

# *Effects of coastal managed retreat on mercury biogeochemistry*

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Accepted Version

Sizmur, T. ORCID: <https://orcid.org/0000-0001-9835-7195>,  
Godfrey, A. and O'Driscoll, N. J. (2016) Effects of coastal  
managed retreat on mercury biogeochemistry. *Environmental  
Pollution*, 209. pp. 99-106. ISSN 0269-7491 doi:  
10.1016/j.envpol.2015.11.016 Available at  
<https://centaur.reading.ac.uk/48003/>

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To link to this article DOI: <http://dx.doi.org/10.1016/j.envpol.2015.11.016>

Publisher: Elsevier

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Effects of coastal managed retreat on mercury biogeochemistry

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## 14    **Abstract**

15    We investigated the impact of managed retreat on mercury (Hg) biogeochemistry at a site subject  
16    to diffuse contamination with Hg. We collected sediment cores from an area of land behind a  
17    dyke one year before and one year after it was intentionally breached. These sediments were  
18    compared to those of an adjacent mudflat and a salt marsh. The concentration of total mercury  
19    (THg) in the sediment doubled after the dyke was breached due to the deposition of fresh  
20    sediment that had a smaller particle size, and higher pH. The concentration of methylmercury  
21    (MeHg) was 27% lower in the sediments after the dyke was breached. We conclude that coastal  
22    flooding during managed retreat of coastal flood defences at this site has not increased the risk of  
23    Hg methylation or bioavailability during the first year. As the sediment becomes vegetated,  
24    increased activity of Hg-methylating bacteria may accelerate Hg-methylation rate.

25    **Keywords:** Mercury, Methylmercury, Biogeochemistry, Sediment deposition, Coastal

26    **Capsule:** Mercury concentration doubled in sediments after coastal flooding but methylmercury  
27    concentration decreased

## Introduction

Coastal wetlands have been subject to dramatic global declines in the past due to dyking and draining for agriculture. However, this practice is now being reversed in many countries because salt marshes are valued as habitats for wildlife and as natural defence against rising sea-levels (Singh et al., 2007). Managed retreat of coastal defences has led to an increase in the number of sites where dykes are breached, agricultural fields are inundated with seawater, sediment is deposited over soils, and new salt marshes are created. Inundation of previously dyked farmland leads to considerable biogeochemical changes, characterised by increased salinity, lower redox potential (Portnoy and Giblin, 1997) and a decaying mat of buried vegetation (Emmerson et al., 2000). There is concern that biogeochemical changes during managed retreat may alter the fate of redox-sensitive contaminants such as mercury (Hg) (Morris et al., 2014).

The Bay of Fundy in Southeastern Canada is renowned for having the largest tidal amplitude in the world, which gives rise to expansive intertidal mudflats and vast areas of salt marsh (Crowell et al., 2011; Desplanque and Mossman, 2004). For centuries the Bay's coastline has been extensively dyked to use the land for agriculture (Wynn, 1979). The land surrounding the Bay of Fundy is designated a 'biological mercury hotspot' due to elevated concentrations of Hg in biota (Evers et al., 2007). The Bay of Fundy itself has been identified as an area of special concern for Hg contamination because the Bay's ecosystem may be critical to concentrations of Hg found in fish, birds and wildlife (Hung and Chmura, 2006).

Mercury enters the Bay of Fundy through seawater inflow and atmospheric deposition (Sunderland et al., 2012). The Hg present in sediments of the Bay of Fundy is strongly associated with organic matter and fine textured sediments (O'Driscoll et al., 2011; Sizmur et al., 2013b). Inorganic Hg in sediments can be converted to methylmercury (MeHg) under anoxic conditions by sulphate-reducing bacteria (Compeau and Bartha, 1985). Methylmercury can biomagnify through food webs (Lavoie et al., 2010) and is a potent neurotoxin affecting higher trophic level animals and humans (Rasmussen et al., 2005).

Increases in MeHg concentrations in sediments and biota have been observed during the decades that follow terrestrial freshwater flooding for dam construction or wetland creation (Kelly et al., 1997; Sinclair et al., 2012). However, little research has been done to assess changes in Hg biogeochemistry after coastal wetland flooding. Terrestrial flooding events, like reservoir or wetland creation, entail a permanent change in sediment redox from oxic to anoxic conditions because the sediments are constantly flooded. However, coastal flooding events subject the land to fluctuating oxic/anoxic conditions due to the tidal cycle. These fluctuations generate an oxic-anoxic interface in the sediment. The temporal fluctuations in redox conditions increases the volume of sediment where sulphate reduction and mercury methylation may occur (Heim et al., 2007; Sizmur et al., 2013a). However, there is also frequent tidal flushing of inundated areas which acts as a significant means of removing MeHg from the surface of coastal sediments (Guédron et al., 2012). Therefore, it is not clear if managed retreat will increase or decrease Hg and MeHg concentrations in sediments.

We investigated the effects of managed retreat on mercury biogeochemistry at Beaubassin Research Station where a dyke has recently been breached, allowing the seawater to inundate land previously drained for agriculture.

## **Materials and Methods**

### Site Description

Beaubassin Research Station (Latitude: 45.852195 Longitude: -64.279631) is located on the Chignecto Isthmus between Nova Scotia and New Brunswick, Canada (Figure 1a). It lies along the Cumberland Basin, a branch of Chignecto Bay, in the Bay of Fundy which is sourced from the Gulf of Maine. The average tidal amplitude at Beaubassin is 11 m (Gordon Jr and Baretta, 1982). Recently, an eroding 150-year-old dyke was replaced with a new dyke built approximately 90 m back from the pre-existing coastline in order to protect transport infrastructure and the historic site of Fort Beausejour from tidal surges. The 40 ha of low lying land between the old dyke and the new dyke (Latitude: 45.851595 Longitude: -64.294379) was flooded in October 2010. Flooding occurred when the old dyke was deliberately breached so that sediment could accumulate to protect the new dyke before the old dyke completely failed (Ollerhead et al., 2011). Tidal re-entry has resulted in the rapid deposition of fresh sediment over the top of the agricultural soil, burying a mat of terrestrial vegetation. At the time of sampling, new salt marsh vegetation was yet to establish.

