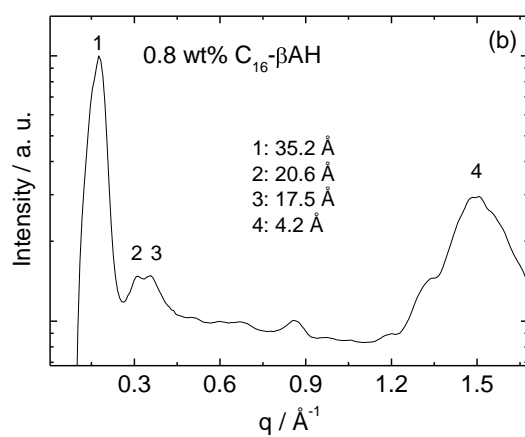
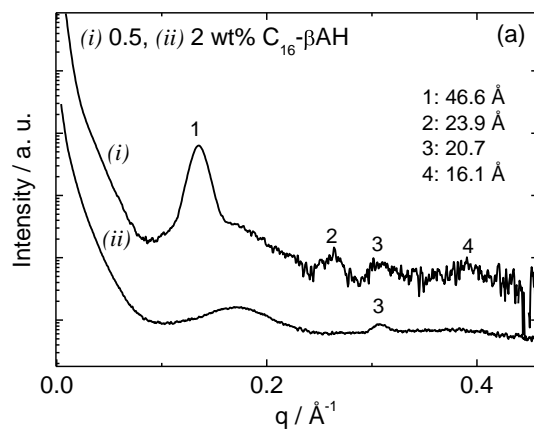


Figure 3. (a) SAXS data for 0.5 and 2 wt% C_{16} - β AH. Data for 0.5 wt% has been shifted in order to enable visualization. (b) XRD data for a stalk dried from a 0.8 wt % C_{16} - β AH solution.

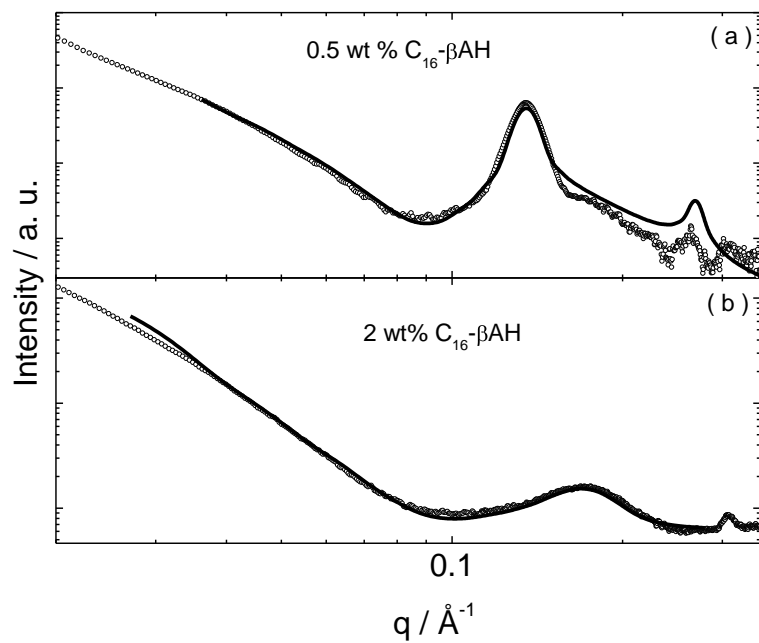


Figure 4. Fitting of the SAXS curves for (a) 0.5 and (b) 2 wt% C_{16} - β AH in Figure 3a according to a Gaussian bilayer form factor together with a modified Caillé structure factor.

