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Ebullition was a major pathway of methane emissions from the aquaculture ponds in southeast China

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Abstract

Aquaculture ponds are hotspots of carbon cycling and important anthropogenic sources of the potent greenhouse gas methane (CH₄). Despite the importance of CH₄ ebullition in aquatic ecosystems, its magnitude and spatiotemporal variations in aquaculture ponds remain poorly understood. In this study, we determined the rates and spatiotemporal variations of ebullitive CH₄ emissions from three mariculture ponds during the aquaculture period of two years at a subtropical estuary in southeast China. Our results showed that the mean ebullitive CH₄ flux from the studied ponds was 14.9 mg CH₄ m⁻² h⁻¹ during the aquaculture period and accounted for over 90% of the total CH₄ emission, indicating the importance of ebullition as a major CH₄ transport mechanism. Ebullitive CH₄ emission demonstrated a clear seasonal pattern, with a peak value during the middle stage of aquaculture. Sediment temperature was found to be an important factor influencing the seasonal variations in CH₄ ebullition. Ebullitive CH₄ fluxes also exhibited considerable spatial variations within the ponds, with 49.7–71.8% of the whole pond CH₄ ebullition being detected in the feeding zone where the large loading of sediment organic matter fueled CH₄ production. Aquaculture ponds have much higher ebullitive CH₄ effluxes than other aquatic ecosystems, which indicated the urgency to mitigate CH₄ emission from aquaculture activities. Our findings highlighted that the importance of considering the large spatiotemporal variations in ebullitive CH₄ flux in improving the accuracy of large-scale estimation of CH₄ fluxes in aquatic ecosystems. Future studies should be conducted to characterize CH₄ ebullitive fluxes over a greater number and diversity of aquaculture ponds and examine the mechanisms controlling CH₄ ebullition in aquatic ecosystems.

1. Introduction

Rising concentrations of atmospheric greenhouse gases since the beginning of the industrial era are of international concern owing to their huge implications to climate change

(IPCC, 2019; Tangen et al., 2016). Methane (CH₄) is a potent greenhouse gas that has a sustained-flux global warming potential of 45 times higher than that of CO₂ on a per mass unit basis over a 100-year time horizon (Neubauer and Megonigal, 2015), and contributes to approximately 20% of the global radiative forcing (IPCC, 2013). Global average atmospheric CH₄ concentration has increased from 722 ppb in 1750 to 1875 ppb in 2019 (National Oceanic and Atmospheric, 2020), exceeding the pre-industrial levels by about 150%. Thus, determining the sources, magnitude, and mechanisms of CH₄ emissions around the world is crucial for mitigating global climate change and the associated adverse effects on the environment and human society.

Aquatic systems are potentially significant contributors to global CH₄ emissions. Previous studies have reported that inland waters including rivers (Borges et al., 2015; Yuan et al., 2019), lakes (Bastviken et al. 2011; Tangen et al., 2016; Yang et al., 2011), reservoirs (Musenze et al., 2014; Yang et al., 2012) are strong sources of atmospheric CH₄. Small ponds, especially those perturbed by humans, form an indispensable part of the global inland water systems, and play an important role in the biogeochemical cycling of carbon and the release of CH₄ (Holgerson, 2015; Holgerson and Raymond, 2016; Rubbo et al., 2006; Yuan et al., 2019). Over the past two decades, large efforts have been made to evaluate the concentrations, fluxes, and drivers of CH₄ in small ponds from different countries, including Sweden (Bastviken et al., 2004; Natchimuthu et al., 2014; Peacock et al., 2019), Germany (Ortega et al., 2019), Canada (Hamilton et al., 1994; Laurion et al., 2010; Pelletier et al., 2014), India (Panneer Selvam et al., 2014), the USA (Holgerson, 2015; Gorsky et al., 2019), Netherlands (van Bergen et al., 2019) and Australia (Grinham et al., 2018a). Aquaculture ponds form an important component of the global man-made aquatic ecosystems. The freshwater and brackish aquaculture ponds, with a global total surface area of around 1.1×10^5 km² (Verdegem and Bosma, 2009), are potential hotspots of CH₄ emission because of their large loadings of organic matter from residual feeds and feces of cultured animals (IPCC, 2019; Yang et al., 2018, 2019; Yuan et al., 2019). Although numerous studies have quantified CH₄ fluxes from the aquaculture ponds (Chen et al., 2016; Ma et al., 2018; Soares and Henry-Silva, 2019; Wu et al., 2018; Yang et al., 2018, 2019; Yuan et al., 2019), the biogeochemical processes involved (e.g., CH₄ production, oxidation, and transport) in the aquaculture systems have thus far received little attention (Avnimelech and Ritvo, 2003).

Biogenic CH₄ is produced during the terminal step of organic matter degradation in anaerobic sediments (Lofton et al., 2015) and in oxic waters with rich planktonic microbes (Bogard et al., 2014; Khatun et al., 2019; Tang et al., 2016). CH₄ can be exported from the sediment to the atmosphere through gas ebullition, molecular diffusion, or plant-mediated transport (Bastviken et al., 2004, 2008; Wu et al., 2019; Yang et al., 2015). The majority of existing studies determined the diffusive CH₄ fluxes from aquatic ecosystems based on the wind-based estimations of gas transfer velocity (piston velocity, k) along with measurements of surface water CH₄ concentrations (Borges et al., 2018; Cole and Caraco, 1998; Cotovicz et al., 2016). On the other hand, CH₄ ebullition in freshwater systems (e.g., lakes, reservoirs, ponds) has traditionally been measured by directly capturing the released bubbles with inverted funnels (Davidson et al., 2018; Wik et al., 2013; Zhou et al., 2019) or indirectly with the use of floating chambers (*FCs*) (Bastviken et al., 2004; Chuang et al., 2017; Natchimuthu et al., 2014; Wu et al., 2019). To estimate the ebullition, the total CH₄ flux from the non-vegetated water bodies measured by the *FCs* are compared against the diffusive CH₄ flux determined by the gas transfer coefficient method (Bastviken et al., 2004; Chuang et al., 2017; Natchimuthu et al., 2014; Wu et al., 2019).

Although the upscaling of chamber-based CH₄ fluxes is subjected to potential bias when ebullition rates are spatially heterogeneous, *FCs* have been widely used in flux measurement owing to their low cost and relative ease of operation (Bastviken et al., 2015; Lorke et al., 2015; Podgrajsek et al., 2014). Recently, some state-of-the-art techniques, such as hyperspectral image analysis (Gålfalk et al., 2016), optical detectors (Grinham et al., 2011), hydroacoustic (sonar) technology (DelSontro et al., 2011) and automated bubble traps (Maher et al., 2019), have also been employed to measure CH₄ ebullition.