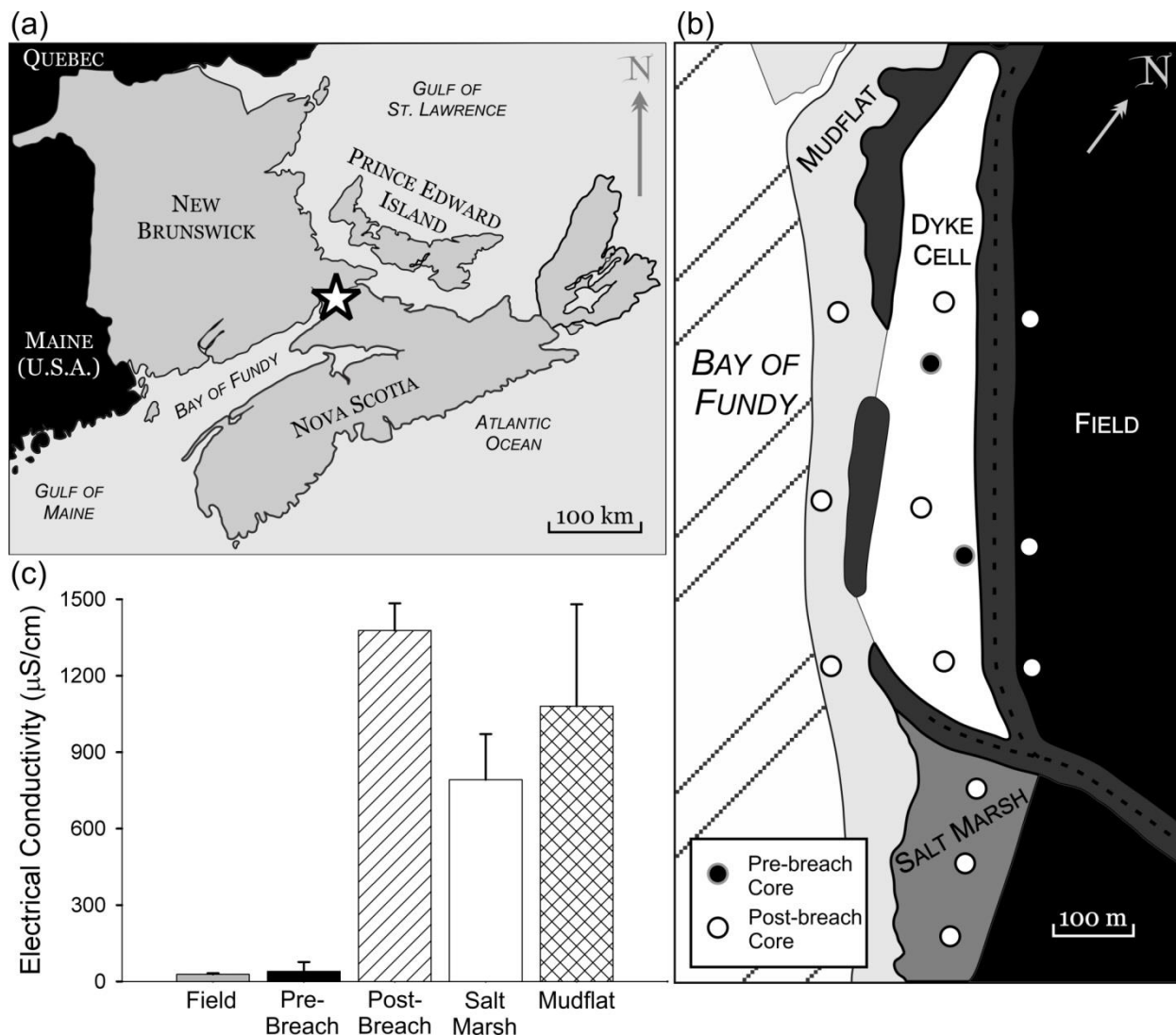


Figure 1. (a) Site location at Beaubassin, New Brunswick, Canada; (b) Location of all cores sampled from the dyke cell (pre-breach and post-breach) along with adjacent sites (mudflat, salt marsh and field). The location of two gaps in the wall of the dyke cell represent where they were deliberately breached in 2010; (c) Electrical conductivity of sediment cores sampled (averaged 0-15 cm) shown here to demonstrate the influence of seawater on the dyke cell pre-breach and post-breach.



98 Sample Collection and Preparation

99 Two 16 cm deep cores were taken in the dyke cell (Figure 1b) between the new and the old  
100 dykes (hereafter referred to as the pre-breach cores) in summer 2009 (before the old dyke was  
101 breached in 2010). We returned to the site in summer 2011 to collect cores one year after the old  
102 dyke was breached. Three 15 cm deep cores were sampled at four locations: (i) The area  
103 previously sampled in the dyke cell between the new and old dykes (hereafter referred to as the  
104 post-breach cores), (ii) the mudflat seaward of the dyke cell, (iii) a pre-existing salt marsh  
105 adjacent to the dyke cell, and (iv) the field landward of the dyke cell (Figure 1b). All cores were  
106 sampled at low tide using polyvinyl chloride (PVC) cores (10 cm internal diameter) that were  
107 dug out with a stainless steel spade.

108

109 Pre-breach cores were sliced in 2 cm intervals to a depth of 16 cm, producing a total of eight  
110 slices per core. Each of the post-breach, mudflat, salt marsh, and field cores were sliced at 1 cm  
111 intervals for the upper 5 cm of sediment and then at 2 cm intervals for the remaining 10 cm,  
112 producing a total of 10 core slices per core. Core slices were individually sealed in Ziploc bags at  
113 the research station and placed in a dark cooler with ice packs for transport back to the  
114 laboratory.

115

116 At the laboratory each sediment slice was thoroughly homogenised by hand in the Ziploc bag  
117 and frozen as a wet homogenate at -20 °C. Sediment samples were later thawed and a subsample  
118 dried at 60 °C for 24 hours. Dried sediment samples were ground with a pestle and mortar and  
119 sieved to < 2 mm. A subsample of wet sediment was analysed for electrical conductivity (EC)

using a VWR Symphony SP90M5 meter and Orion electrical conductivity probe. The field was only sampled to demonstrate that the pre-breach sediments had not been inundated by seawater prior to the breach. Since the EC of the pre-breach and field cores (Figure 1c) revealed no significant difference ( $p > 0.05$ ), further analysis of the field cores was deemed unnecessary. Each slice of the remaining cores was analysed for total mercury (THg), MeHg, percentage organic matter (%OM), particle size distribution, water-extractable organic carbon (WEOC) and pH.

#### Analytical Procedures

Total mercury in sediment was determined using thermal degradation – gold amalgamation atomic absorbance spectroscopy as outlined in EPA Method 7473 (1998) using a Nippon MA-2000 non-dispersive double-beam cold-vapor atomic absorption Hg analyzer. Methylmercury was determined in sediments by alkaline digestion, ethylation purge and trap Gas Chromatography - Cold Vapour Atomic Fluorescence Spectrometry (GC-CVAFS) following Sizmur et al (2013b). A 100 mg sample of sediment was digested in 2.5 ml of basic methanol (25 % KOH/MeOH) by shaking on a reciprocal shaker for 1 hour and then heating for 1 hour at 90 °C. Within 24 hours of digestion, a 60 µl aliquot was transferred to a glass reaction bubbler, ethylated with  $\text{NaB}(\text{C}_2\text{H}_5)_4$ , purged with argon, collected on a Tenax trap and analysed for MeHg using GC-AFS (Brooks Rand Model III).

Organic matter in sediment was determined by loss on ignition at 500 °C (Byers et al., 1978) and particle size distribution by the micro-pipette method (Miller and Miller, 1987). Sand was

calculated as particles 2000-63  $\mu\text{m}$ , silt as 63-2  $\mu\text{m}$ , and clay as  $< 2 \mu\text{m}$  in diameter. Water-extractable organic carbon was determined following Sizmur et al (2011) by shaking 1 g of sediment with 40 ml of Milli Q water for 2 hours on a reciprocal shaker at 120 shakes  $\text{min}^{-1}$ , followed by centrifuging at 4000 rpm (2647 G) for 20 min and filtering to  $< 0.45 \mu\text{m}$  with polypropylene membrane filters, before TOC/TIC analysis with a Shimadzu TOC-V CPH Total Organic Carbon Analyzer. Sediment pH was analysed in WEOC vials prior to centrifuging.

