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Effects of dietary selenium supplementation on tissue selenium distribution and glutathione peroxidase activity in Chinese Ring Necked Pheasants

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The objective of this study was to determine the concentration of total selenium (Se) and the proportions of total Se comprised as selenomethionine (SeMet) and selenocysteine (SeCys) in the postmortem tissues of female pheasants (*Phasianus Colchicus Torquator*) offered diets that contained graded additions of selenised-enriched yeast (SY) or a single comparative dose of sodium selenite (SS). Thiobarbituric acid reactive substances (TBARS) and tissue glutathione peroxidase (GSH-Px) activity of breast (Pectoralis Major) were assessed at 0 and 5 days postmortem. A total of 216 female pheasant chicks were enrolled into the study. Twenty-four birds were euthanased at the start of the study, and samples of blood, breast muscle, leg muscle (M. Peroneus Longus and M. Gastrocnemius), heart, liver, kidney and gizzard were collected for determination of total Se. Remaining birds were blocked by live weight and randomly allocated to one of four dietary treatments ($n = 48$ birds/treatment) that either differed in Se source (SY v. SS) or dose (control (0.17 mg total Se/kg), SY-L and SS-L (0.3 mg/kg total Se as SY and SS, respectively) and SY-H (0.45 mg total Se/kg)). Following 42 and 91 days of treatment, 24 birds per treatment were euthanased, and samples of blood, breast muscle, leg muscle, heart, liver, kidney and gizzard were retained for determination of total Se and the proportion of total Se comprised as SeMet or SeCys. Whole blood GSH-Px activity was determined at each time point. Tissue GSH-Px activity and TBARS were determined in breast tissue at the end of the study. There were increases in both blood and tissues to the graded addition of SY to the diet ($P < 0.001$), but the same responses were not apparent with the blood and tissues of selenite-supplemented birds receiving a comparable dose (SY-L v. SS-L). Although there were differences between tissue types in the distribution of SeMet and SeCys, there were few differences between treatments. There were effects of treatment on erythrocyte GSH-Px activity ($P = 0.012$) with values being higher in treatments SY-H and SS-L when compared with the negative control and treatment SY-L. There were no effects of treatment on tissue GSH-Px activity, which is reflected in the overall lack of any treatment effects on TBARS.

Keywords: pheasant, selenium, selenocysteine, selenomethionine, glutathione peroxidase

Implications

The results of this study indicate that selenium (Se) source can influence the distribution of total Se within the tissues of game birds. Furthermore, the responses to dietary Se and the distribution of Se species within the tissues of unimproved, wild type birds are markedly different from those of developed commercial breeds. This would imply that the Se requirements of game type birds may be appreciably different from those of commercial genotypes, despite the fact that literature suggests that they are similar.

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Introduction

Selenium (Se) is an important trace element, essential for all selenoproteins, which contain a functional selenocysteine (SeCys) core. Before the late 1950s, much of the work on Se focussed on the toxic and detrimental effects that high concentrations of dietary Se had on animal health (Underwood and Suttle, 2001). However, during the latter part of the 1950s, it was noted that Se played an important physiological role in higher animals, as Se-deficient animals were seen to suffer from liver necrosis and muscular dystrophy (Underwood and Suttle, 2001). In the early part of the 1970s, a specific biological role for Se became apparent with

the discovery of the first selenoprotein, glutathione peroxidase (GSH-Px; Rotruck *et al.*, 1973). GSH-Px catalyses the reduction of lipid and hydrogen peroxides to less harmful hydroxides via the oxidation and subsequent reduction of the SeCys active centre of the enzyme (Surai, 2006). The antioxidant functions of Se, via the actions of tissue GSH-Px, have been shown to persist *postmortem* in poultry muscle tissue (Juniper *et al.*, 2011) and delay the onset of oxidation reactions (DeVore *et al.*, 1983).

During the last 40 years, a number of selenoproteins have been identified. These include those involved in the protection of cellular membranes against the effects of oxidative stress, namely, the GSH-Pxs and thioredoxin reductases, deiodinases, which are essential for proper thyroid function, and selenoproteins S, W and P, the latter of which is involved in Se transport. The expression and subsequent activity of all selenoenzymes is dependent upon an adequate Se supply. Furthermore, Se within an organism is not evenly distributed as there is a hierarchy of selenoenzymes (Burke *et al.*, 2005). Consequently, when Se is limiting there maybe full expression of some selenoenzymes and restricted expression of others.

Pheasants are typically wild type and have not undergone the same rigorous genetic selection that has been applied to commercial lines of poultry to improve rates of growth and feed conversion efficiency. However, despite this lack of selection, the Se requirements of Ring-necked Pheasants are given to be similar to those of growing turkeys (National Research Council (NRC), 1994). Differences in responses to dietary Se have been reported between a commercial porcine genotype and a pure breed (Banoch *et al.*, 2009), indicating that Se metabolism and subsequent requirements may differ between animals selected for growth and efficiency when compared with unimproved breeds.

The objective of this study was to determine tissue Se distribution, erythrocyte and tissue GSH-Px activity in growing pheasants fed diets containing additions of dietary Se from either sodium selenite (SS) or seleno-yeast (SY) sources.

Material and methods

The work was conducted in accordance with the UK Animals (Scientific procedures) Act 1986 (Home office, 1986) and procedures were undertaken by competent technical staff holding appropriate license authorities under the Act. All birds were housed on white wood shavings with a stocking density of 1.3 m²/bird. Fresh water was available at all times via a gravity fed bell drinker.