Ebullition is often considered as a dominant CH₄ emission pathway in the shallow area of reservoirs (Deshmukh et al., 2016), rivers (Wu et al., 2019), and lakes (Bastviken et al., 2004; Natchimuthu et al., 2016; Xiao et al., 2017). However, the magnitude of CH₄ ebullition can vary from time to time both within and across ecosystems depending on the CH₄ production rate (Zhou et al., 2019), trophic state (Bastviken et al., 2008; Walter et al., 2008; Zhou et al., 2019), water depth, and other environmental factors (e.g., water temperature and atmospheric pressure) (Maeck et al., 2014; Zhu et al., 2016). Organic matter in sediment is a major source of substrate for CH₄ production in aquatic ecosystems (Bastviken et al., 2008; Walter et al., 2008; Zhu et al., 2016). CH₄ ebullition generally increases with organic matter content (DelSontro et al., 2011; Zhou et al., 2019; Zhu et al., 2016), but this process is highly stochastic (Bastviken et al., 2004) with large spatiotemporal variations (e.g., DelSontro et al., 2011; Natchimuthu et al., 2016; Wilkinson et al., 2015). Recent studies have suggested that ebullition could dominate CH₄ emissions from shallow ponds (e.g., Natchimuthu et al., 2014; Yang et al., 2019a). Without *in situ* sampling and observation, the magnitude and spatiotemporal variations of ebullitive CH₄ fluxes in aquaculture ponds remain largely unknown, which limit our ability in accurately determining CH₄ fluxes from shallow ponds and their contributions to the global CH₄ budget.

China has the largest area of aquaculture ponds in the world (approximately 25,700 km²; 23% of the global total), with many of them being widely located in the subtropical estuaries along the coastal regions (Ren et al., 2019; Yang et al., 2018). Shrimp mariculture is one of the dominant types of land-based aquaculture in the coastal areas of China (Yang et al., 2017a). These shrimp ponds are generally maintained through daily supply of feeds (Yang et al., 2017a), with a rather low feed utilization efficiency in the range of ~4.0% - 27.4% (Chen et al., 2015; Molnar et al., 2013). Consequently, the majority of the uneaten feeds, along with animal excreta and dead phytoplankton, deposit on the sediment surface. The microbial decomposition of these organic-rich materials can stimulate CH₄ production and thereby contribute to a large amount of ebullitive CH₄ flux. However, the contribution of ebullitive CH₄ fluxes in aquaculture ponds is largely understudied, considering the enormous number and size of these ponds globally. Furthermore, mariculture ponds are largely affected by topography, environmental factors (e.g., sediment organic matter and temperature), and management practices (Yang et al., 2018; 2019). This could lead to considerable spatiotemporal variations in ebullitive CH₄ fluxes, which should be further characterized and tested by field observation.

To the best of our knowledge, this study made the first attempt to estimate ebullitive CH₄ fluxes from the intensive mariculture ponds. The intensity of mariculture pond culture has increased progressively over the last few decades (Wang et al., 2018). Intensive shrimp pond is the most dominant type of land-based aquaculture ponds in the coastal region in China with an area of 2.4×10³ km² (Bureau of Fisheries of the Ministry of Agriculture, 2019), accounting for approximately 12% of the total global area of shrimp aquaculture ponds. This research aimed to: (1) determine the magnitude, spatiotemporal variations, and main controls

of ebullitive CH₄ fluxes within the ponds; and (2) examine the relative contribution of ebullition to total CH₄ fluxes from the mariculture ponds. We hypothesized that: (1) ebullitive CH₄ fluxes would exhibit remarkable temporal variations in response to differences in environmental variables (e.g., temperature and substrate availability); (2) ebullitive CH₄ fluxes would vary significantly among different zones (nearshore area, feeding area, and aeration area) within the ponds owing to differences in oxygen and substrate supply; and (3) ebullition would contribute markedly to the total CH₄ emissions from the mariculture ponds owing to the large organic matter load and shallow water depth.

2. Materials and methods

2.1. Study area and shrimp pond system

The study was conducted in the intensive aquaculture ponds with monoculture of white shrimps (*Litopenaeus vannamei*) located at the Shanyutan Wetland in the Min River Estuary (MRE) southeast China (26°00'36"–26°03'42"N, 119°34'12"–119°40'40"E, Fig. 1). White shrimp is the most important type of maricultured crustacean species in China with an annual yield of approximately 875,000 tons, which is over 6 times higher than the yield of mud crab (Wang et al., 2018). Moreover, intensive shrimp is particularly dominant in the southern part of China (Yuan et al., 2006). Hence, our study ponds represented a very common management regime of mariculture ponds in the region. The study area is influenced by a subtropical monsoonal climate with annual mean temperature of 19.60 °C and annual precipitation of 1,350 mm (Tong et al., 2010). The total area of shrimp ponds at the Shanyutan wetland is about 234 ha (Yang et al., 2017a). These shrimp ponds were created in 2011 by complete removal of original marsh vegetations (Yang et al., 2017b). Field measurement was conducted in three selected ponds, which were representative of the physical setting and management practices of mariculture ponds in the surrounding region. These three ponds had a small size of 19,000 m², which was close to the range of 20,000–30,000 m² found in Thailand and Taiwan (Kongkeo, 1997) and 33,300–66,700 m² reported in Chinese shrimp ponds (Xie & Yu, 2007). Moreover, our ponds had shallow water depths ranging between 1.1 and 1.8 m over the culture period (Yang et al., 2019a), which were close to the mean of 1.7 m reported in tropical China (Herbeck et al., 2013) and 1.0 m in shrimp ponds across China (Cao et al., 2007).