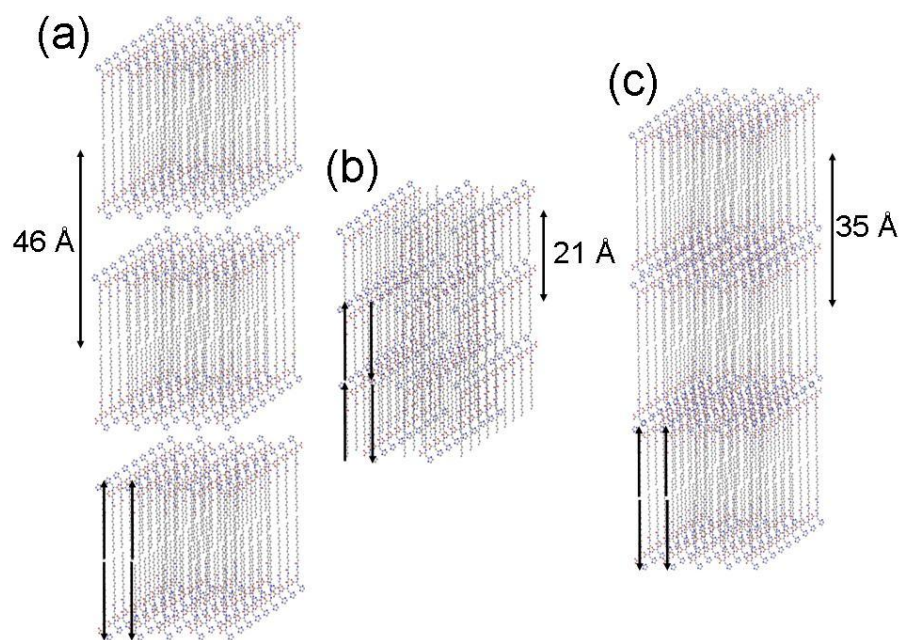


Figure 5. Model layer structures for $C_{16}\text{-}\beta\text{AH}$ in solution.

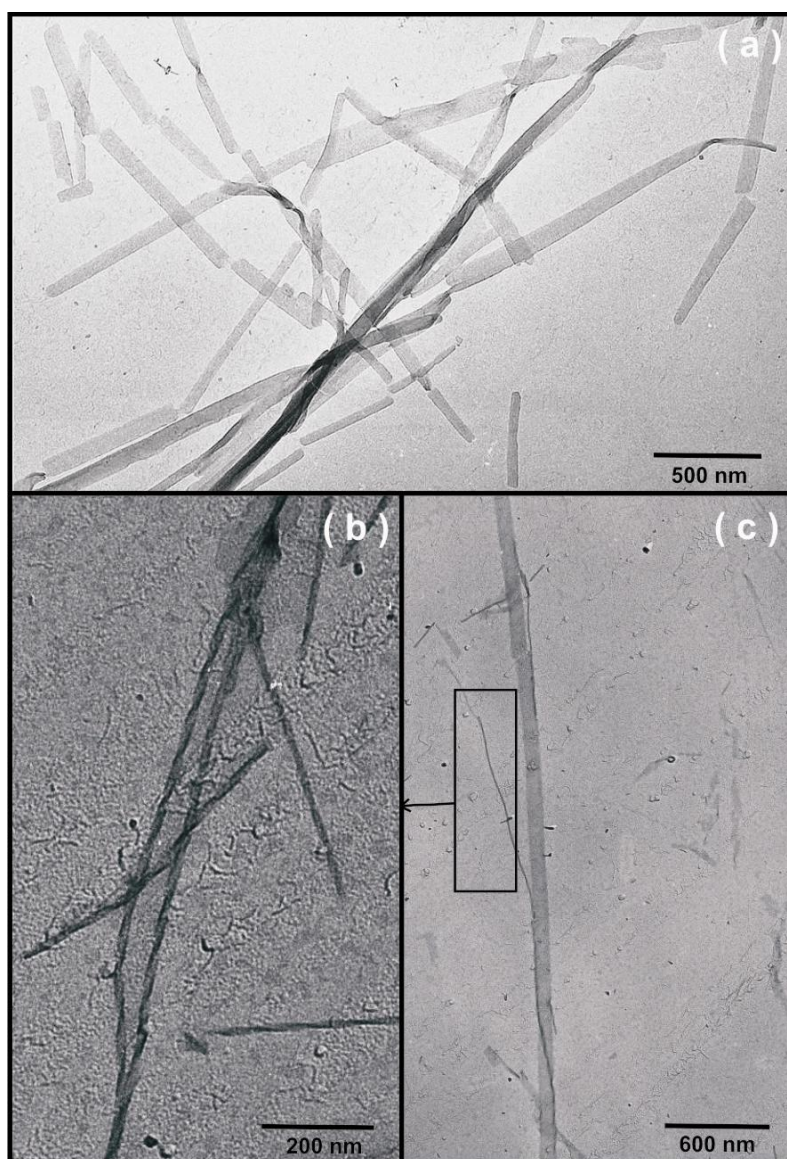


Figure 6. TEM images for (a) 0.05 and (b-c) 1 wt% C₁₆-βAH; (b) corresponds to a magnification of a twisted fibril similar to that framed in (c).