A total of 500 pheasant chicks (Holme Park Game Hatcheries, Wokingham, Berks, UK) were purchased and brooded according to commercial shoot specifications. During this pre-trial period all chicks were offered a starter ration, formulated to meet nutritional requirements (NRC, 1994) that had not been augmented with supplementary Se. At 5 weeks of age, when gender identification was possible following sufficient plumage development, all male pheasant chicks were removed and all female pheasant chicks weighed

to provide data for experimental blocking. A total of 216 female chicks were selected on the basis of live weight (mean BW 527.6 ± 4.1 g). Of these 216 selected birds, 24 were randomly selected to form a pretreatment slaughter group, which were subsequently euthanased by cervical dislocation. Samples of blood were taken from all euthanased birds and tissue samples (breast, leg, heart and liver) were retrieved from six randomly selected birds for determination of pretreatment Se concentrations. All remaining birds were blocked by live weight and randomly allocated to one of four dietary treatments: an unsupplemented control (Con), supplemented with 0.08 or 0.23 mg Se/kg dry matter (DM) of SY (SY-L and SY-H, respectively; Sel-Plex, Alltech, Nicholasville, KY, USA), or 0.08 mg/kg DM of an inorganic Se source (SS-L), to achieve final total Se concentrations of 0.30, 0.45 and 0.30 mg/kg DM, respectively. All diets were formulated to meet the nutritional requirements of growing Ring-necked Pheasants (NRC, 1994).

Birds were penned in groups of eight birds per pen with six pens per treatment. All birds remained within established pen groups throughout the study. Pens were bedded with white wood shavings, which were replenished regularly to maintain litter quality. Each pen was equipped with one bell type automatic drinker and one plastic poultry tube feeder with a feeding circumference of 60 cm. Birds were offered the experimental diets, with respect to treatment designation, for the duration of the study.

Feeding stuffs and feed intake

Diets were formulated and manufactured by Dairy Direct (Mill Feed Services, Anglia, UK). Before manufacture, the Se content of individual compositional ingredients of the diets was sent to UT2A laboratories (Pau, France) for determination of background Se of compositional ingredients (Table 1). The quantities of either SY or SS supplements required to achieve target doses with respect to treatment were calculated and mineral supplements were subsequently manufactured. Following manufacture, representative samples of each mineral supplement were analysed to confirm the correct level of Se.

Birds were offered their respective diets (according to experimental design) *ad libitum* daily throughout the study. Feed offered and refused were weighed and recorded daily with DM contents being determined on a weekly basis. Weekly samples of each diet were stored at -20°C for subsequent laboratory determination (Euorofins, Wolverhampton, UK) of DM, CP, starch, NDF, total sugars, oil, ash and Se.

Following 42 days exposure to experimental diets (T₄₂), four birds were randomly selected from each pen (*n* = 24 birds per treatment) and euthanased by cervical dislocation, and samples of blood and tissue (breast, leg, gizzard, kidneys and liver) retrieved for determination of Se concentrations. Following an additional 49 days exposure (T₉₁), all remaining birds (*n* = 24 birds per treatment) were euthanased by cervical dislocation, and samples of blood and tissue (breast, leg, gizzard, kidneys and liver) retrieved for determination of Se concentrations.

Table 1 Constituent ingredients, inclusion rates, Se contents and estimated Se contents of experimental diets

Ingredient	Inclusion (g/kg as fed unless otherwise stated)	Se content (mg/kg as fed)	Se (mg/kg as fed)
Wheat	719.75	0.112	0.081
Hydro soya	200.0	0.263	0.053
Full fat soya	37.5	0.023	0.001
Vitamin and mineral premix	25.0	1.359	0.024
Di-calcium phosphate	17.25	0.137	0.013
Diet Se estimates (unsupplemented)	—	—	0.172
Seleno-yeast	^a	2027	—
Sodium selenite	^a	8628	—
SY-L SY inclusion (g/tonne as fed)	64.1	SY-L estimated total Se (mg/kg as fed)	0.30
SY-H SY inclusion (g/tonne as fed)	138.1	SY-H estimated total Se (mg/kg as fed)	0.45
SS-L SS inclusion (g/tonne as fed)	15.1	SS-L estimated total Se (mg/kg as fed)	0.30

Se = selenium; SY-L = lowest level of selenised-enriched yeast; SY-H = highest level of selenised-enriched yeast; SS-L = lowest level of sodium selenite.

^aDetermined by experimental protocol.

Vitamin and mineral premix composition: calcium pantothenate 400 mg/kg, riboflavin (B₂) 200 mg/kg, vitamin B₁₂ 320 µg/kg, thiamin (B₁) 40.0 mg/kg, pyridoxine (B₆) 40.0 mg/kg, biotin 2.0 mg/kg, cobalt (cobalt carbonate) 10 mg/kg, iodine (as sulphate) 40 mg/kg, copper (as sulphate) 800 mg/kg, iron (as sulphate) 400 mg/kg, manganese (manganous oxide) 3200 mg/kg, zinc (zinc oxide) 2400 mg/kg, choline chloride 1740 mg/kg, phosphorus 140.6 g/kg, vitamin A 400 000 IU/kg, vitamin D₃ 120 000 IU/kg, vitamin E 800 mg/kg, vitamin K 80 mg/kg, folic acid 40 mg/kg, niacin 800 mg/kg.

At each time point (T₄₂ and T₉₁) blood was collected following exsanguination into three pretreated lithium heparin tubes per bird (Sarstedt, Leicester, UK). One tube was sent for determination of whole blood GSH-Px activity (Veterinary Laboratories Agency, Shropshire, UK), one for determination of Se content (UT2A) and the third centrifuged at 1252 × g for 10 min at room temperature in a bench top centrifuge (Weiss-Gallenkamp, Loughborough, UK) after which the plasma fraction was decanted and sent for determination of Se content (UT2A).