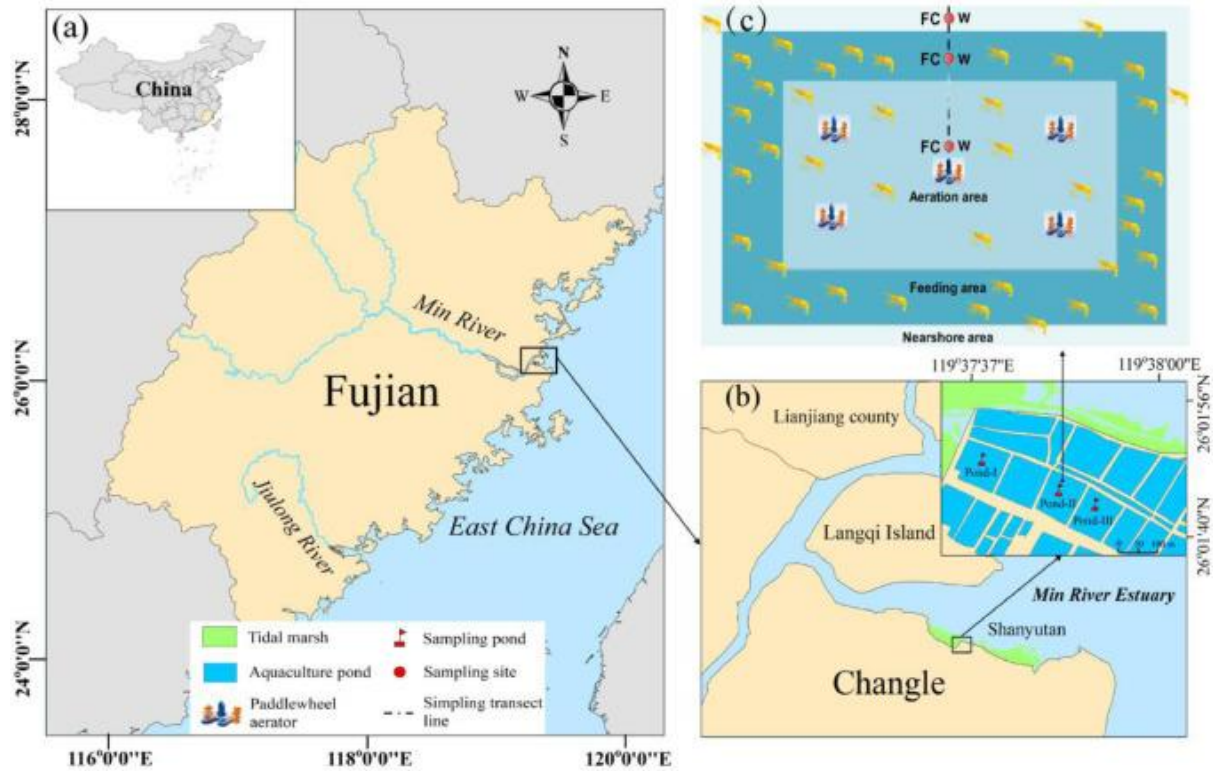


Fig. 1. (a) Location of the study area; (b) sampling sites and ponds in Shanyutan wetland of Min River estuary, southeast China; and (c) schematic diagram of aquaculture shrimp pond and the location of sampling sites (red dots). W = water samples; FC = flux measurement using floating chambers.

Shrimp aquaculture usually starts in June and ends in November, with only one single crop being produced each year (Yang et al., 2017a). Before the start of the production cycle, these ponds were filled with seawater from the Min River estuary using a submerged pump (Yang et al., 2018, 2019). There was no water exchange during the culture period, except occasional input by rainfall events. The stocking density of shrimps in our ponds was around 35–45 individuals m^{-2} (Yang et al., 2017a), which was comparable to 54 and 38–73 individuals m^{-2} in other intensive shrimp ponds in China (Xie & Yu, 2007), and the Philippines and Taiwan (Kongkeo, 1997), respectively. The shrimps were regularly fed with artificial feeds (Hangsheng and Tianma Chia Tai Feed Co., Ltd., Fuzhou, China) twice per day (07:00 AM and 16:00 PM, local standard time) manually from a boat. These pellet feeds have been commonly used in mariculture ponds across China (Wang et al., 2018). The feed conversion ratio in our ponds ranged between 1.11 and 1.72, which was lower than 2.1 reported in other intensive shrimp ponds in China (Xie and Yu, 2007) but similar to the range of 1.38–1.67 in other Asian countries including Indonesia, Taiwan and Thailand (Kongkeo, 1997). Generally, three to five automatic paddlewheel aerators were operated four times per day (00:00–03:00, 07:00–09:00, 12:00–14:00, and 18:00–20:00) to increase the oxygen content in water. Paddlewheel aerators are equipped in most of the intensive shrimp ponds in China and operated either continuously or a few hours in the day depending on weather (Herbeck et al., 2013). According to the spatial variations in microtopographic features, water depth, and management practices, the shrimp ponds were divided into three distinct zones, namely Zone N (near-shore area with sparse submerged vegetation), Zone F

(deep-water area for feeding activities) and Zone A (shallow-water area for aeration) (Fig. S1 and Table S1, Yang et al., 2019a; Zhang et al., 2019).

2.2. Experimental design

To determine the spatiotemporal variations in CH₄ emissions within the ponds during the culture period, we collected water, sediment, and gas samples from three replicate shrimp ponds at the Shanyutan Wetland (Fig. 1). A wooden bridge was built in each pond to facilitate the access to the sampling sites and minimize the potential disturbance to the pond ecosystem. Field sampling campaigns were carried out at all three zones (N, F, and A) in the ponds (Fig. 1) between June and November in 2017, and May and October in 2018 following the timing of aquaculture management practices. Gas fluxes were measured and water samples were taken 2–3 times each month. Sediment samples were collected during the initial (June or May), middle (August), and final (October) stages of shrimp production. On each sampling day, all samples were taken between 9:00 am and 11:00 am to minimize the potential discrepancies owing to diurnal variations (Chen et al., 2016; Wu et al., 2019; Zou et al., 2015).

2.3. Dissolved CH₄ concentration and diffusive CH₄ flux

Bubble-free water samples were collected at the depth of 20 cm using a gas-tight water sampler for dissolved CH₄ concentration analysis. Subsequently, 55-mL gas-tight serum glass bottles were flushed with the collected water samples thoroughly for three times before being completely filled with the water samples. Prior to sealing the bottles, approximately 0.2 mL of saturated HgCl₂ solution was injected into each sample to inhibit bacterial activity (Borges et al., 2018). The glass bottles were then immediately sealed without headspace using a halobutyl rubber septum to exclude any air bubbles (Borges et al., 2018; Xiao et al., 2017). The water samples were then stored in an ice-packed cooler, transported to the laboratory within 4–6 h, and analyzed within two days.

The concentrations of dissolved CH₄ in the water samples were determined using the headspace equilibrium technique and a gas chromatograph (GC-2010, Shimadzu, Kyoto, Japan) equipped with a flame ionization detector. Five CH₄ standards, namely 2, 8, 500, 1000, and 10,000 ppm, were used in the calibration. The CH₄ detection limit was 0.3 ppm, and the relative standard deviations of CH₄ analyses were $\leq 2.0\%$ in 24 h. Details about the gas sampling procedures and configurations of the gas chromatograph can be found in our previous paper (Yang et al., 2019a). The *in situ* dissolved CH₄ concentration in surface water (C_{water}) was calculated based on CH₄ concentration in the headspace of the glass bottles after equilibration (Eq. (1); Hu et al., 2018):

$$C_{\text{Water}} = \frac{(C_{\text{H}} - C_{\text{A}}) \times V_{\text{H}} + \alpha \times C_{\text{H}} \times V_{\text{W}}}{V_{\text{W}}}$$

where C_{H} is CH₄ concentration ($\mu\text{mol L}^{-1}$) in the headspace of the glass bottles at equilibrium; C_{A} is the atmospheric CH₄ concentration ($\mu\text{mol L}^{-1}$) at the prevailing *in situ* conditions; V_{H} is the volume of headspace gas in the glass bottles (L); α is the Bunsen coefficient (Wanninkhof, 1992); and V_{W} is the volume of water in the glass bottles (L).