#### Quality control

Sediments were analysed in triplicate alongside certified reference materials MESS-3 (National Research Council Canada) and SQC-1238 (Sigma Aldrich RTC) for THg and MeHg respectively. Mean recovery of THg from MESS-3 was 102.2 % (SD = 1.4 %). Mean recovery of MeHg from SQC-1238 was 94.4 % (SD = 12.0 %). Detection limits for MeHg and THg were 0.65 and 1.21  $\text{pmol g}^{-1}$ , respectively. Both samples and reference materials during Hg analysis were corrected for background by subtracting averaged method blanks from the analysed samples.

#### Statistical Analysis

Statistical analysis was carried out using Genstat version 16. Normality and homoscedasticity was assessed by inspecting residual plots. Two-way analysis of variance was carried out on all data (MeHg, THg, pH, clay, %OM, WEOC and EC) using 'site' and 'depth' as the factors and allowing for interactions. Fisher's least significant difference was used to identify differences between individual treatments. Multiple linear regression was carried out by forward selection; first the variable that resulted in the highest  $R^2$  values was included in the model, then variables

that resulted in the greatest increase were added. Data presented in text as average values at each site are calculated from the concentrations in cores averaged across all depths. All the raw data is provided in the supporting information.

## Results

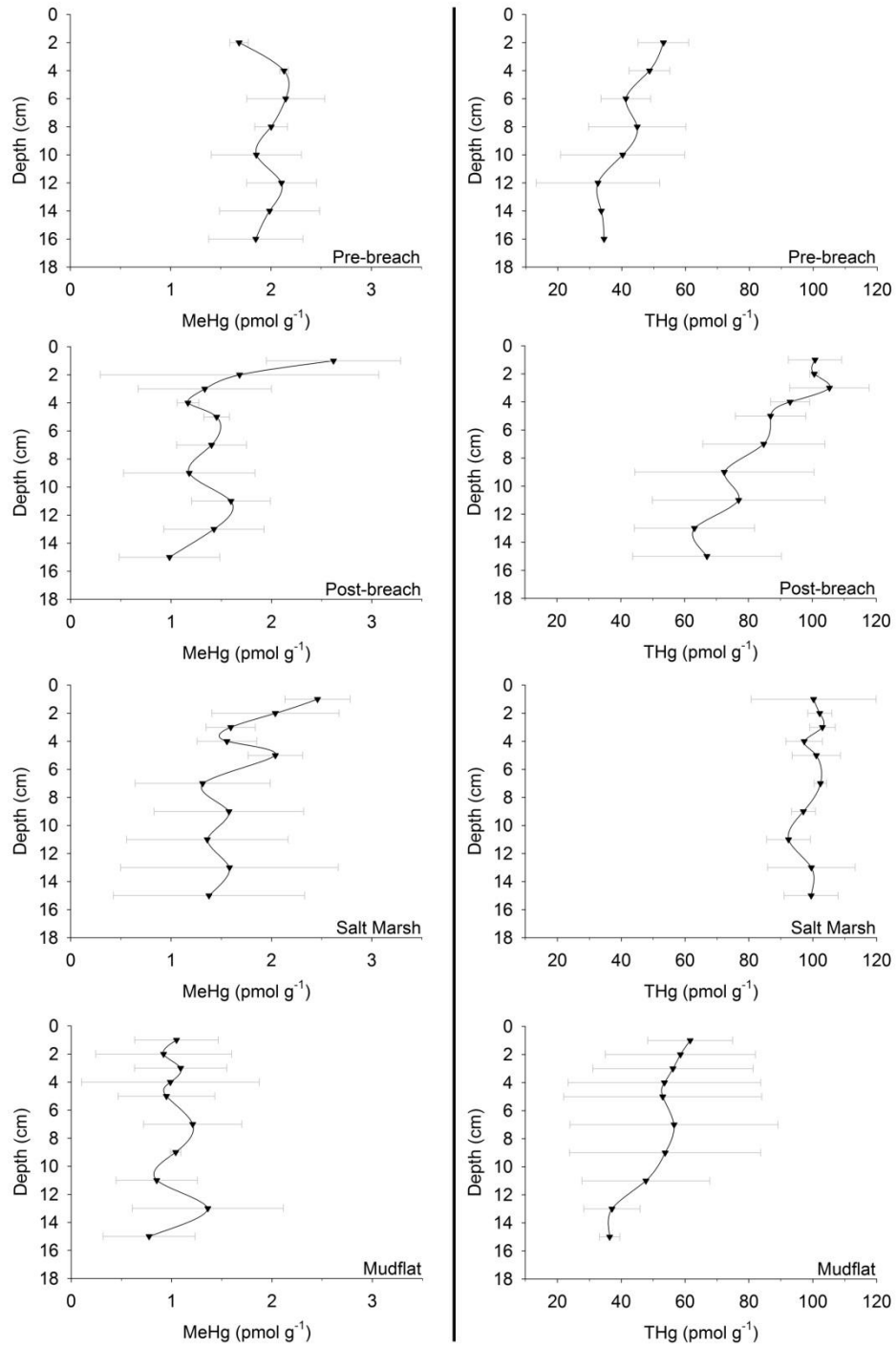
### *Mercury and Methylmercury*

The average concentration of THg in the post-breach cores was 85.1 pmol g<sup>-1</sup> (SD = 15.6) which was approximately double the concentration in the pre-breach cores (41.1 pmol g<sup>-1</sup>, SD = 9.52). THg decreased significantly ( $p < 0.001$ ) with depth (Table 1) in the post-breach and mudflat cores but this decrease was not observed in the pre-breach or the salt marsh cores (Figure 2). The THg concentration in the salt marsh cores was significantly ( $p < 0.05$ ) greater than the mudflat or the dyke cell pre- or post-breach. The post-breach cores had significantly ( $p < 0.05$ ) greater Hg concentrations than the pre-breach cores.

Table 1 Analysis of variance from physiochemical sediment variables; Water Extractable Organic Carbon (WEOC), pH, Electrical Conductivity (EC), Clay content and Organic Matter (OM).

Variable	Site F value	Depth F value	Site-depth interaction F value
THg	62.19***	2.34*	0.41
MeHg	12.83***	2.06*	0.55
% MeHg	31.41***	0.95*	0.62
% OM	15.03***	0.29	0.93
pH	16.87***	0.97*	0.79
% Clay	24.93***	1.74	0.38
WEOC	32.14***	1.68	1.36
EC	69.15***	0.82	1.23

\* =  $p < 0.05$ , \*\*\* =  $p < 0.001$



181

182 Figure 2. Total mercury (THg) and methylmercury (MeHg) concentrations of sediment  
 183 slices of cores sampled from the dyke cell (pre-breach and post-breach) along with  
 184 adjacent sites (mudflat and salt marsh). The error bars represent the standard deviation  
 185 of three replicate cores ( $n = 2$  for the pre-breach cores).

MeHg significantly ( $p < 0.05$ ) decreased with depth (Table 1) in the post-breach and salt marsh cores (Figure 2). Although THg was greater after inundation, MeHg concentration was significantly ( $p < 0.05$ ) lower in post-breach sediments (Figure 2). Methylmercury concentrations were 27% lower in the post-breach cores ( $1.48 \text{ pmol g}^{-1}$ ,  $SD = 0.54$ ) compared to the pre-breach cores ( $1.97 \text{ pmol g}^{-1}$ ,  $SD = 0.31$ ). However, we did measure 36% higher MeHg concentrations in the upper 2 cm of the post-breach sediment than in the top 2 cm of the pre-breach cores (Figure 2). The post-breach MeHg concentration was not significantly ( $p > 0.05$ ) different than that measured in the salt marsh ( $1.69 \text{ pmol g}^{-1}$ ,  $SD = 0.60$ ) but was significantly ( $p > 0.05$ ) greater than MeHg in the mudflat ( $1.02 \text{ pmol g}^{-1}$ ,  $SD = 0.51$ ). The percentage of MeHg as a proportion of the THg (%MeHg) was significantly ( $p < 0.05$ ) greater in the pre-breach cores (5.97%,  $SD = 2.99$ ) than the post-breach cores (2.02%,  $SD = 0.58$ ). %MeHg in the post-breach sediment was not significantly ( $p > 0.05$ ) different from the mudflat (2.34%) or salt marsh (1.84%) sediments.