Following slaughter at T₄₂, one bird per pen (six birds per treatment) was randomly selected and dissected. The ventral aspect of the carcass was skinned, and a sample of breast tissue (*M. Pectoralis Major* (PM)) from the left side of the carcase retained for Se analysis. The musculature of the left thigh was exposed and samples of muscle tissue (*M. Gastrocnemius* (MG) and *M. Peroneus Longus* (PL)) retained for Se analysis. The heart, liver, kidneys and gizzard were removed from the body cavity and retained for Se analysis.

At the T₉₁ slaughter point, the remaining 24 birds per treatment were plucked and dressed, and one bird per pen (six birds per treatment) was randomly selected and underwent dissection as previously described. In addition, the whole breasts were removed and skinned from 12 birds per treatment, packaged in a zip-lock bag, refrigerated and transported in a cool box on the day of slaughter to the University of Bristol for determination of thiobarbituric acid reactive substances (TBARS) and tissue GSH-Px activity.

Total Se in feeding stuffs, whole blood samples, plasma and tissues was determined according to the method of Mester *et al.* (2006). Briefly, 1 g of each sample was mineralised in 4 ml 16 M HNO₃ and 2 ml 9.8 M H₂O₂ within a closed-vessel heating block system. The solution was further diluted with water and Se subsequently determined using inductively coupled plasma mass spectrometry (ICP-MS; Perkin Elmer Elan 6100 ICPMS, Waltham MA, USA).

The selenised amino acid (AA) contents of blood were determined by speciation using the method of Bierla *et al.* (2008a). Samples were initially incubated for 5 h with DL-dithiothreitol and iodoacetamide to reduce and alkylate SeCys. Samples were then spiked with selenomethionine (SeMet⁷⁷), and subsequently incubated for 24 h at 37°C with a mixture of protease and lipase maintained at a pH 7.5. Following incubation, the mixture was centrifuged and the supernatant separated and purified by size exclusion liquid chromatography. Aliquots of the supernatant were analysed by reversed-phase HPLC using an ICP-MS equipped with a collision cell (Perkin Elmer Elan 6100 ICP-MS). The selenised AA content of tissues (breast, thigh, liver and kidney) was determined by speciation according to the method of Bierla *et al.* (2008b). Samples were mixed and sieved after which a representative subsample was taken. Urea was added to the subsample and the subsample sonicated after which it was reduced, alkylated and submitted to proteolysis. The extract was then purified by size exclusion chromatography and the AAs quantified by reverse phase HPLC-ICP-MS.

Breast tissue was stored at 3°C for 5 days after which TBARS were determined by the method of Tarladgis *et al.* (1960), modified by the use of a Büchi 321 distillation unit (Büchi Labortechnik AG, Postfach, Switzerland). Tissue GSH-Px activity was determined in samples of breast tissue taken immediately post slaughter and from breast tissue that had undergone 5 days aging in MAP, using the coupled assay procedure of Paglia and Valentine (1967), modified by DeVore and Greene (1982) and Daun *et al.* (2000). Results are presented as nmol of NADPH oxidised per mg of protein/min. GSH-Px activity in whole blood was determined using the Olympus AU400 Chemistry Analyser (Olympus UK, Watford, UK) based on the method of Anderson *et al.* (1978). Results are presented as units per ml.

The study was of a complete block randomised design. Statistically significant differences between treatments for

all traits at T_{42} and T_{91} were determined by ANOVA using a GLM (Minitab v. 15, Minitab Inc., State College, PA, USA). Sources of variation within the model included treatment (3 d.f.) and block (5 d.f.). Data pertaining to animal performance (feed intake, live weight gain and feed conversion ratio) were analysed on a group pen basis. Data pertaining to carcass characteristics (carcass weight, dressing out percentage), Se analysis and meat quality were analysed on an individual bird basis. Results are presented as least square means with the standard error of the mean (s.e.m.). Tukey's simultaneous test was used to establish statistical differences ($P < 0.05$) among individual treatment means.

Results

With the exception of Se concentrations, which were dictated by treatment structure, there were no appreciable differences in any determined or calculated feed nutritional parameter. Diets had a mean (\pm s.d.) DM content of 871 ± 1.1 g/kg fresh weight (FW), CP content of 204.8 ± 3.5 g/kg FW, oil content of 27.8 ± 0.5 g/kg FW and estimated Metabolisable Energy content of 12.9 ± 0.6 MJ/kg FW. Diets had a mean (\pm s.d.) Se concentration of 0.12, 0.30, 0.46 and 0.27 ± 0.02 mg/kg FW for treatments Con, SY-L, SY-H and SS-L, respectively.

Although intakes and rates of gain were lower than anticipated, there were no appreciable effects of treatment on any aspect of bird physical performance; birds within all treatments started and finished at similar weights and consumed similar quantities of feed over the course of the study (Table 2).

There were effects of treatment ($P < 0.001$) at both T_{42} and T_{91} on the whole blood total Se contents (Table 3). At both time points the total Se content of whole blood from Con birds did not change appreciably from those recorded at T_0 . Total Se values of SS-L birds were similar to those of Con and SY-L birds at T_{42} , although whole blood total Se contents were greater in SY-L birds when compared with both T_0 and Con birds ($P < 0.05$). Whole blood total Se contents at T_{42} were greater in treatment SY-H when compared with Con and SS-L ($P < 0.05$). At T_{91} , whole blood total Se values of treatments SY-L and SY-H were similar when compared with each other, but were greater than Con and SS-L ($P < 0.05$). This trend is also reflected in the total Se content of plasma at T_{91} ; plasma total Se values of SY-L and SY-H birds were similar when compared with each other but greater than Con and SS-L birds ($P < 0.05$).