The diffusive CH₄ fluxes (F_{D} , $\mu\text{mol m}^{-2} \text{h}^{-1}$) across the water-atmosphere interface were calculated based on the gas transfer coefficient method (Eq. (2); Musenze et al., 2014; Xiao et al., 2017):

$$F_{\text{D}} = k \times (C_{\text{Water}} - C_{\text{Eq}})$$

where C_{Water} ($\mu\text{mol L}^{-1}$) is the dissolved CH₄ concentration measured at the water surface; C_{Eq} ($\mu\text{mol L}^{-1}$) is the CH₄ concentration in

equilibrium with the atmospheric concentration at the prevailing *in situ* conditions; and k_x is the gas transfer velocity (cm h^{-1}). In the present study, we report the unit of CH_4 diffusive flux as $\text{mg m}^{-2} \text{h}^{-1}$ ($= F_D \times M \times 10^6$), where M is molar mass of CH_4 (16 mg mol^{-1}).

Considering the influence of varying wind speeds on the estimated k_x value, we adopted the model of Cole and Caraco (1998), which was developed under wind conditions most similar to our studied aquaculture ponds, to

calculate k_x values : (3) $k=(Sc/600)-0.5 \times (2.07+0.215 \times U_{10}^{1.7})$ where Sc is the Schmidt number for CH_4 corrected by *in situ* water temperature (Jahne et al., 1987; MacIntyre et al., 1995; Xiao et al., 2017); and U_{10} is the frictionless wind speed at 10 m height (m s^{-1}) according to Eq. (4) (Musenze et al., 2014; Sibgh et al., 2007): (4) $U_{10}=U_z[1+(C_{d10})^{1/2}K\ln(10z)]$ where U_z (m s^{-1}) is the measured wind speed at height z (m) above the water surface (here at 2.5 m); C_{d10} is the drag coefficient at 10 m above the water surface (0.0013 m s^{-1}); and K is the von Karman constant (0.41). Wind speed was measured and recorded by an automatic meteorological station (Vantage Pro 2, China) installed at the MRE weather station, China Wetland Ecosystem Research Network.

2.4. Ebullitive CH_4 flux estimation

In order to estimate ebullitive CH_4 flux from the aquaculture ponds, the total CH_4 fluxes across the water-atmosphere interface were measured using the floating chamber method. On each sampling day, three plastic FC s were deployed on one transect spanning across Zones N, F, and A in each pond (Fig. 1c). The FC s were similar to those used in previous studies (Chuang et al., 2017; Lorke et al., 2015; Natchimuthu et al., 2014, 2017). Briefly, the opaque FC covering an area of 0.1 m^2 with a volume of 5.2 L was made from plastic basin (polyethylene/plexiglas®) with floats at the sides, and covered with reflective aluminum foil to minimize internal heating by sunlight. A fan was installed inside the chamber to mix the headspace air during gas sampling. In each sampling location, four gas samples (approximately 50 mL each) were taken from the chamber air headspace at 15-minute intervals over a total duration of 45 min, and transferred into 100 mL pre-evacuated airtight gas sampling bags (Dalian Delin Gas Packing Co., Ltd., China). The CH_4 concentration in the gas samples were determined using a gas chromatograph (GC-2010, Shimadzu, Kyoto, Japan) within 24 h after sampling. The rates of CH_4 flux ($\text{mg m}^{-2} \text{h}^{-1}$) were calculated by the regression of CH_4 concentration against time (Yang et al., 2019a). If the plastic floating chambers showed distinct nonlinear increases in CH_4 concentration, it was considered that the CH_4 fluxes from both diffusion and ebullition were captured (e.g., Chuang et al., 2017; Keller and Stallard, 1994; Zhu et al., 2016). In the current study, all chambers showed distinct nonlinear increases in CH_4 concentration, with the chamber-based fluxes being substantially larger than the diffusive fluxes calculated based on the gas transfer coefficient method. Therefore, the ebullitive CH_4 flux from our ponds was estimated by deducting the diffusive flux determined by the gas transfer model from the total CH_4 flux measured by the FC s (Chuang et al., 2017; Xiao et al., 2017; Yang et al., 2019a).

2.5. Measurement of sediment CH_4 production rates

Sediment cores (~15 cm length) were collected from the three different zones in each pond in May/mid-June, August, and October, which represented the initial, middle, and final culture stages, respectively, in 2017. The sediment cores were collected using a steel sediment sampler (internal diameter of 5 cm), and placed into incubation chambers made of transparent

Plexiglas (inner diameter = 5 cm, height = 25 cm). All the sediment samples were stored in an ice box and transported back to the laboratory within 4 h. Considering that the dissolved oxygen concentrations of the surface sediment in aquaculture ponds were almost close to zero, the sediment CH₄ production rates were measured using the anoxic incubation method in a dark and shaking incubator (Inglett et al., 2012; Wassmann et al., 1998; Yang et al., 2018). Before the start of incubation, the chambers were purged with ultrapure nitrogen (N₂) gas for 5–8 min to create an anaerobic condition (Vizza et al., 2017; Wassmann et al., 1998). The incubation chambers were then incubated for 6 days at *in situ* temperatures. Approximately 5 mL of gas samples were collected from the chamber using a polypropylene syringe equipped with a three-way stopcock at two-day intervals over the incubation period. After gas sampling, 5 mL of N₂ gas was added back into the chamber to re-establish the ambient atmospheric pressure. CH₄ concentrations in the gas samples were analyzed using a gas chromatograph. Sediment CH₄ production rates [$\mu\text{g CH}_4 \text{ g}^{-1}$ (dry weight) day^{-1}] were calculated based on the rate of change in chamber headspace CH₄ concentrations over the incubation period (Wassmann et al., 1998). In the present study, the dry weight was calculated by multiplying the wet weight of incubation sediment sample by (1 - the moisture content of incubation sediment sample).

2.6. Measurement of ancillary data

During each sampling campaign, various parameters of surface water (e.g., water temperature, salinity, pH, and dissolved oxygen) were determined *in situ* at a depth of 20 cm at each site. Water temperature and pH were measured using a portable pH/mV/Temperature meter (IQ150, IQ Scientific Instruments, USA). Water salinity and dissolved oxygen (DO) were determined using a salinity meter (Eutech Instruments-Salt6, USA) and a multiparameter probe (550A YSI, USA), respectively. The detection limits for pH, salinity, and DO were 0.01, 0.1 ppt and 0.1 mg L⁻¹, respectively. The relative standard deviations of pH, salinity, and DO analyses were $\leq 1.0\%$, $\leq 1.0\%$, and $\leq 2.0\%$, respectively. Meteorological parameters (e.g., air temperature, atmospheric pressure, and precipitation) were obtained from the automatic meteorological station (Vantage Pro 2, China) at the MRE weather station. The precision levels of air temperature, atmospheric pressure, and precipitation measurements were ± 0.2 °C, ± 1.5 hPa, and ± 0.4 mm min⁻¹, respectively.