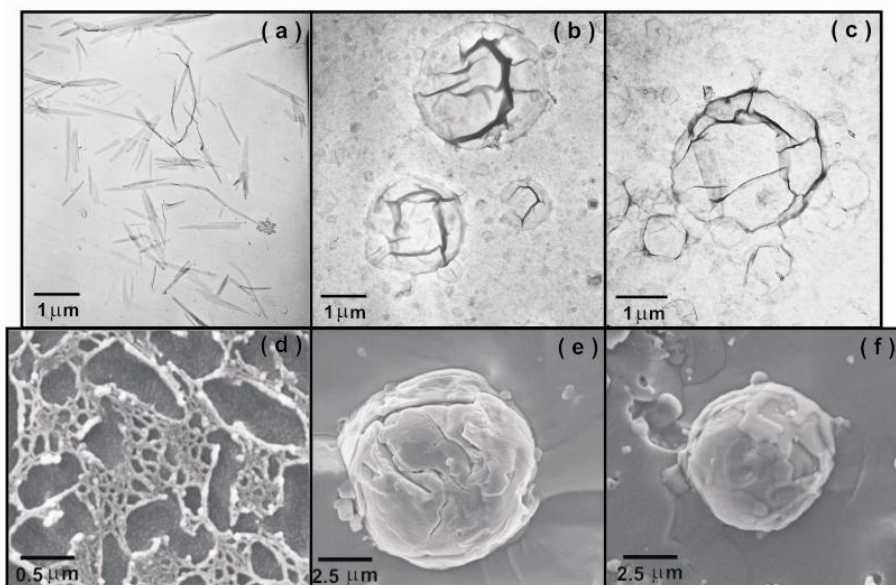


Figure 7. TEM images measured for films dried from (a) 0.03 wt% $C_{16}\text{-}\beta\text{AH}$, (b) 1.5 wt% DPPC and (c) 1.5 wt% DPPC+0.03 wt% $C_{16}\text{-}\beta\text{AH}$. Cryo-SEM for (d) 0.03 wt% $C_{16}\text{-}\beta\text{AH}$, (e) 1.5 wt% DPPC and (f) 1.5 wt% DPPC+0.03 wt% $C_{16}\text{-}\beta\text{AH}$.