The Se content of tissues at T_0 differed between the different tissue types (Table 4) with total Se concentrations being greatest in kidney, followed by liver, heart, gizzard and skeletal muscle. Although there were effects of treatment on the total Se content of each tissue at successive time points (T_{42} and T_{91}), the hierarchy of tissue Se concentrations seen at T_0 was maintained within the treatment structure at each time point. There were effects ($P < 0.005$) on the total Se content of skeletal muscles at T_{42} to the graded addition of SY to the diet. Total Se values in the skeletal tissues of Con birds at both T_{42} and T_{91} were not appreciably different from those recorded at T_0 . Similarly, the total Se contents of skeletal tissue of SS-L birds at both time points did not differ from those of Con birds or T_0 . However, total Se values of

Table 2 Physical performance of female Ring-necked Pheasants, following 91 days exposure to diets containing graded additions of supplementary Se

	Treatment					
	Con	SY-L	SY-H	SS-L	s.e.m.	P-value
Start weight (g)	523.4	533.1	538.8	515.1	8.2	0.182
Finish weight (g)	878.1	883.7	938.6	886.9	17.6	0.055
Average daily feed intake (g/day)	52.3	49.9	45.4	51.6	5.3	0.959
Total weight gain (g)	342.7	357.5	397.2	346.8	18.0	0.128
Average live weight gain (g/day)	4.1	4.3	4.7	4.1	0.2	0.128
Feed conversion ratio (g intake/g gain)	13.30	11.16	11.82	13.77	1.4	0.513

Se = selenium; Con = control; SY-L = lowest level of selenised-enriched yeast; SY-H = highest level of selenised-enriched yeast; SS-L = lowest level of sodium selenite.

Table 3 Se content of whole blood, plasma and whole blood GSH-Px activities of female Ring-necked Pheasants, following 42 (T_{42}) and 91 (T_{91}) days exposure to diets containing graded additions of supplementary Se

	Treatment					Contrasts		
	Con	SY-L	SY-H	SS-L	s.e.m.	Con v. Se supplemented	SY-L v. SS-L	SY-L v. SY-H
T_{42} whole blood Se (ng/g FW)	200.8 ^x	246.5 ^{yz}	273.7 ^z	220.9 ^{xy}	9.2	<0.001	0.206	0.171
T_{91} whole blood Se (ng/g FW)	192.9 ^x	276.7 ^{yz}	299.0 ^z	221.6 ^{xy}	11.4	<0.001	0.005	0.517
T_{91} plasma Se (ng/g FW)	78.0 ^x	98.0 ^{yz}	103.8 ^z	82.1 ^{xy}	4.4	0.004	0.059	0.805
T_{91} GSH-Px (U/ml)	60.6 ^x	59.6 ^x	72.4 ^y	68.5 ^{xy}	3.1	0.105	0.186	0.027

Se = selenium; GSH-Px = glutathione peroxidase; Con = control; SY-L = lowest level of selenised-enriched yeast; SY-H = highest level of selenised-enriched yeast; SS-L = lowest level of sodium selenite; FW = fresh weight.

^{xyz}Different superscripts within a row differ significantly ($P < 0.05$).

Table 4 Total Se contents (mg/kg dry matter) of postmortem tissues of female Ring-necked Pheasants offered diets containing graded additions of supplementary Se for either for 42 (T_{42}) or 91 (T_{91}) days

	Treatment					Contrasts		
	Con	SY-L	SY-H	SS-L	s.e.m.	Con v. Se supplemented	SY-L v. SS-L	SY-L v. SY-H
<i>M. Pectoralis Major</i>								
T_{42}	0.64 ^x	1.05 ^y	1.27 ^z	0.65 ^x	0.03	0.063	<0.001	0.004
T_{91}	0.55 ^x	1.00 ^y	1.26 ^z	0.64 ^x	0.02	0.028	<0.001	<0.001
<i>M. Gastrocnemius</i>								
T_{42}	0.71 ^x	1.05 ^y	1.41 ^z	0.75 ^x	0.04	0.066	0.002	<0.001
T_{91}	0.73 ^x	1.23 ^y	1.50 ^z	0.81 ^x	0.04	0.033	0.006	<0.001
<i>M. Peroneus Longus</i>								
T_{42}	0.71 ^x	0.96 ^{xy}	1.24 ^y	0.68 ^x	0.09	0.180	0.231	0.247
T_{91}	0.73 ^x	1.05 ^{yz}	1.37 ^z	0.86 ^{xy}	0.05	0.028	0.004	0.072
Heart								
T_{42}	1.59	2.25	2.16	1.82	0.17	0.045	0.350	0.980
T_{91}	1.70 ^x	2.33 ^y	2.37 ^y	1.78 ^{xy}	0.14	0.067	0.081	0.997
Liver								
T_{42}	2.40 ^{xy}	2.68 ^{yz}	2.86 ^z	2.31 ^x	0.08	0.231	0.035	0.390
T_{91}	2.48 ^x	3.03 ^{yz}	3.16 ^z	2.63 ^{xy}	0.09	0.022	0.063	0.787
Kidney								
T_{42}	3.96 ^x	4.20 ^{xy}	4.88 ^y	4.05 ^{xy}	0.20	0.207	0.948	0.163
T_{91}	4.05	4.46	4.91	4.23	0.19	0.098	0.838	0.393
Gizzard								
T_{42}	1.18 ^x	1.57 ^y	1.77 ^y	1.20 ^x	0.05	0.061	0.002	0.062
T_{91}	1.04 ^x	1.11 ^x	1.48 ^y	1.06 ^x	0.05	0.197	0.913	0.002

Se = selenium; Con = control; SY-L = lowest level of selenised-enriched yeast; SY-H = highest level of selenised-enriched yeast; SS-L = lowest level of sodium selenite.

^{xyz}Different superscripts within a row differ significantly ($P < 0.05$).

breast and leg tissue of SY-L birds were generally greater at both T_{42} and T_{91} when compared with Con and SS-L birds at the same time points ($P < 0.05$) and greater in treatment SY-H when compared with SY-L ($P < 0.05$), the exceptions to this being in the PL at T_{42} .