Sediment pH and electrical conductivity (EC) were determined using a pH meter (Orion 868, USA) and an EC meter (2265FS, Spectrum Technologies Inc., Phoenix, AZ, USA), respectively, in a slurry with a sediment to water ratio of 1:2.5 (w/v). A subsample of sediment was freeze-dried, homogenized, and then grounded to a fine powder for the analysis of total carbon (TC), total nitrogen (TN), N-NO₃⁻ and SO₄²⁻. Sediment TC and TN was determined using an elemental analyser (Elementar Vario MAX CN, Germany). The concentrations of N-NO₃⁻ and SO₄²⁻ were determined following the methods of Tu et al. (2010), and Chen and Sun (2020), respectively, using a flow injection analyser (Skalar Analytical SAN⁺⁺, Netherlands). The detection limits for TC and TN were 4 $\mu\text{g L}^{-1}$ and 3 $\mu\text{g L}^{-1}$, respectively, and the measurement reproducibilities were $\leq 1.0\%$ and $\leq 2.0\%$, respectively. The physico-chemical properties of the shrimp pond sediment in 2017 were measured from the sediment samples at 0–15 cm depth from the three different zones in each pond, while the sediment properties in 2018 were the averages at two sampling depths (0–5, and 5–15 cm) at each sampling site.

2.7. Statistical analyses

Data were tested for normality and homogeneity of variance. Log-transformation of data was conducted for ebullitive CH₄ fluxes and sediment physicochemical parameters when necessary prior to the analysis of variance (ANOVA). Two-way ANOVA was conducted to examine the effects of sampling zones (Zones N, F, and A), sampling time, and their interactions on ebullitive CH₄ fluxes (or sediment physicochemical parameters) in aquaculture ponds with sampling ponds being specified as a random variable. The differences in mean ebullitive CH₄ fluxes and environmental variables between 2017 and 2018 were also tested using the independent samples *t*-test. Spearman correlation analyses were conducted to estimate the relationships (1) between ebullitive CH₄ fluxes (or sediment CH₄ production rates) and the environmental variables, and (2) between ebullitive CH₄ fluxes and sediment CH₄ production rates. Principal component analysis (PCA) was also performed to analyze the relationships between the ebullitive CH₄ fluxes and environmental parameters at different culture stages. All statistical analyses were conducted using SPSS statistical software (v. 17.0, SPSS Inc., USA) at a significance level of 0.05. The results were presented as means \pm 1 standard error. All statistical plots were generated using OriginPro 7.5 (OriginLab Corp. USA).

3. Results

3.1. Surface sediment characteristics and gas transfer velocities

The spatial variations in sediment physico-chemical variables across the different zones within the ponds over the study period were shown in Figs. 2 and S1a. Overall, the mean sediment TC (Fig. 2a) and TN (Fig. 2b) concentrations were significantly higher in Zone F than the other two zones ($p < 0.01$; Table S1). In contrast, sediment SO₄²⁻ (Fig. 2c), N-NO₃⁻ (Fig. 2d) and EC (Fig. 2f) were lowest in Zone F, followed by Zone N and Zone A (ANOVA, $p < 0.05$; Table S1). However, no significant difference in sediment pH (Fig. 2e) and temperature (Fig. S1a) was observed among the three zones (ANOVA, $p > 0.05$; Table S1).

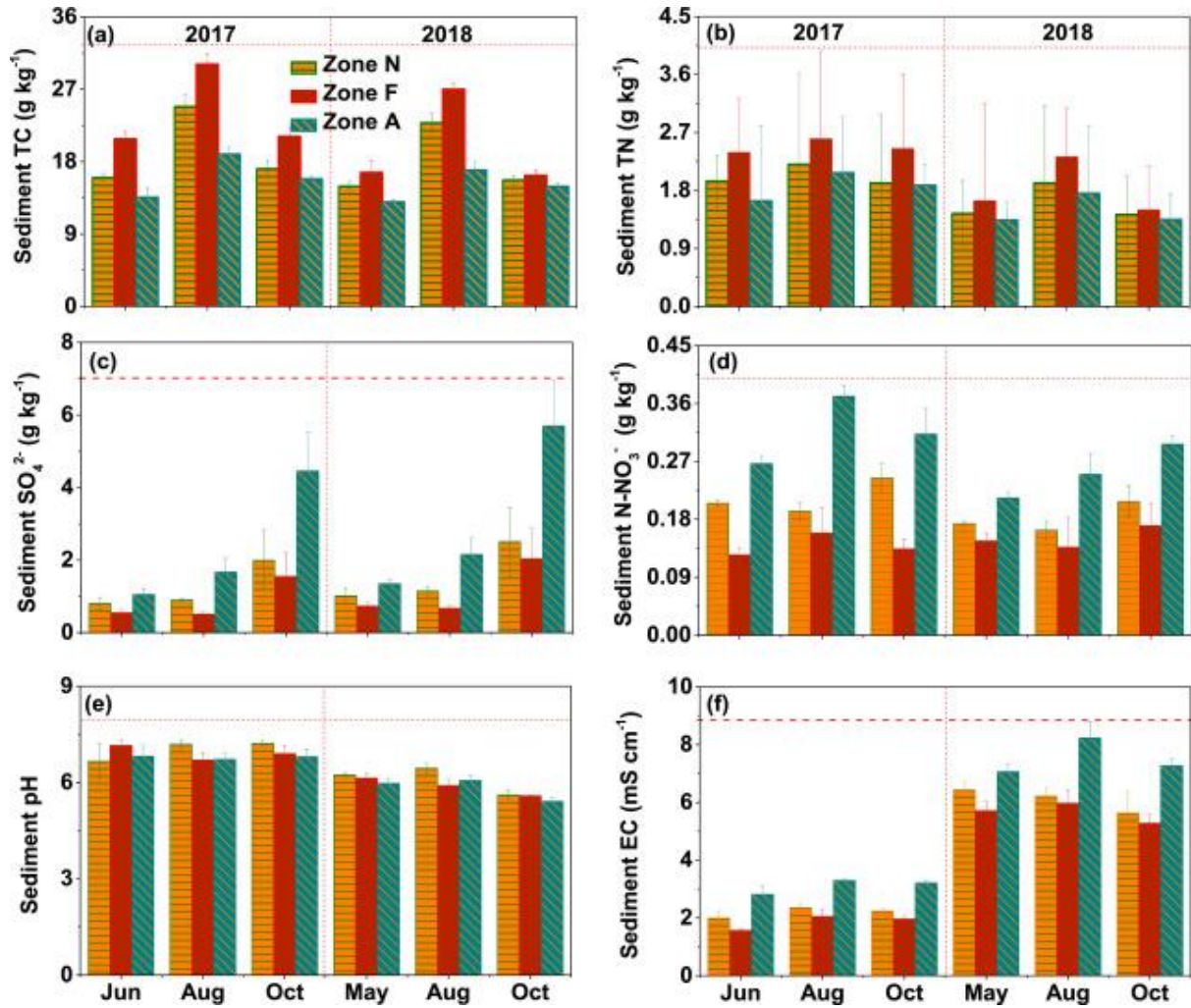


Fig. 2. Variations in (a) TC, (b) TN, (c) SO_4^{2-} , (d) N-NO_3^- , (e) pH, and (f) EC in the surface sediment (15 cm depth) in the three zones of aquaculture ponds in the Min River estuary during the aquaculture period. Zones N, F and A are nearshore area, feeding area and aeration area, respectively. Bars represent mean ± 1 SE ($n = 3$).