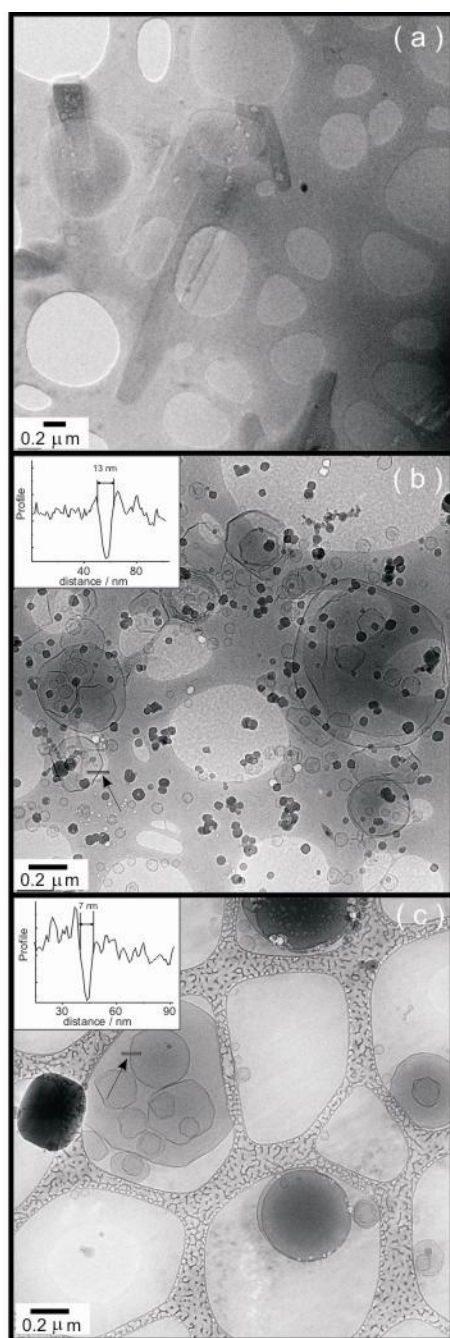


Figure 8. Cryo-TEM images for (a) 0.03 wt% C_{16} -βAH, (b) 1.5 wt% DPPC and (c) 1.5 wt% DPPC+0.03 wt% C_{16} -βAH. (b) and (c) The arrows point to the region corresponding to the intensity profile.

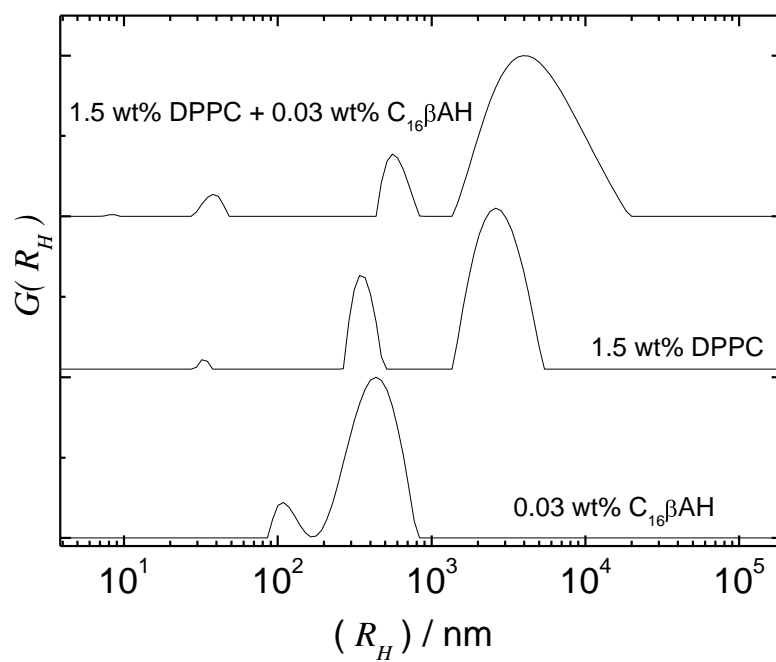


Figure 9. Distribution of hydrodynamic radii calculated for 0.03 wt% C₁₆-βAH, 1.5 wt% DPPC and 1.5 wt% DPPC+0.03 wt% C₁₆-βAH