There were effects in the total Se contents of heart and liver to the graded addition of Se to the diet, differences being more pronounced at T_{91} than at T_{42} . The total Se contents of heart tissue from Con and SS-L birds at T_{42} and T_{91} did not differ appreciably from those recorded in the baseline group at T_0 . Conversely, total Se values were greater in treatments SY-L and SY-H ($P < 0.05$) at both T_{42} and T_{91} than those of Con and SS-L birds at the same time points. The total Se contents of liver tissue in Con and SS-L birds at T_{42} were similar to those of T_0 and did not change appreciably by T_{91} , whereas those of SY-L and SY-H were higher at T_{42} and T_{91} when compared with T_0 samples ($P < 0.05$).

Kidney tissue total Se contents were generally higher in those birds that had received some degree of Se supplementation ($P < 0.05$) and only numerically higher in those birds that had received the highest level of supplementation (SY-H) when compared with the lowest levels (SY-L and SS-L). There were no discernable differences between SS and SY-supplemented birds at a comparable dose (SY-L v. SS-L). The total Se content of gizzard tissue at T_{42} was greatest in those birds that had received SY supplements ($P < 0.05$), irrespective of dose, with there being no difference between Con and SS-L birds, which had values similar to those at T_0 .

At T_{91} , total Se values of gizzard tissue had appeared to decline slightly in all treatments, with the greatest Se concentration being in the gizzards of birds receiving the highest level of Se supplementation ($P < 0.05$). However, this decline may reflect the fact that the slaughter groups at T_{42} and T_{91} comprised different birds.

SeCys was the predominant selenised AA ($P < 0.05$) within whole blood samples at all time points irrespective of treatment (Figure 1) with values being similar, regardless of treatment or time point. This is reflected in erythrocyte GSH-Px activities, which were generally similar between treatments, although values were marginally higher in SY-H when compared with SY-L and Con birds ($P < 0.05$). SeMet although comprising a smaller proportion of total Se, was greater at both T_{42} and T_{91} in SY-L and SY-H birds when compared with Con birds ($P < 0.05$), but similar between SY-L, SY-H and SS-L birds.

SeCys was the predominant selenised AA in liver and kidney tissue, irrespective of treatment, accounting for ~75% of total Se (Figure 2). SeMet comprised a much smaller fraction of total Se in liver and kidney tissue and was not different between treatments. There were no appreciable differences between treatments in the selenised AA contents of breast and thigh tissue, although SeMet values were numerically higher than SeCys values in the breast tissue of SY-supplemented birds ($P < 0.1$) when compared with selenised AA in breast tissue of all birds, irrespective of treatment, although values were markedly higher in the breast tissue of birds that had received diets containing SY supplements ($P < 0.05$). Conversely, in leg

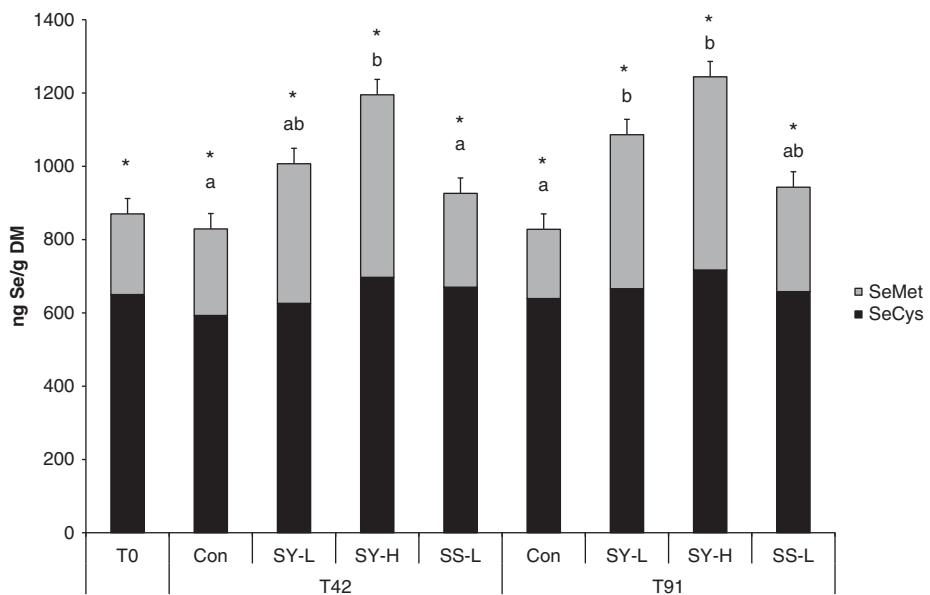


Figure 1 Total Se and the proportion of total Se comprised as selenomethionine (SeMet) or selenocysteine (SeCys) in the whole blood of female Ring-necked Pheasants at day 0 (T_0), 42 (T_{42}) and 91 (T_{91}). Different lowercase letters within time point denote SeMet differences ($P < 0.05$) between bars. Asterisks denote differences between SeMet and SeCys within bars. DM = dry matter.