#### Physiochemical variables

The EC of the pre-breach cores was not significantly different to the samples taken from the field behind the new dyke. EC was significantly increased ( $p < 0.05$ ) by periodic tidal inundation of the dyke cell, increasing over 2000% from pre- to post-breach (Figure 1c). Post-breach sediment EC was also significantly ( $p < 0.05$ ) greater than the salt marsh and mudflat but the magnitude of the difference was much smaller.

Sediment pH was significantly ( $p < 0.001$ ) greater after inundation of the dyke cell, rising from 5.08 ( $SD = 1.2$ ) in the pre-breach cores to 7.43 ( $SD = 0.6$ ) in the post-breach cores (Figure 3).

209 There was no significant ( $p > 0.05$ ) pH difference between the post-breach cores and the salt  
210 marsh or mudflat cores.

211

212 The texture of the sediment in the top 15 cm of the dyke cell significantly ( $p < 0.001$ ) changed  
213 during salt marsh restoration as fresh sediment with a smaller particle size distribution was  
214 deposited over the top of the drained agricultural field (Figure 3). Percentage clay was  
215 significantly ( $p < 0.05$ ) greater and sand significantly ( $p < 0.05$ ) lower in the sediment after the  
216 inundation. Percentage clay in the post-breach cores (29.8%,  $SD = 1.52$ ) was nearly double that  
217 in the pre-breach cores (16.6%,  $SD = 13.8$ ). The proportions of sand, silt and clay in the post-  
218 breach cores were not significantly ( $p > 0.05$ ) different to the sediments sampled from the  
219 mudflat (Figure 3) but clay content was significantly ( $p < 0.05$ ) greater than sediments sampled  
220 from the salt marsh.

221

222 The post-breach sediments had significantly ( $p < 0.05$ ) higher organic matter (%OM) and WEOC  
223 than the pre-breach cores (Table 1 and Figure 3). There was no significant ( $p > 0.05$ ) difference  
224 between the post-breach cores and the salt marsh and mudflat cores, for either %OM or WEOC.  
225 The concentration of both WEOC ( $44.9 \text{ mmol kg}^{-1}$ ,  $SD = 4.64$ ) and %OM (6.3%,  $SD = 0.8$ ) in  
226 the post-breach cores was greater than the mudflat cores ( $32.1 \text{ mmol kg}^{-1}$ ,  $SD = 8.87$  and 5.8%,  
227  $SD = 1.2$ ) but lower than the salt marsh cores ( $47.9 \text{ mmol kg}^{-1}$ ,  $SD = 7.17$  and 6.7%,  $SD = 0.7$ ).

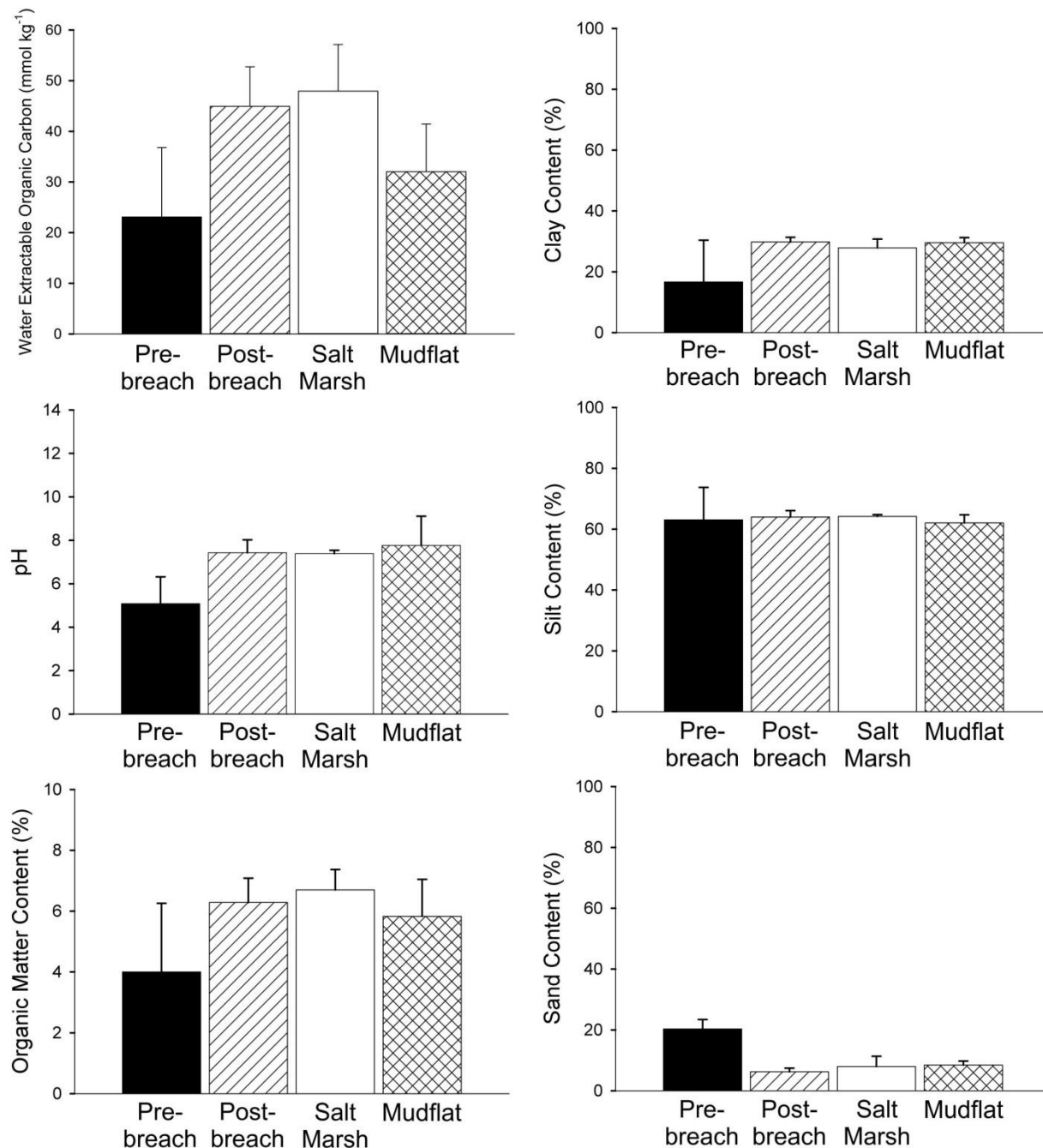


Figure 3. Physiochemical variables measured in cores (averaged 0-15 cm) sampled from the dyke cell (pre-breach and post-breach) along with adjacent sites (mudflat and salt marsh). The error bars represent standard deviation of three replicate cores (n = 2 for the pre-breach cores).