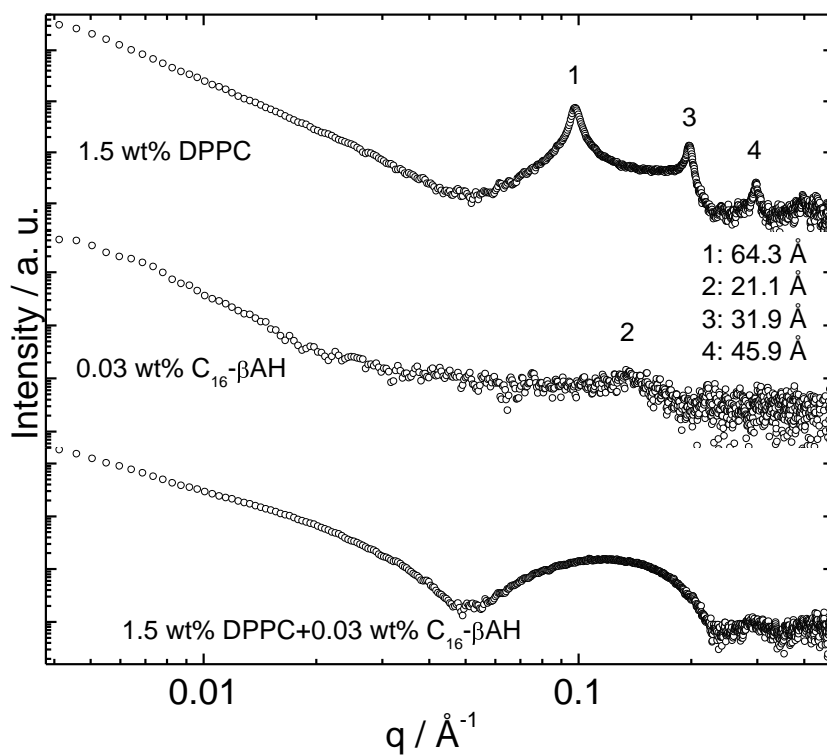


Figure 10. SAXS profiles for 0.03 wt% C₁₆-βAH, 1.5 wt% DPPC and 1.5 wt% DPPC+0.03 wt% C₁₆-βAH . The SAXS curves have been shifted in order to enable visualization of the data.

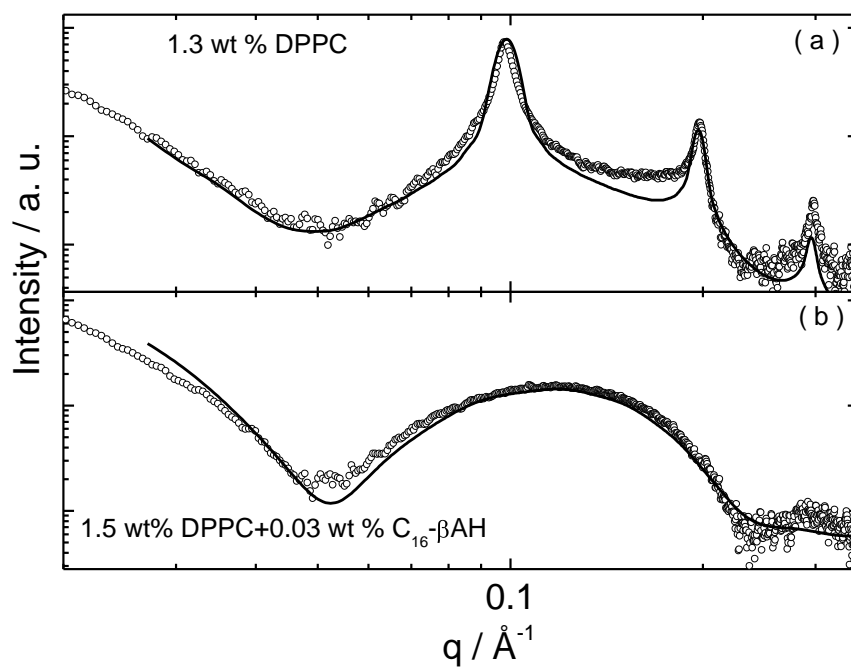


Figure 11. Fitting of the SAXS curves for (a) 1.5 wt% DPPC and (b) 1.5 wt% DPPC+ 0.03 wt% $C_{16}\text{-}\beta\text{AH}$ in Figure 9 according to a Gaussian bilayer form factor together with a modified Caillé structure factor.