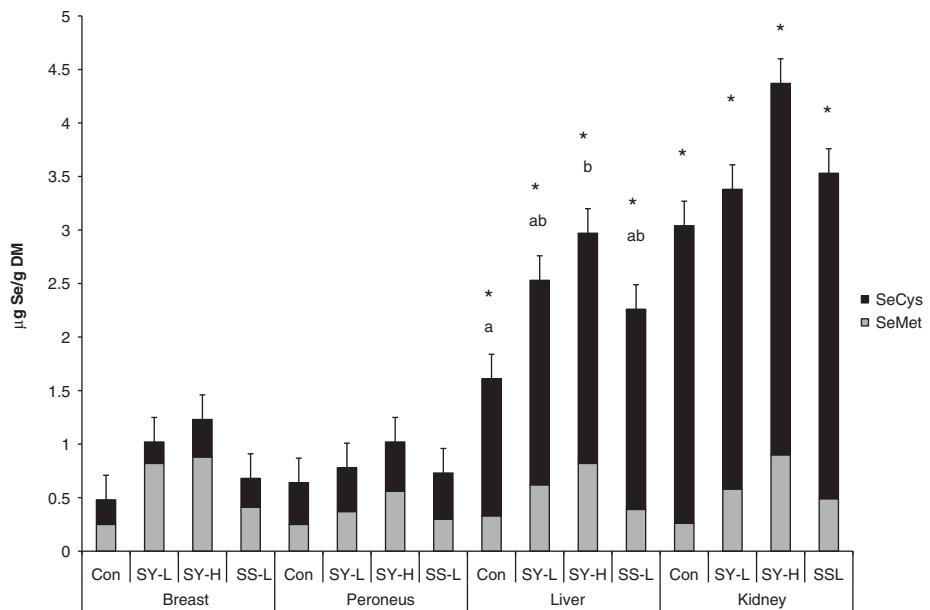


Figure 2 Total selenium (Se) and the proportion of total Se comprised as selenomethionine (SeMet) or selenocysteine (SeCys) in the tissues of female Ring-necked Pheasants following 91 days of dietary treatment. Different lowercase letters within tissue denote SeCys differences ($P < 0.05$) between bars. Asterisks denote differences between SeMet and SeCys within bars. Con = control; SY-L = lowest level of selenised-enriched yeast; SY-H = highest level of selenised-enriched yeast; SS-L = lowest level of sodium selenite.

tissue, SeMet was the predominant form of Se in those birds that had received diets supplemented with SY and SeCys the predominant form in those birds that had received diets that had not been augmented with additional Se (Con) or had been supplemented with selenite (SS-L). However, absolute SeCys values were similar in leg tissue irrespective of treatment indicating that increases in total Se in the leg tissue of SY-supplemented birds was predominantly the result of increases in the SeMet content of the tissue.

There were no effects of treatment on breast muscle TBARS or tissue GSH-Px activities at both 0 and 5 days postmortem (Table 5).

Discussion

In general, there were no major differences between the different treatments in nutritive composition of the experimental diets as would be expected, given that all compositional

Table 5 TBARS and tissue GSH-Px activity in the breast tissue of female Ring-necked Pheasants offered diets containing graded additions of supplementary Se for 91 days

	Treatment					Contrasts		
	Con	SY-L	SY-H	SS-L	s.e.m.	Con v. Se supplemented	SY-L v. SS-L	SY-L v. SY-H
TBARS (mg MDA/kg)	1.23	1.18	0.96	1.04	0.12	0.214	0.826	0.537
GSH-Px 0 days postmortem (U/mg protein)	19.55	19.19	18.69	20.83	1.40	0.992	0.995	0.839
GSH-Px 5 days postmortem (U/mg protein)	20.08	19.03	19.02	20.66	1.33	0.738	0.826	0.999

TBARS = thiobarbituric acid reactive substances; GSH-Px = glutathione peroxidase; Se = selenium; Con = control; SY-L = lowest level of selenised-enriched yeast; SY-H = highest level of selenised-enriched yeast; SS-L = lowest level of sodium selenite; MDA = malondialdehyde.

ingredients and mineral mixes, with the exception of Se, were derived from the same source.

The lack of effect of any significant response over time in whole blood and plasma Se concentrations of Con birds would be expected and is consistent with the findings of other authors (Echevarria *et al.*, 1988; Pan *et al.*, 2007; Yoon *et al.*, 2007; Juniper *et al.*, 2011). However, the Se values recorded in the T₀ group and Con birds were higher than those recorded in commercial lines of poultry in other studies that had received diets, which had not been augmented with supplementary Se (Petrović *et al.*, 2006; Yoon *et al.*, 2007 and Juniper *et al.*, 2011). The differences seen between comparable doses of SY and SS are indicative of improved uptake and incorporation of Se derived from SY and are consistent with the findings in sheep (Van Ryssen *et al.*, 1989; Juniper *et al.*, 2008a), cattle (Gunter *et al.*, 2003; Phipps *et al.*, 2008; Juniper *et al.*, 2008b), pigs (Mahan and Parrett, 1996; Mahan *et al.*, 1999; Kim and Mahan, 2001) and poultry (Petrović *et al.*, 2006; Juniper *et al.*, 2011). However, the degree of response to supplementary Se is markedly lower than those recorded in other studies and most probably reflect the much higher blood and plasma Se levels seen at T₀ and in Con pheasants. This may be indicative of a more efficient Se retention system in wild type classes of bird when consuming diets low in Se. However, as dietary Se levels increase this efficient scavenging system may become quickly saturated, hence why responses in pheasants are not as pronounced as those seen in commercial lines of poultry (Petrović *et al.*, 2006; Juniper *et al.*, 2011). Similar observations have been reported in different breeds of pig, whereby Se uptake and metabolism of a commercial hybrid was different to that of a pure breed when fed identical diets (Banoch *et al.*, 2009).

The hierarchy of Se concentrations observed in different tissues tends to reflect the relative metabolic activity of each tissue. Glandular visceral tissues tend to have greater total Se concentrations than muscle tissue and cardiac muscle has a greater Se content than skeletal tissue or gizzard tissue. These data agree with those reported in laying hens (Petrović *et al.*, 2006; Pan *et al.*, 2007), broilers (Wang and Xu, 2008), turkeys (Juniper *et al.*, 2011) and are also similar to those reported in lambs (Juniper *et al.*, 2008a) and beef cattle (Juniper *et al.*, 2008b).