### Multiple linear regressions

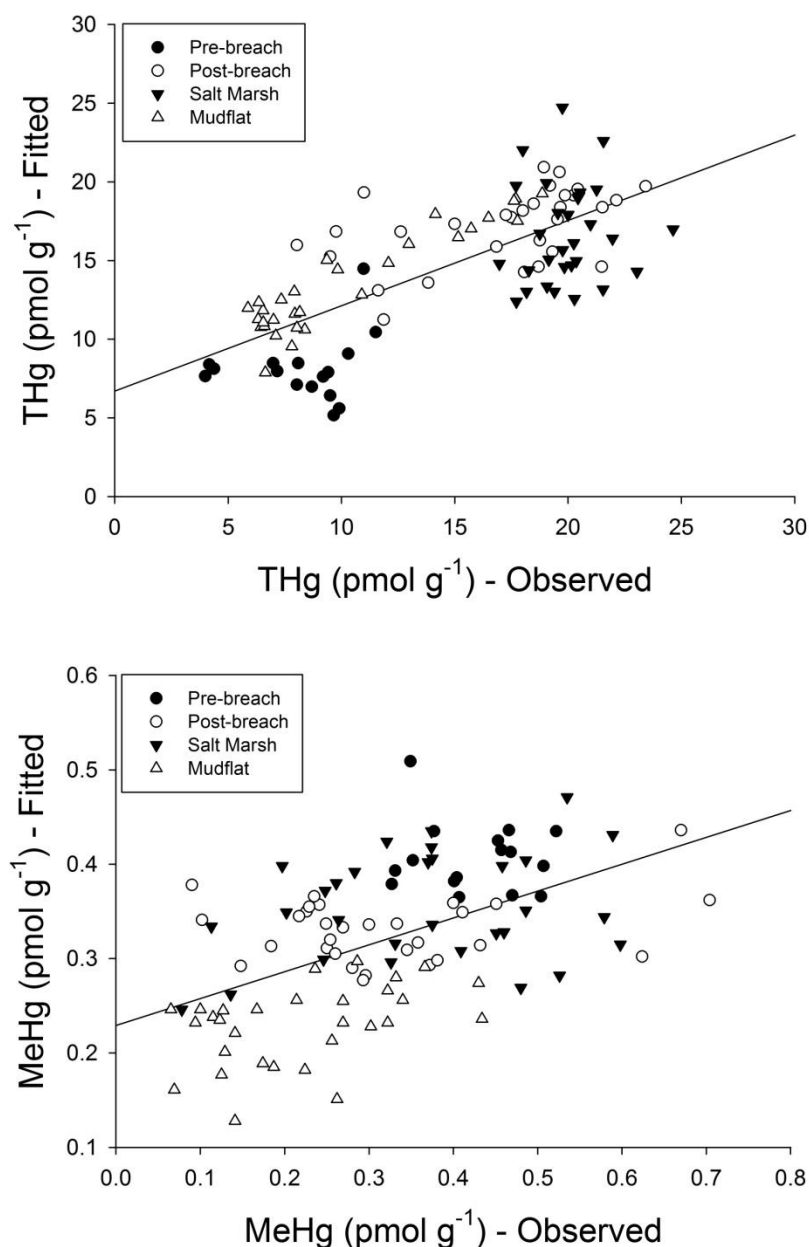
The correlation between the best multiple linear regression model and the THg concentrations measured in the sediments (Figure 4) yielded an  $R^2$  value of 0.524 and a p value  $< 0.001$  (Table 2). The explanatory variables in order of decreasing importance were WEOC, pH, EC and %Clay. Adding the next most important variable (%OM) decreased the  $R^2$  value to 0.519. WEOC alone explained 36.7% of the variation in the observed data.

Table 2 Significance and correlation results of forward multiple linear regression models for the prediction of THg and MeHg from physiochemical sediment variables; Water Extractable Organic Carbon (WEOC), pH, Electrical Conductivity (EC), Clay content and Organic Matter (OM).

Response variable	Fitted terms	F value	$R^2$
THg	WEOC	61.8***	0.367
	WEOC+pH	48.6***	0.476
	WEOC+pH+EC	35.5***	0.496
	WEOC+pH+EC+Clay	29.9***	0.524
MeHg	EC	7.02**	0.54
	EC+THg	8.36***	0.123
	EC+THg+pH	7.07***	0.148
	EC+THg+pH+Clay	6.05***	0.161
	EC+THg+pH+Clay+WEOC	5.50***	0.176
	EC+THg+pH+Clay+WEOC+OM	6.56***	0.241

\*\* =  $p < 0.01$ , \*\*\* =  $p < 0.001$

The variability in MeHg concentrations was more difficult to explain than the THg concentrations using the physiochemical variables measured. The multiple linear regression model for MeHg (Figure 4) had a lower  $R^2$  value than the model for THg. The fit which included EC, THg, pH, %Clay, WEOC and %OM (in order of decreasing importance) had an  $R^2$  value of 0.241 and a p value of  $< 0.001$ . Although EC accounted for the greatest extent of the variability in the MeHg dataset, when considered on its own EC accounted for only 5.4% of the variation. This indicates that variables that we measured could not adequately explain the concentration of MeHg in the sediments.



253

254 Figure 4. Total mercury (THg) and methylmercury (MeHg) concentrations observed in  
 255 cores sampled from the dyke cell (pre-breach and post-breach) along with adjacent  
 256 sites (mudflat and salt marsh) plotted against fitted THg and MeHg concentrations that  
 257 were predicted using multiple linear regression models (Table 2) created with the same  
 258 data. The THg model included WEOC, pH, EC, and %Clay as explanatory variables,  
 259 explaining 51.9% of the variability. The MeHg model included EC, THg, pH, %Clay,  
 260 WEOC and %OM as explanatory variables, explaining 24.1% of the variability.

## **Discussion**

*Post-breach sediments are chemically more similar to the salt marsh and mudflat than pre-breach sediments*

The breaching of the dyke and inundation of the dyke cell deposited a large quantity of fresh sediment over the top of the pre-existing soil. This event changed the biogeochemistry of the system by increasing the EC, pH, %OM and WEOC. The impact of this change is best demonstrated by the considerable increase in the EC observed (Figure 1c) as the dyke cell changed from a terrestrial environment to a coastal environment due to inundation with saline water. While the topography of the mudflat and the salt marsh gently slopes down towards the sea, the soil in the dyke cell was relatively flat prior to breaching and inundation. The deposition of fresh sediment in the dyke cell was unevenly distributed leaving puddles of seawater which we observed in depressed areas at low tide. Evaporation of water and precipitation of salts in these depressed areas (Mouneimne and Price, 2007) has resulted in the EC of the post-breach sediments being elevated above levels measured in the mudflat or the salt marsh (Figure 1c).

The objective of the managed retreat is for salt marsh vegetation to colonise the freshly deposited sediment once the depth of the sediment raises the wetland to an elevation high enough for vegetation to survive (Williams and Orr, 2002). During the post-breach sampling in 2011 the dyke cell was still unvegetated and looked more similar to a mudflat than a salt marsh. This observation is supported by the textural analysis of the sediment deposited in the dyke cell (post-breach) which was similar to the sediment sampled from the mudflat (Figure 3). The chemistry of the post-breach sediments (WEOC, pH and %OM) was more similar to the salt marsh and

mudflat sediments than the samples collected pre-breach. However, this data must be interpreted with caution since only two cores were collected prior to the dyke being breached.