TC and TN exhibited similar temporal variations over the two years, with higher and lower concentrations being found at the middle (August) and initial stages (May or June), respectively (Fig. 2a and b). The sediment SO_4^{2-} content in the aquaculture ponds increased with time (Fig. 2c). The seasonal means of sediment TC, TN, N-NO_3^- and pH in 2017 were slightly higher than those in 2018 ($p > 0.05$), while the mean of sediment EC in 2017 was significantly lower than that in 2018 ($p < 0.05$).

Gas transfer velocities (k_x) showed no clear seasonal patterns but varied significantly among measurement events (Fig. S1b). Across the sampling sites, k_x values ranged from 0.7 to 5.7 cm h^{-1} and from 0.5 to 3.0 cm h^{-1} in 2017 and 2018, respectively (Fig. S1b). The seasonal averages of k_x were 2.6 ± 0.5 and 1.6 ± 0.2 cm h^{-1} in 2017 and 2018, respectively.

3.2. Dissolved CH_4 concentrations and sediment CH_4 production rates

Dissolved CH₄ concentrations showed large spatiotemporal variations within the ponds. The average dissolved CH₄ concentrations during the aquaculture period ranged from 2.29±0.29 to 50.48±20.91 µM in 2017 (unpublished data), which were higher than those in 2018 (0.18±0.03–1.68±0.17 µM) (Fig. S2). In general, dissolved CH₄ concentrations were lowest in May/June, and highest in August/ September (Fig. S2). Over the two years, the mean dissolved CH₄ concentration was highest at Zone F (16.8 µM), followed by Zone N (4.2 µM) and Zone A (2.2 µM).

Sediment CH₄ production rates in the shrimp ponds ranged from 1.09 to 6.34 µg g⁻¹ d⁻¹ over the aquaculture period in 2017, which were higher than those in 2018 (0.03–0.53 µg g⁻¹ d⁻¹, unpublished data). CH₄ production rates varied significantly among the three aquaculture stages in 2017 in the following descending order ($p<0.01$; Table 1): middle stage (August) > final stage (October) > initial stage (June) (Fig. 3). Among the three sampling zones, the highest and lowest CH₄ production rates were found in Zone F (2.51±0.25 – 5.64±0.63 µg g⁻¹ d⁻¹) and Zone A (1.17±0.04 – 2.31±0.31 µg g⁻¹ d⁻¹), respectively ($p<0.01$; Fig. 3; Table 1).

Table 1. Summary of two-way ANOVAs (with ponds ID specified as the random term) that examining the effect of sampling zones, sampling time (or stage) and their interactions on ebullitive CH₄ Flux (or sediment CH₄ production rates) at the aquaculture ponds in the Min River estuary.

Fixed effect	Ebullitive CH ₄ Flux					Sediment CH ₄ production rates				
	Sum of squares	Mean square	df	F values	P values	Sum of squares	Mean square	df	F values	P values
Zones	21,776.744	10,888.372	2	43.678	0.002	19.332	9.666	2	270.856	<0.001
Time	115,379.591	3978.607	39	3.222	<0.001	22.655	11.328	2	21.990	0.007
Zones × Time	34,418.041	593.414	58	1.084	0.338	3.981	0.995	4	8.031	0.007
Residuals	98,495.369	547.196				5.324	0.296			
Random effect										
Ponds			2	0.331	0.720			2	2.494	0.246

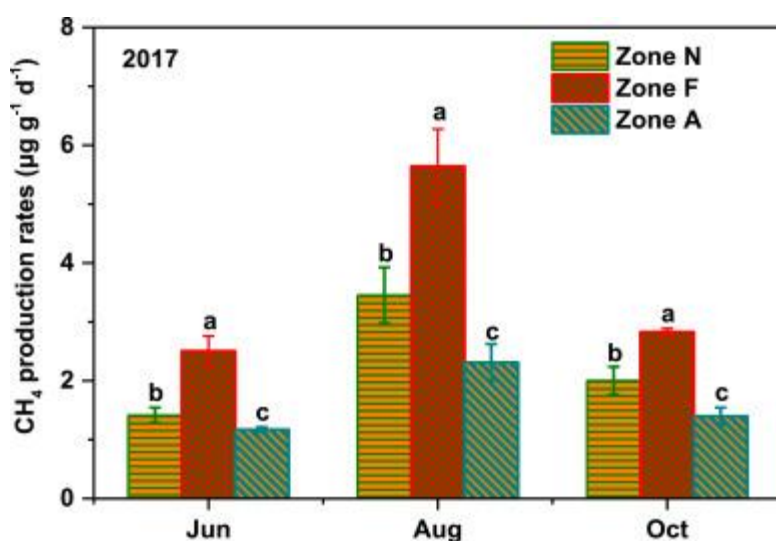


Fig. 3. Variations in sediment CH₄ production rates (0–15 cm depth) in the three zones of aquaculture ponds in the Min River estuary across the three aquaculture stages during the

aquaculture period in 2017. Zones N, F and A are nearshore area, feeding area and aeration area, respectively. Bars represent mean \pm 1 SE ($n = 3$). Different lowercase letters above the bars indicate significant differences ($p < 0.05$) between sampling zones during each culture stage.

3.3. Spatiotemporal variations in ebullitive CH₄ flux within ponds

Over the two years, ebullitive CH₄ fluxes differed significantly among the three zones within the ponds ($p < 0.01$; Table 1), with the ranges of 0.17–77.14, 0.67–123.34, and 0.04–53.40 mg m⁻² h⁻¹ in Zones N, F and A, respectively (Fig. 4). When averaging the monthly fluxes, a strong spatial pattern in ebullitive CH₄ emissions emerged (Figure S3) with the rates decreasing in the order: Zone F > Zone N > Zone A.