The differences seen in cardiac, skeletal and gizzard muscle tissue total Se concentrations, when comparing comparable doses of SS and SY, are indicative of improved uptake and assimilation of Se derived from SY. However, these responses in muscle tissue lack the same degree of response reported in other studies on commercial lines of poultry (Petrović *et al.*, 2006; Juniper *et al.*, 2011) reflecting the responses seen in blood and plasma within this study. Furthermore, muscle tissue Se concentrations at T₀ and in Con birds were greater than those reported in commercial lines of poultry receiving unsupplemented diets (Petrović *et al.*, 2006; Juniper *et al.*, 2011). This may further indicate that wild type birds, such as pheasants, may have more efficient means of Se capture than those of commercial lines of poultry at low dietary levels of Se and that this system becomes very quickly saturated as dietary Se levels increase.

SeCys was the predominant selenised AA within blood, irrespective of treatment or time point (Figure 1). This observation in pheasants is unlike those reported for turkeys (Juniper *et al.*, 2011) in which SeMet was the predominant selenised AA, but similar to those reported for lambs (Juniper *et al.*, 2008a), beef cattle (Juniper *et al.*, 2008b) and dairy cattle (Phipps *et al.*, 2008) where SeCys was the predominant form.

SeCys was the predominant selenised AA in liver and kidney tissue, irrespective of treatment (Figure 2). The predominance of SeCys in liver and kidney tissue has been reported in turkeys (Juniper *et al.*, 2011), as well as in lambs (Juniper *et al.*, 2008a) and beef cattle (Juniper *et al.*, 2008b), and may reflect the greater metabolic activity of these tissues. Much of the difference in total Se content of skeletal muscle with SY-supplemented diets is attributable to increases in SeMet content rather than changes in SeCys content, as SeCys content is comparable between treatments. Furthermore, the lack of any difference in SeCys content of skeletal muscle is reflected in the lack of any difference in tissue GSH-Px activity.

The predominance of SeMet in skeletal muscle in SY-supplemented treatments is similar to observations made in the breast tissue of turkeys (Juniper *et al.*, 2011) and the skeletal tissue of beef cattle (Juniper *et al.*, 2008b), although the specific SY dose response that was reported in these studies is not apparent in this study.

The lack of any effect of treatment on breast tissue TBARS and GSH-Px activity in poultry is not uncommon.

Radmilla *et al.* (2008) and Sevcikova *et al.* (2006) have both reported an absence of effect in broilers and a similar lack of response has also been reported in turkeys (Juniper *et al.*, 2011). In addition, both pigs (Mahan *et al.*, 1999) and cattle (Skřivanová *et al.*, 2007; Taylor *et al.*, 2008; Juniper *et al.*, 2008b) have shown a lack of response in both tissue TBARS and GSH-Px activity in skeletal muscle to additional dietary Se. These results would suggest that higher tissue total Se contents tend not to reflect improvements in GSH-Px activity to the oxidative stability of skeletal tissue meat keeping quality.

Conclusion

The incorporation of graded additions of SY into the diets of growing pheasants increased total Se in tissues and blood in a dose-dependent manner. The high levels of total Se seen in T₀ birds at the start of the study, and in control birds throughout the study, maybe indicative of a more efficient Se capture system in wild type birds. Furthermore, the limited responses seen in both blood and tissue of both control and SS-L birds would also indicate that this efficient Se capture system may become very quickly saturated, as dietary Se concentrations increase. However, the moderate increases in the Se content of tissues of SY-supplemented birds appear to be the result of increases in the proportion of total Se comprised as SeMet, reflecting improved uptake and incorporation of SeMet.