*Post-breach sediments have greater total Hg but lower MeHg concentrations*

The total Hg concentrations in the reclaimed region were similar to those found in other studies of non-vegetated intertidal mudflats (O'Driscoll et al., 2011; Sizmur et al., 2013b) and salt marshes (Hung and Chmura, 2006; Sunderland et al., 2004) in the Bay of Fundy. Over a period of two years (and only one year after the dyke was breached) the concentration of total Hg in the dyke cell was considerably greater (Figure 2). We acknowledge, however, that this dataset has limitations since there were only two replicate cores collected prior to the dyke being breached. Despite this apparent increase, the concentration of Hg in the post-breach sediments had not yet reached that of the salt marsh, which is the target ecosystem. There was a clear decrease in Hg concentration with sediment depth in both the mudflat and the post-breach sediments but not in the salt marsh sediments which is a further indication that the sediment characteristics more closely match the mudflat at this stage of restoration.

While the total Hg concentrations were greater in the dyke cell after inundation, and the MeHg concentrations were greater at the surface of the sediment, MeHg was observed to be lower overall in the post-breach cores (Figure 2). This lower MeHg concentration was reflected by %MeHg in the sediments of the dyke cell decreasing from 6% pre-breach to 2% post-breach when averaged over all the depths. This observation indicates that methylation has not rapidly

occurred in the newly deposited Hg(II) species in the sediment. If the lower MeHg in the post-breach sediments was due to greater export of MeHg from the sediments by tidal flushing then we would have expected to see MeHg depleted in the top few cm of sediment. However, MeHg concentrations were greatest in the top few cm (Figure 2). Therefore, tidal flushing is probably not the reason for the lower MeHg in the post-breach sediments.

Because total Hg concentrations are greater post-breach and MeHg concentrations are lower, Hg methylation cannot be limited by the supply of total Hg. Hg methylation is rather limited by the bioavailability of Hg to Hg methylating bacteria or the activity of these bacteria (Sunderland et al., 2006). Canário et al. (2007) showed that %MeHg in unvegetated coastal wetland sediments were only 0.6%, while vegetated sediments had up to 18% MeHg. The authors explained that this discrepancy is likely to occur because the presence of vegetation increases microbial activity and favours Hg methylation. Colonisation of the dyke cell by benthic invertebrates (e.g. polychaete worms) may also increase the sediment-water interface and the concentration of MeHg in their burrows (Sizmur et al., 2013a). Therefore, the MeHg concentrations in the dyke cell may increase as the restoration progresses and the dyke cell becomes colonised by vegetation and fauna. This prediction must be contrasted with the observation by Morris et al. (2014) that restored salt marshes have lower MeHg concentrations several decades after inundation when compared to adjacent natural salt marshes. It is therefore unclear whether the MeHg concentration in the dyke cell sediments will increase beyond the concentrations in the adjacent natural salt marsh in the long term.

327 THg concentrations are influenced by soluble carbon, particle size, pH, and salinity but MeHg  
328 concentrations are poorly predicted

329 Water-extractable organic carbon, pH, EC, and clay content of sediments all contributed to the  
330 multiple linear regression models that explained 52.4% of the variability associated with the  
331 concentration of THg, but only 24% of the variability associated with MeHg concentrations in  
332 the sediments (Table 2 and Figure 4). Clay content was positively correlated with THg sediment  
333 concentration. A reduction in sediment particle size (here observed by an increase in clay  
334 content) increases the surface-area-to-volume ratio of particulates in a system. The high surface  
335 area and cation exchange capacity of clays results in the adsorption of Hg to fine particles  
336 (Bengtsson and Picado, 2008). Suspension of fine sediments in the tidal water increases the  
337 likelihood of sediments to scavenge Hg from the water column by settling and retaining Hg in  
338 the accumulating sediment (Covelli et al., 2009; Hung and Chmura, 2006; Sunderland et al.,  
339 2006). Sediments comprised of fine particles also increase the proportion of particle-bound Hg  
340 (Bengtsson and Picado, 2008) and may thus reduce the bioavailability of Hg to methylating  
341 bacteria.

342

343 Dissolved organic matter (DOM) is a major binding phase for Hg in aquatic environments  
344 (Haitzer et al., 2002; Haverstock et al., 2012; Le Faucheur et al., 2014; O'Driscoll and Evans,  
345 2000; Ravichandran, 2004). Here we use WEOC as a proxy for DOM in the sediments following  
346 Sizmur et al (2013b). Although we found a positive correlation between THg and both %OM and  
347 WEOC, the WEOC explains the THg concentration in the sediments better (Table 2). This  
348 observation indicates that the changes in Hg in the sediments are due to a greater fraction that is  
349 bound to soluble carbon complexes. The concentration of WEOC in the post-breach sediments

(Figure 3) was higher than the mudflat and (unlike the salt marsh) was not associated with vegetation growing in situ. It is therefore likely that the cause of higher concentrations of WEOC (and THg) in the post-breach sediments, compared to the mudflat, is the decaying mat of terrestrial vegetation underneath the freshly deposited sediment. Hg complexation with DOM reduces the bioavailability of Hg to methylating bacteria because the complexes are generally too large to penetrate their biological membranes (Le Faucheur et al., 2014; Ravichandran, 2004). However, soluble organic matter also provides an energy source for methylating bacteria and may increase their activity resulting in greater methylation rates (Ullrich et al., 2001). Further increases in DOM (and microbial activity) are likely to occur as the dyke cell becomes vegetated (Canário et al., 2007) which may increase methylation rates in the future. The deposition of plankton is likely to increase the %MeHg in the fresh sediment as they contain approximately 6% to 15% MeHg in the Northwest Atlantic Ocean (Hammerschmidt et al., 2013).

The solubility and speciation of Hg and the binding of dissolved Hg species to DOM or sediment particles is pH dependent (Gabriel and Williamson, 2004). At low pH,  $H^+$  competes with Hg for binding sites on DOM or the surface of sediment particles, which releases Hg into solution but they also both compete for uptake by negatively charged bacterial cells. In this study pH correlated positively with Hg but negatively with MeHg. This contrast indicates that the greater pH of the mudflat, salt marsh, and post-breach sediments, compared to pre-breach sediments (Figure 3) resulted in greater Hg retention (Hung and Chmura, 2006) but may have reduced Hg bioavailability to methylating bacteria (Barkay et al., 1997; Gilmour and Henry, 1991; Le Faucheur et al., 2014).

The increase in EC that resulted from the inundation of the dyke cell with sea water (Figure 1c) is due to the high salinity of the seawater (Mouneimne and Price, 2007). The high salinity of the sediment deposited after the dyke cell was inundated with seawater created an environment with a higher ionic strength. As ionic strength increases, the concentration of Hg desorbed into solution decreases (Duarte et al., 1991) resulting in greater Hg retention in sediments and a decrease in the bioavailability of Hg to methylating bacteria (Barkay et al., 1997). Seawater contains high concentrations of chloride ions which can form strong ( $\log_{10} K_1^\circ = 7.31$ ) complexes with mercury species (Powell et al., 2005). The greater the concentration of chloride, the more negatively charged mercuric chloride ions ( $\text{HgCl}_3^-$  and  $\text{HgCl}_4^{2-}$ ) will be present in solution and these negative ions also have a lower bioavailability (Barkay et al., 1997) to methylating bacteria with negatively charged cell walls. Therefore, the increase in ionic strength and formation of trivalent or tetravalent mercuric chloride species in the high EC sediments of the post-breach sediments may have reduced their bioavailability to mercury methylating bacteria. These Hg-chloride complexes may also be less susceptible to photoreduction and loss to the atmosphere (Qureshi et al., 2009).