References

- (1) Bonfanti, L.;Peretto, P.;De Marchis, S.;Fasolo, A. *Prog. Neurobiol.* **1999**, *59*, 333-353.
- (2) Gariballa, S. E.;Sinclair, A. J. *Age and Ageing* **2000**, *29*, 207-210.
- (3) Mehta, A. D.;Seidler, N. W. *Journal of Enzyme Inhibition and Medicinal Chemistry* **2005**, *20*, 199-203.
- (4) Hipkiss, A. R.;Michaelis, J.;Syrris, P. *FEBS Lett.* **1995**, *371*, 81-85.
- (5) Brownson, C.;Hipkiss, A. R. *Free Radical Biology and Medicine* **2000**, *28*, 1564-1570.
- (6) Boldryev, A. A.;Dupin, A. M.;Bunin, A. Y.;Babizhaev, M. A.;Severin, S. E. *Biochem. Int.* **1987**, *15*, 1105-1113.
- (7) McFarland, G. A.;Holliday, R. *Experimental Cell Research* **1994**, *212*, 167-175.
- (8) Torreggiani, A.;Fini, G.;Bottura, G. *Journal of Molecular Structure* **2001**, *565-566*, 341-346.
- (9) Torreggiani, A.;Bonora, S.;Fini, G. *Biopolymers* **2000**, *57*, 352-364.
- (10) Lenz, G. R.;Martell, A. E. *Biochemistry* **1964**, *3*, 750-753.
- (11) Lenz, G. R.;Martell, A. E. *Biochemistry* **1964**, *3*, 745-750.
- (12) Hobart, L. J.;Seibel, I.;Yeargans, G. S.;Seidler, N. W. *Life Sci.* **2004**, *75*, 1379-1389.
- (13) Seidler, N. W.;Yeargans, G. S.;Morgan, T. G. *Arch. Biochem. Biophys.* **2004**, *427*, 110-115.
- (14) Attanasio, F.;Cataldo, S.;Fisichella, S.;Nicoletti, S.;Nicoletti, V. G.;Pignataro, B.;Savarino, A.;Rizzarelli, E. *Biochemistry* **2009**, *48*, 6522-6531.
- (15) Castelletto, V.;Cheng, C.;Greenland, B. W.;Hamley, I. W.;Harris, P. J. F. *Langmuir* **2011**, *27*, 2980-2988.
- (16) Hamley, I. W. *Soft Matter* **2011**, *7*, 9533-9534.
- (17) Matson, J. B.;Zha, R. H.;Stupp, S. I. *Current Opinion in Solid State & Materials Science* **2011**, *15*, 225-235.
- (18) Silva, G. A.;Czeisler, C.;Niece, K. L.;Beniash, E.;Harrington, D. A.;Kessler, J. A.;Stupp, S. I. *Science* **2004**, *303*, 1352-1355.
- (19) Tysseling-Mattiace, V.;Sahni, V.;Niece, K.;Birch, D.;Czeisler, C.;Fehlings, M.;Stupp, S.;Kessler, J. *J. Neurosci.* **2008**, *28*, 3814-3823.
- (20) Shah, R.;Shah, N.;Lim, M.;Hsieh, C.;Nuber, G.;Stupp, S. *PNAS* **2010**, *107*, 3293-3298.
- (21) Bond, C.;Angeloni, N.;Harrington, D.;Stupp, S.;McKenna, K.;Podlasek, C. *Journal of Sexual Medicine* **2011**, *8*, 78-89.
- (22) Lintner, K.;Peschard, O. *International Journal of Cosmetic Science* **2000**, *22*, 207-218.
- (23) Szoka, F.;Papahadjopoulos, D. *PNAS* **1978**, *75*, 4194-4198.
- (24) David, G.;Perez, J. *J. Appl. Cryst.* **2009**, *42*, 892-900.
- (25) Pabst, G.;Rappolt, M.;Amenitsch, H.;Laggner, P. *Physical Review E* **2000**, *62*, 4000-4008.
- (26) Zhang, R.;Suter, R. M.;Nagle, J. F. *Phys. Rev. E* **1994**, *50*, 5047-5060.
- (27) Caillé, M. A. C. *R. Acad. Sci. Paris* **1972**, *274*, 891-893.
- (28) Zhang, R.;Suter, R. M.;Nagle, J. F. *Phys. Rev. E* **1964**, *50*, 5047-5060.
- (29) Berne, B. J.;Pecora, R. (1976) *Dynamic Light Scattering*, Wiley-Interscience, New York.
- (30) Provencher, S. W. *Makromolekulare Chemie* **1979**, *180*, 201.
- (31) www.molinspiration.com. (2010).