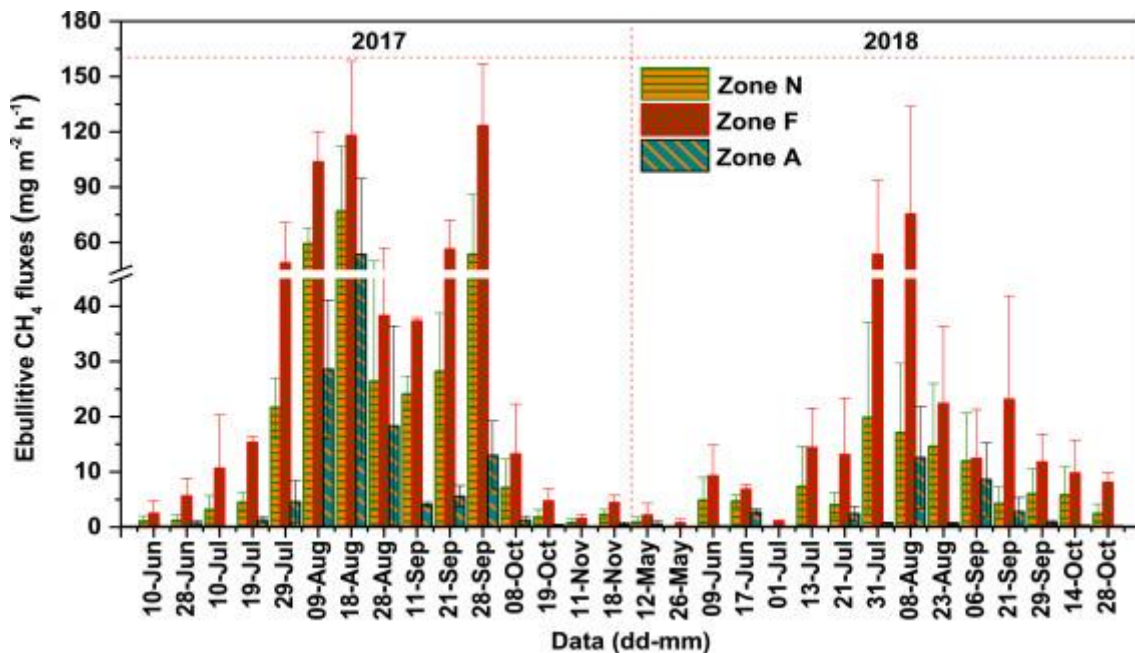


Fig. 4. Spatiotemporal variations in ebullitive CH₄ fluxes from aquaculture ponds in the Min River estuary during the aquaculture period. Zones N, F and A are nearshore area, feeding area and aeration area, respectively. Bars represent mean \pm 1 SE ($n = 3$).

Ebullitive CH₄ fluxes also showed significant variations over time ($p < 0.01$; Table 1), with lower fluxes in May/June and October/November, and higher fluxes in August/September (Fig. 4). Across all the sampling sites, the seasonal average of ebullitive CH₄ fluxes ranged from 0.85 \pm 0.04 to 82.89 \pm 18.90 mg m⁻² h⁻¹ over the aquaculture period in 2017, which was higher than those in 2018 (0.35 \pm 0.22 – 35.06 \pm 20.23 mg m⁻² h⁻¹) ($p < 0.01$).

3.4. Diffusive and ebullitive fluxes

During the aquaculture period in 2017, total CH₄ fluxes in Zones N, F, and A varied in the ranges of 1.16–78.75, 2.31–125.63, and 0.22–54.9 mg m⁻² h⁻¹, respectively. On average, ebullition accounted for 95%, 97% and 88% of the total CH₄ emissions in Zones N, F, and A, respectively, with a minor contribution from diffusion (Fig. 5 and Table 2).

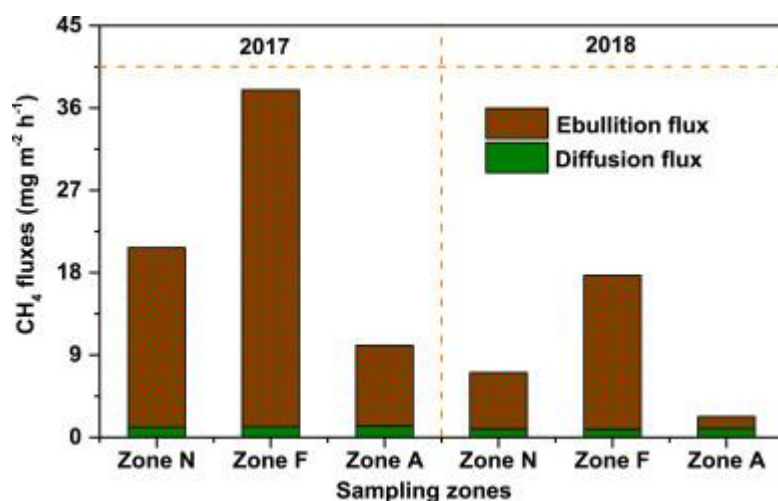


Fig. 5. CH₄ ebullition fluxes vs diffusion fluxes in the three zones of aquaculture ponds in the Min River estuary during the culture period in 2017 and 2018. Zones N, F, and A are nearshore area, feeding area, and aeration area, respectively.

Table 2. Contribution of ebullitive CH₄ fluxes to the total CH₄ fluxes (Mean ± SE, mg m⁻² h⁻¹) across the different zones within ponds during the aquaculture period in 2017 and 2018. Zones N, F, and A are nearshore area, feeding area, and aeration area, respectively.

	2017				2018			
	Zone N	Zone F	Zone A	Average	Zone N	Zone F	Zone A	Average
Total fluxes	20.71±6.50	37.96±11.43	10.04±3.87	22.91±4.79	7.09±1.59	17.70±5.30	2.28±0.94	9.03±2.07
Ebullition fluxes	19.59±6.34	36.81±3.83	8.81±3.83	21.74±4.76	6.99±1.59	16.81±5.29	1.32±0.93	8.09±4.57
Contribution of Ebullition (%)	95	97	88	95	99	95	58	90

During the aquaculture period in 2018, total CH₄ fluxes also differed greatly among the three zones with the ranges of 0.25–19.92, 0.85–75.58, and 0.16–12.84 mg m⁻² h⁻¹ in Zones N, F, and A, respectively. On average, ebullition accounted for 99%, 95% and 58% of the total CH₄ emissions in Zones N, F, and A, respectively (Fig. 5 and Table 2). When averaging the fluxes obtained at all sampling sites over the two years, ebullition contributed over 90% to the total CH₄ emission from ponds.

3.5. Relationships between ebullitive CH₄ flux and environmental variables

The results of Spearman correlation analysis showed that ebullitive CH₄ fluxes in the three zones of our ponds were positively correlated with sediment TC ($r=0.36-0.82$, $p<0.05$; Fig. S4) and TN content ($r=0.56-0.88$, $p<0.05$; Fig. S5), and negatively correlated with sediment SO₄²⁻ content ($r=0.41-0.78$, $p<0.05$; Fig. S6). Ebullitive CH₄ fluxes were also positively correlated with sediment CH₄ production rates across the sites on each sampling campaign ($r=0.58-0.85$, $p<0.01$; Fig. S7).

When data from the six sampling campaigns were analyzed together, ebullitive CH₄ fluxes were found to be positively correlated with sediment temperature ($r=0.49$, $p<0.01$), TC ($r=0.76$, $p<0.01$) and TN ($r=0.59$, $p<0.01$), and negatively correlated with water DO concentration ($r=-0.55$, $p<0.01$) and sediment SO₄²⁻ content ($r=-0.31$, $p<0.01$) (Table S2).