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References

- Anderson PH, Berrett S and Patterson DSP 1978. Glutathione peroxidase activity in erythrocytes and muscle of cattle and sheep and its relationship to selenium. *Journal of Comparative Pathology* 88, 181–189.
- Banoch T, Fajt Z, Odehnalova S, Drabek J and Svoboda M 2009. Evaluation of selenium status in pure bred Duroc sows and their progeny. *Neuroendocrinology Letters* 30 (Suppl), 143–147.
- Bierla K, Vacchina V, Szpunar J, Bertin G and Lobinski R 2008a. Simultaneous derivatization of selenocysteine and selenomethionine in animal blood prior to their specific determination by 2D size exclusion ion-pairing reversed phase HPLC-ICP MS. *Journal of Analytical Atomic Spectrometry* 23, 508–513.
- Bierla K, Vacchina V, Szpunar J, Bertin G and Lobinski R 2008b. Determination of selenocysteine and selenomethionine in edible animal tissues by 2D size-exclusion reversed-phase HPLC-ICP MS following carbamidomethylation and proteolytic extraction. *Analytical and Bioanalytical Chemistry* 390, 1789–1798.
- Burke RF, Hill KE and Xia Y 2005. Selenoproteins and the human selenium requirement. 12th International Symposium on Trace Elements in Man and Animals (TEMA), 19–23 June 2005, Coleraine, N. Ireland.
- Daun C, Johansson M, Önning G and Åkesson B 2000. Glutathione peroxidase activity, tissue and soluble selenium content in beef and pork in relation to meat ageing and pig RN phenotype. *Food Chemistry* 73, 313–319.
- DeVore VR and Greene BE 1982. Glutathione peroxidase in post-rigor bovine semitendinosus muscle. *Journal of Food Science* 47, 1406–1409.
- DeVore VR, Colnago GL, Jensen LS and Greene BE 1983. Thiobarbituric acid values and glutathione peroxidase activity in meat from chickens fed a selenium supplemented diet. *Journal of Food Science* 48, 300–301.
- Echevarria MG, Henry CB, Ammerman PV and Miles RD 1988. Estimation of the relative bioavailability of inorganic selenium sources for poultry. 1. Effect of time and high dietary selenium on tissue selenium uptake. *Poultry Science* 67, 1295–1301.
- Gunter SA, Beck PA and Phillips JM 2003. Effects of supplementary selenium source on the performance of blood measurements in beef cows and their calves. *Journal of Animal Science* 81, 856–864.
- Home Office 1986. Animal Scientific Procedures Act 1986. Her Majesty's Stationery Office, London.
- Juniper DT, Phipps RH and Bertin G 2011. Effect of dietary supplementation with selenium-enriched yeast or sodium selenite on selenium tissue distribution and meat quality in commercial-line turkeys. *Animal* 5, 1751–1760.
- Juniper DT, Phipps RH, Ramos-Morales E and Bertin G 2008a. Effects of dietary supplementation with selenium enriched yeast or sodium selenite on selenium tissue distribution and meat quality in lambs. *Animal Feed Science and Technology* 149, 228–239.
- Juniper DT, Phipps RH, Ramos-Morales E and Bertin G 2008b. Effect of dietary supplementation with selenium enriched yeast or sodium selenite on selenium tissue distribution and meat quality in beef cattle. *Journal of Animal Science* 86, 3100–3109.
- Kim YY and Mahan DC 2001. Effect of dietary selenium source, level, and pig hair colour on various selenium indices. *Journal of Animal Science* 79, 949–955.
- Mahan DC and Parrett NA 1996. Evaluating the efficacy of selenium-enriched yeast and sodium selenite on tissue selenium retention and serum glutathione peroxidase activity in grower and finisher swine. *Journal of Animal Science* 74, 2967–2974.
- Mahan DC, Cline TR and Richert B 1999. Effects of dietary levels of selenium-enriched yeast and sodium selenite as selenium sources fed to growing-finishing pigs on performance, tissue selenium, serum glutathione peroxidase activity, carcass characteristics, and loin quality. *Journal of Animal Science* 77, 2172–2179.
- Mester Z, Willie S, Sturgeon R, Caruso JA, Fernandez M, Fodor P, Goldschmidt RJ, Goenaga-Infante H, Lobinski R and Wolf WR 2006. Certification of a new selenized yeast reference material (selm-1) for methionine, selenomethionine and total selenium content and its use in an intercomparison exercise for quantifying of these analytes. *Analytical and Bioanalytical Chemistry* 385, 168–180.
- National Research Council (NRC) 1994. Nutrient Requirements of Poultry, 9th edition. The National Academies Press, Washington, DC.
- Pan C, Huang K, Zhao Y, Qin S, Chen F and Hu Q 2007. Effect of selenium source and level in hen's diet on tissue selenium deposition and egg selenium concentrations. *Journal of Agricultural Food Chemistry* 55, 1027–1032.
- Paglia DE and Valentine WN 1967. Studies on the quantitative and qualitative characterization of erythrocyte glutathione peroxidase. *Journal of Laboratory and Clinical Medicine* 70, 158–168.
- Petrović V, Boldižárová K, Faix S, Mellen M, Arpašová H and Leng L 2006. Antioxidant and selenium status of laying hens fed with diets supplemented with selenite or Se-yeast. *Journal of Animal Feed Science* 15, 435–444.
- Phipps RH, Grandison AS, Jones AK, Juniper DT, Ramos-Morales E and Bertin G 2008. Selenium supplementation of lactating dairy cows: effects on milk production an total selenium content and speciation in blood, milk and cheese. *Animal* 2, 1610–1618.
- Radmilla M, Jovanovic BI, Baltic ZM, Sefer D, Petrujkic B and Sinovec Z 2008. Effects of selenium supplementation as sodium selenite or selenized yeast and different amounts of vitamin E on selenium and vitamin E status in broilers. *Acta Veterinaria-Beograd* 58, 369–380.
- Rotruck JT, Pope AL, Ganther HE, Swanson AB, Hafeman DG and Hoekstra WG 1973. Selenium: biochemical role as a component of glutathione peroxidase. *Science* 179, 588–590.
- Sevcikova S, Skrivan M, Dlouha G and Koucký M 2006. The effect of selenium source on the performance and meat quality of broiler chickens. *Czech Journal of Animal Science* 51, 449–457.
- Skřivanová E, Marounek M, De Smet S and Raes K 2007. Influence of dietary selenium and vitamin E on quality of veal. *Meat Science* 76, 495–500.
- Surai PF 2006. Selenium absorption and metabolism. In *Selenium nutrition and health* (ed. PF Surai), pp. 161–171. Nottingham University Press, Nottingham, UK.

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- Tarladgis BG, Watts BM and Younathan MT 1960. A distillation method for the quantitative determination of malonaldehyde in rancid foods. *Journal of the American Chemical Society* 37, 44–48.
- Taylor JB, Marchello MJ, Finley JW, Neville TL, Combs GF and Caton JS 2008. Nutritive value and display life attributes of selenium enriched beef muscle foods. *Journal of Food Composition and Analysis* 21, 183–186.
- Underwood EJ and Suttle NF 2001. The mineral nutrition of livestock, 3rd edition. CAB International, Wallingford, UK.
- Van Ryssen JBJ, Deagen JT, Beilstein MA and Whanger PD 1989. Comparative metabolism of organic and inorganic selenium by sheep. *Journal of Agricultural Food Chemistry* 37, 1358–1363.
- Wang YB and Xu BH 2008. Effect of different selenium source (sodium selenite and selenium yeast) on broiler chickens. *Animal Feed Science and Technology* 144, 306–314.
- Yoon I, Werner TM and Butler JM 2007. Effect of source and concentration of selenium on growth performance and selenium retention in broiler chickens. *Poultry Science* 86, 727–730.