In summary, the chemical changes that occur in the sediment after inundation may have impacted on the bioavailability of Hg to methylating bacteria. The decrease in particle size distribution and subsequent increase in sediment surface area may have increased sorption of Hg out of the water column but lowered its bioavailability. Higher organic matter levels may provide a food source for methylating bacteria and increase their activity. Greater soluble organic carbon may mobilise Hg from the surface of sediments but also complex it in a form that is unavailable to methylating bacteria. An increase in sediment pH increases the concentration that can be



adsorbed from the solution phase and reduces the bioavailability. Finally, the higher ionic strength leads to a greater proportion of inorganic complexes and a lower bioavailability of Hg. This final conclusion assumes that the uptake of Hg by methylating bacteria occurs by passive diffusion of neutral or ionic lipophilic Hg species but there is now a considerable body of evidence to suggest that uptake may occur by facilitated diffusion or active transportation by protein pumps (Hsu-Kim et al., 2013).

#### Conclusions and Implications for Coastal Managed Retreat

Despite a doubling of Hg concentration within the dyke cell after the dyke was breached, Hg concentrations are still below the Canadian Sediment Quality Guidelines (CCME, 2002). The reason for the Hg increase in this study was the fresh deposition of sediments with a smaller particle size distribution that are able to scavenge and retain Hg due to their higher surface area, negative charge, and higher pH. This site can therefore be considered a net sink for mercury during the first year after the dyke was breached. The more sediment that is deposited, the larger the sink will become. In contrast to considerable increases in mercury methylation observed during freshwater wetland creation (Kelly et al., 1997; Sinclair et al., 2012), we observed a 27% decrease in MeHg concentrations in the dyke cell after the dyke was breached. This decrease may have been due to greater Hg retention and lower Hg bioavailability to methylating bacteria but ultimately cannot be fully explained with the available data and is limited by the low number of replicate cores collected. Further work will be required to explain the precise mechanisms for this decrease.

Our data provides no evidence for a flush of Hg methylation during the first year of managed retreat. As the restoration progresses and vegetation colonises, the soluble carbon concentration and microbial activity may increase and the rate of Hg methylation may also increase. However, contradictory data from other studies indicate that it is unclear whether MeHg will be elevated beyond the concentration found in natural wetlands (Canário et al., 2007; Kelly et al., 1997; Morris et al., 2014; Sinclair et al., 2012). We conclude that coastal flooding of sediments subject to diffuse Hg contamination during managed retreat of coastal flood defences does not pose a significant risk of increasing Hg methylation or bioavailability during the first year.

## **Acknowledgements**

Financial support for this research was provided by the Canada Research Chairs, the Ducks Unlimited Canada - Acadia University Partnership Grant, and an NSERC Discovery Grant awarded to NJO.

## **References**

- Barkay, T., Gillman, M., Turner, R.R., 1997. Effects of dissolved organic carbon and salinity on bioavailability of mercury. *Applied and environmental microbiology* 63, 4267-4271.
- Bengtsson, G., Picado, F., 2008. Mercury sorption to sediments: Dependence on grain size, dissolved organic carbon, and suspended bacteria. *Chemosphere* 73, 526-531.
- Byers, S.C., Mills, E.L., Stewart, P.L., 1978. A comparison of methods of determining organic carbon in marine sediments, with suggestions for a standard method. *Hydrobiologia* 58, 43-47.
- Canário, J., Caetano, M., Vale, C., Cesário, R., 2007. Evidence for elevated production of methylmercury in salt marshes. *Environmental science & technology* 41, 7376-7382.

441 CCME, 2002. Canadian Sediment Quality Guidelines for the Protection of Aquatic Life.

442 Compeau, G., Bartha, R., 1985. Sulfate-reducing bacteria: principal methylators of mercury in  
 443 anoxic estuarine sediment. *Applied and environmental microbiology* 50, 498-502.

444 Covelli, S., Acquavita, A., Piani, R., Predonzani, S., De Vittor, C., 2009. Recent contamination  
 445 of mercury in an estuarine environment (Marano lagoon, Northern Adriatic, Italy). *Estuarine,*  
 446 *Coastal and Shelf Science* 82, 273-284.

447 Crowell, N., Webster, T., O'Driscoll, N.J., 2011. GIS modelling of intertidal wetland exposure  
 448 characteristics. *Journal of Coastal Research* 27, 44-51.

449 Desplanque, C., Mossman, D.J., 2004. Tides and their seminal impact on the geology,  
 450 geography, history, and socio-economics of the Bay of Fundy, eastern Canada. *Atlantic Geology*  
 451 40.

452 Duarte, A., Pereira, M., Oliveira, J., Hall, A., 1991. Mercury desorption from contaminated  
 453 sediments. *Water Air & Soil Pollution* 56, 77-82.

454 Emmerson, R., Birkett, J., Scrimshaw, M., Lester, J., 2000. Solid phase partitioning of metals in  
 455 managed retreat soils: field changes over the first year of tidal inundation. *Science of the Total*  
 456 *Environment* 254, 75-92.

457 Evers, D.C., Han, Y.-J., Driscoll, C.T., Kamman, N.C., Goodale, M.W., Lambert, K.F., Holsen,  
 458 T.M., Chen, C.Y., Clair, T.A., Butler, T., 2007. Biological mercury hotspots in the northeastern  
 459 United States and southeastern Canada. *Bioscience* 57, 29-43.

460 Gabriel, M.C., Williamson, D.G., 2004. Principal biogeochemical factors affecting the speciation  
 461 and transport of mercury through the terrestrial environment. *Environmental geochemistry and*  
 462 *health* 26, 421-434.

463 Gilmour, C.C., Henry, E.A., 1991. Mercury methylation in aquatic systems affected by acid  
 464 deposition. *Environmental Pollution* 71, 131-169.

465 Gordon Jr, D.C., Baretta, J.W., 1982. A preliminary comparison of two turbid coastal  
 466 ecosystems: The Dollard (Netherlands-FRG) and the Cumberland Basin (Canada).  
 467 Hydrobiological Bulletin 16, 255-267.

468 Guédron, S., Huguet, L., Vignati, D., Liu, B., Gimbert, F., Ferrari, B., Zonta, R., Dominik, J.,  
 469 2012. Tidal cycling of mercury and methylmercury between sediments and water column in the  
 470 Venice Lagoon (Italy). Marine Chemistry 130, 1-11.

471 Haitzer, M., Aiken, G.R., Ryan, J.N., 2002. Binding of mercury (II) to dissolved organic matter:  
 472 the role of the mercury-to-DOM concentration ratio. Environmental science & technology 36,  
 473 3564-3570.

474 Hammerschmidt, C.R., Finiguerra, M.B., Weller, R.L., Fitzgerald, W.F., 2013. Methylmercury  
 475 accumulation in plankton on the continental margin of the northwest atlantic ocean.  
 476 Environmental science & technology 47, 3671-3677.

477 Haverstock, S., Sizmur, T., Murimboh, J., O'Driscoll, N.J., 2012. Modeling the photo-oxidation  
 478 of dissolved organic matter by ultraviolet radiation in freshwater lakes: Implications for mercury  
 479 bioavailability. Chemosphere 88, 1220-1226.

480 Heim, W.A., Coale, K.H., Stephenson, M., Choe, K.-Y., Gill, G.A., Foe, C., 2007. Spatial and  
 481 habitat-based variations in total and methyl mercury concentrations in surficial sediments in the  
 482 San Francisco Bay-Delta. Environmental science & technology 41, 3501-3507.