- (32) Johnsson, M.;Hansson, P.;Edwards, K. *J. Phys. Chem. B* **2001**, *105*, 8420-8430.
- (33) Wilhelm, M.;Zhao, C.-L.;Wang, Y.;Xu, R.;Winnik, M. A.;Mura, J.-L.;Riess, G.;Croucher, M. D. *Macromolecules* **1991**, *24*, 1033-1040.
- (34) Astafieva, I.;Zhong, X. F.;Eisenberg, A. *Macromolecules* **1993**, *26*, 7339-7352.
- (35) Astafieva, I.;Khogaz, K.;Eisenberg, A. *Macromolecules* **1995**, *28*, 7127-7134.
- (36) Johnsson, M.;Silvander, M.;Karlsson, G.;Edwards, K. *Langmuir* **1999**, *15*, 6314-6325.
- (37) (Innovagen's Peptide Property Calculator) Copyright © 2012 Innovagen AB.
- (38) Yang, Z.;Xu, B. *Chemical Communications* **2004**, 2424-2425
- (39) Yang, Z.;Gu, H.;Fu, D.;Gao, P.;Lam, J. K. W.;Xu, B. *Adv. Mat.* **2004**, *16*, 1440.
- (40) Cheng, G.;Castelletto, V.;Jones, R.;Connon, C. J.;Hamley, I. W. *Soft Matter* **2010**, in press.
- (41) Bellamy, L. J. (1975) *The Infra-Red Spectra of Complex Molecules*, London.
- (42) Zhang, Y. P.;Lewis, R.;Hodges, R. S.;McElhaney, R. N. *Biochemistry* **1992**, *31*, 11572-11578.
- (43) Reis, O.;Winter, R.;Zerda, T. W. *Biochimica et Biophysica Acta - Biomembranes* **1996**, *1279*, 5-16.
- (44) Moore, D. J.;Rerek, M. E.;Mendelsohn, R. *J. Phys. Chem. B* **1997**, *101*, 8933-8940.
- (45) Hamley , I. W. *Angewandte Chemie, International Edition in English* **2007**, *46*, 8128-8147.
- (46) Safinya, C. R.;Sirota, E. B.;Bruinsma, R. F.;Jeppesen, C.;Plano, R. J.;Wenzel, L. J. *Science* **1993**, *261*, 588-591.
- (47) Kohlbrecher, J.;Bressler, I. (2011) Software package SASfit for fitting small-angle scattering curves.
- (48) Castelletto, V.;Hamley, I. W.;Adamcik, J.;Mezzenga, R.;Gummel, J. *Soft Matter* **2012**, *8*, 217-226.
- (49) Khosravi-Darani, K.;Pardakhty, A.;Honarpisheh, H.;Rao, V. S. N. M.;Mozafari, M. R. *Micron* **2007**, *38*, 804-818.
- (50) Kim, S. H.;Haimovich-Caspi, L.;Omer, L.;Talmon, Y.;Franses, E. I. *Journal of Colloid and Interface Science* **2007**, *311*, 217-227.
- (51) Nagle, J. F.;Tristram-Nagle, S. *Biochimica et Biophysica Acta* **2000**, *1469*, 159-195.
- (52) Sun, W.-J.;Tristram-Nagle, S.;Suter, R. M.;Nagle, J. F. *Biophysical Journal* **1996**, *71*, 885-891.
- (53) McManus, J. J.;Radler, J. O.;Dawson, K. A. *Langmuir* **2003**, *19*, 9630-9637.
- (54) Madden, T. D.;Tilcock, C. P. S.;Wong, K.;Cullist, P. R. *Biochemistry* **1988**, *27*, 8724-8730.
- (55) Haris, P.;Chapman, D. *Biopolymers* **1995**, *37*, 251-263.
- (56) Stuart, B. (1997) *Biological Applications of Infrared Spectroscopy*, Wiley, Chichester.
- (57) Rosler, A.;Klok, H.-A.;Hamley , I. W.;Castelletto, V.;Mykhaylyk, O. O. *Biomacromolecules* **2003**, *4*, 859-863.
- (58) Miyazawa, T.;Blout, E. R. *Journal of the American Chemical Society* **1961**, *83*, 712-719.
- (59) Gaussier, H.;Morency, H.;Lavoie, M. C.;Subirade, M. *Appl. Environ. Microbiol.* **2002**, *68*, 4803-4808.

- (60) Pelton, J. T.;McLean, L. R. *Anal. Biochem.* **2000**, 277, 167-176.
- (61) Lukton, A.;Sisti, A. *The Journal of Organic Chemistry* **1961**, 26, 617-619.
- (62) Baran, E. J.;Parajon-Costa, B. S.;Rojo, T.;Saez-Puche, R.;Fernandez, F.;Totaro, R. M.;Apella, M. C.;Etcheverry, S. B.;Torre, M. H. *Journal of Inorganic Biochemistry* **1995**, 58, 279-289.

Comment [IW4]: Give web address for innovagen. Cheng et al. reference is out of date

Table of Contents Figure