483 Hsu-Kim, H., Kucharzyk, K.H., Zhang, T., Deshusses, M.A., 2013. Mechanisms Regulating  
 484 Mercury Bioavailability for Methylating Microorganisms in the Aquatic Environment: A Critical  
 485 Review. Environmental science & technology 47, 2441-2456.

486 Hung, G.A., Chmura, G.L., 2006. Mercury accumulation in surface sediments of salt marshes of  
 487 the Bay of Fundy. Environmental Pollution 142, 418-431.

488 Kelly, C., Rudd, J., Bodaly, R., Roulet, N., St. Louis, V., Heyes, A., Moore, T., Schiff, S.,  
 489 Aravena, R., Scott, K., 1997. Increases in fluxes of greenhouse gases and methyl mercury  
 490 following flooding of an experimental reservoir. Environmental science & technology 31, 1334-  
 491 1344.

492 Lavoie, R.A., Hebert, C.E., Rail, J.-F., Braune, B.M., Yumvihoze, E., Hill, L.G., Lean, D.R.,  
 493 2010. Trophic structure and mercury distribution in a Gulf of St. Lawrence (Canada) food web  
 494 using stable isotope analysis. *Science of the Total Environment* 408, 5529-5539.

495 Le Faucheur, S., Campbell, P.G.C., Fortin, C., Slaveykova, V.I., 2014. Interactions between  
 496 mercury and phytoplankton: Speciation, bioavailability, and internal handling. *Environmental*  
 497 *Toxicology and Chemistry* 33, 1211-1224.

498 Miller, W., Miller, D., 1987. A micro-pipette method for soil mechanical analysis.  
 499 *Communications in Soil Science & Plant Analysis* 18, 1-15.

500 Morris, M.A., Spencer, K.L., Belyea, L.R., Branfireun, B.A., 2014. Temporal and spatial  
 501 distributions of sediment mercury in restored coastal saltmarshes. *Marine Chemistry* 167, 150-  
 502 159.

503 Mouneimne, S.M., Price, J.S., 2007. Seawater contamination of a harvested bog: hydrological  
 504 aspects. *Wetlands* 27, 355-365.

505 O'Driscoll, N.J., Evans, R.D., 2000. Analysis of methyl mercury binding to freshwater humic  
 506 and fulvic acids by gel permeation chromatography/hydride generation ICP-MS. *Environmental*  
 507 *science & technology* 34, 4039-4043.

508 O'Driscoll, N.J., Canário, J., Crowell, N., Webster, T., 2011. Mercury speciation and distribution  
 509 in coastal wetlands and tidal mudflats: relationships with sulphur speciation and organic carbon.  
 510 *Water, Air, & Soil Pollution* 220, 313-326.

511 Ollerhead, J., Spicer, C., Bams, R., 2011. Monitoring a salt marsh restoration at Fort Beausejour,  
 512 Aulac, NB - Final Report, Ducks Unlimited Canada.

513 Portnoy, J., Giblin, A., 1997. Effects of historic tidal restrictions on salt marsh sediment  
 514 chemistry. *Biogeochemistry* 36, 275-303.

515 Powell, K.J., Brown, P.L., Byrne, R.H., Gajda, T., Hefter, G., Sjöberg, S., Wanner, H., 2005.  
 516 Chemical speciation of environmentally significant heavy metals with inorganic ligands. Part 1:  
 517 The  $\text{Hg}^{2+}$ - $\text{Cl}^-$ ,  $\text{OH}^-$ ,  $\text{CO}_3^{2-}$ ,  $\text{SO}_4^{2-}$ , and  $\text{PO}_4^{3-}$ -aqueous systems (IUPAC Technical Report).  
 518 *Pure and applied chemistry* 77, 739-800.

519 Qureshi, A., O'Driscoll, N.J., MacLeod, M., Neuhold, Y.-M., Hungerbühler, K., 2009.  
 520 Photoreactions of mercury in surface ocean water: gross reaction kinetics and possible pathways.  
 521 Environmental science & technology 44, 644-649.

522 Rasmussen, R.S., Nettleton, J., Morrissey, M.T., 2005. A review of mercury in seafood: Special  
 523 focus on tuna. Journal of Aquatic Food Product Technology 14, 71-100.

524 Ravichandran, M., 2004. Interactions between mercury and dissolved organic matter—a review.  
 525 Chemosphere 55, 319-331.

526 Sinclair, K.A., Xie, Q., Mitchell, C.P., 2012. Methylmercury in water, sediment, and  
 527 invertebrates in created wetlands of Rouge Park, Toronto, Canada. Environmental Pollution 171,  
 528 207-215.

529 Singh, K., Walters, B.B., Ollerhead, J., 2007. Climate change, sea-level rise and the case for salt  
 530 marsh restoration in the Bay of Fundy, Canada. Environments: a journal of interdisciplinary  
 531 studies 35.

532 Sizmur, T., Canário, J., Edmonds, S., Godfrey, A., O'Driscoll, N.J., 2013a. The polychaete worm  
 533 Nereis diversicolor increases mercury lability and methylation in intertidal mudflats.  
 534 Environmental Toxicology and Chemistry 32, 1888-1895.

535 Sizmur, T., Canário, J., Gerwing, T.G., Mallory, M.L., O'Driscoll, N.J., 2013b. Mercury and  
 536 methylmercury bioaccumulation by polychaete worms is governed by both feeding ecology and  
 537 mercury bioavailability in coastal mudflats. Environmental Pollution 176, 18-25.

538 Sizmur, T., Palumbo-Roe, B., Hodson, M.E., 2011. Impact of earthworms on trace element  
 539 solubility in contaminated mine soils amended with green waste compost. Environmental  
 540 Pollution 159, 1852-1860.

541 Sunderland, E.M., Amirbahman, A., Burgess, N.M., Dalziel, J., Harding, G., Jones, S.H., Kamai,  
 542 E., Karagas, M.R., Shi, X., Chen, C.Y., 2012. Mercury sources and fate in the Gulf of Maine.  
 543 Environmental research 119, 27-41.

544 Sunderland, E.M., Gobas, F.A., Branfireun, B.A., Heyes, A., 2006. Environmental controls on  
 545 the speciation and distribution of mercury in coastal sediments. Marine Chemistry 102, 111-123.

546 Sunderland, E.M., Gobas, F.A., Heyes, A., Branfireun, B.A., Bayer, A.K., Cranston, R.E.,  
547 Parsons, M.B., 2004. Speciation and bioavailability of mercury in well-mixed estuarine  
548 sediments. *Marine Chemistry* 90, 91-105.

549 U.S.EPA, 1998. Method 7473: Mercury in solids and solutions by thermal decomposition,  
550 amalgamation and atomic spectrophotometry, U.S. Environmental Protection Agency,  
551 Washington, DC.

552 Ullrich, S.M., Tanton, T.W., Abdrashitova, S.A., 2001. Mercury in the aquatic environment: a  
553 review of factors affecting methylation. *Critical Reviews in Environmental Science and*  
554 *Technology* 31, 241-293.

555 Williams, P.B., Orr, M.K., 2002. Physical evolution of restored breached levee salt marshes in  
556 the San Francisco Bay estuary. *Restoration Ecology* 10, 527-542.

557 Wynn, G., 1979. Late eighteenth-century agriculture on the Bay of Fundy marshlands.  
558 *Acadiensis*, 80-89.

559

560