Our PCA results showed that the two principal components, i.e. PC1 and PC2, explained 95.6% and 3.7%, respectively, of the total variance in ebullitive CH₄ fluxes (Fig. 6). Ebullitive CH₄ efflux, sediment temperature, TC, TN and pH all displayed positive PC1 loading, while sediment SO₄²⁻, N-NO₃⁻ and EC showed negative PC1 loading (Fig. 6).

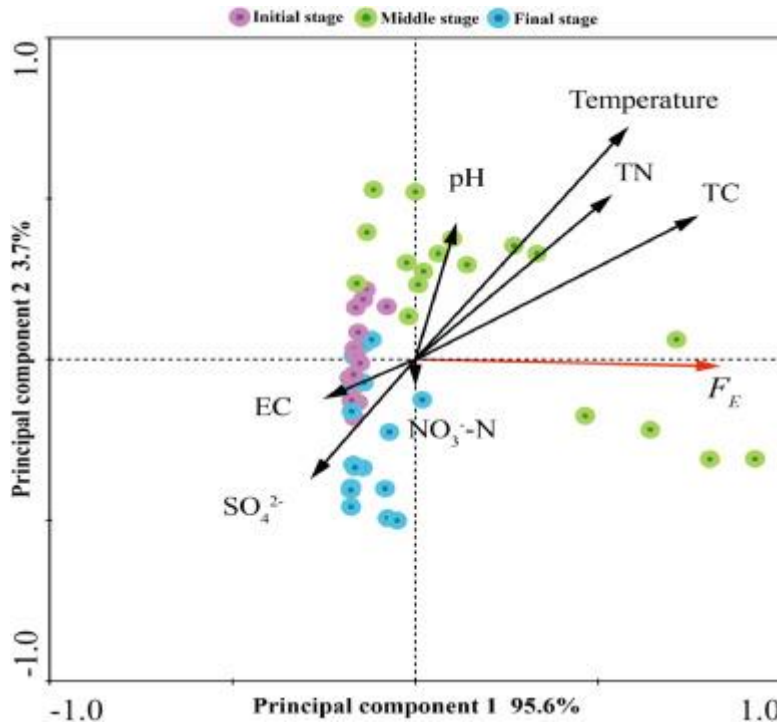


Fig. 6. The principal component analysis (PCA) biplots of the ebullitive CH₄ fluxes (F_E) and surface sediment characteristics of the aquaculture ponds, showing the loadings of environmental factors (arrows) and the scores of observations in three aquaculture stages (initial, middle and final stages).

4. Discussion

4.1. High CH₄ emissions largely attributed to ebullition

Previous studies have found that CH₄ ebullition plays a crucial role in influencing the magnitude of total CH₄ emissions from inland aquatic ecosystems (e.g., Bastviken et al., 2004, 2011; Wu et al., 2019; Zhu et al., 2016). The contributions of ebullition to total CH₄ emissions showed remarkable variations between different inland waters, ranging from 10% to 95% (e.g., Baulch et al., 2011; Deshmukh et al., 2016; Sawakuchi et al., 2014; Wu et al., 2019). For organic-rich shallow zones or freshwater systems, ebullitive fluxes were responsible for more than 50% of the total CH₄ emissions (e.g., Bastviken et al., 2008; Sawakuchi et al., 2014; Zhu et al., 2016). The extremely high CH₄ ebullitions from these aquatic systems are mainly attributed to the relatively shallow water depth and low hydrostatic pressure, which are more favorable for bubble formation and release in sediments (Casper et al., 2000; DelSontro et al., 2011; Zhu et al., 2016). Across the two years, the average total CH₄ emission (including both diffusive and ebullitive fluxes) from the shrimp ponds was 16.0 mg m⁻² h⁻¹, which was equivalent to an annual emission of 69.5 g m⁻² yr⁻¹ (only considering the aquaculture period). Ebullitive fluxes were responsible for

more than 90% of the total CH₄ emissions over the two years. Both ebullitive CH₄ fluxes and total CH₄ emissions showed substantial inter-annual variations (Figs. 4 and 5), with lower rates in 2018 and higher values in 2017 (Table 2). Our results highlighted the significant contribution of ebullition to total CH₄ emissions from aquaculture ponds even in different years.

The total CH₄ flux in our estuarine mariculture ponds was lower than that reported in Lake Medo at the eastern Tibetan Plateau (Zhu et al., 2016), a eutrophic reservoir in Ohio, USA (Beaulieu et al., 2018), and the Gold Creek Reservoir in Australia (Sturm et al., 2014) (Table S3). However, the total CH₄ emission in our mariculture ponds was fairly comparable to the results in Lake Thimmapuram in south India (Attermeyer et al., 2016) and the Lixiahe River in southeast China (Wu et al., 2019). Moreover, the magnitude of total CH₄ fluxes in our ponds was one to two orders higher than that observed in various lakes in different climatic regions (e.g., Huttunen et al., 2001; Natchimuthu et al., 2016; Walter et al., 2008; Wik et al., 2013). The net CH₄ emission rates in our mariculture ponds were also substantially higher than those in a hydropower reservoir in Switzerland (Delsontro et al., 2010), several rivers in the Amazon (Sawakuchi et al., 2014), as well as many Chinese lakes (e.g., Chen et al., 2009a; Li et al., 2018; Wang et al., 2006; Xiao et al., 2017; Xing et al., 2005), reservoirs (e.g., Chen et al., 2009b; Li et al., 2018; Wang et al., 2017; Zhao et al., 2013; Zheng et al., 2011), and freshwater aquaculture ponds (e.g., Chen et al., 2016; Hu et al., 2014, 2016; Liu et al., 2015; Ma et al., 2018; Wu et al., 2018). In addition, the total CH₄ emission from our ponds during the aquaculture period was higher than the average of 1.4 mg m⁻² h⁻¹ in China's natural wetlands (Wei and Wang, 2017), and was approximately 6 times higher than the average of 2.6 mg m⁻² h⁻¹ in the *Cyperus malaccensis* marsh from an adjacent estuary (Yang et al., 2019b). As seen in Table S3, ebullitive fluxes generally accounted for more than 80% of the total CH₄ emission in most aquatic ecosystems. Hence, the much higher total CH₄ fluxes observed in our mariculture ponds, compared to many other aquatic ecosystems, might imply the presence of a considerably high rate of CH₄ ebullition.

4.2. Feeding areas as hotspots of ebullitive CH₄ emission

Ebullitive CH₄ emission in aquaculture ponds at a high spatial resolution remains poorly characterized. As the first attempt to investigate the fine-scale spatial variations in CH₄ ebullition in aquaculture ponds, this study showed that the mean ebullitive CH₄ efflux at Zone F was higher than those in Zones A and N by over 5 and 2 times, respectively (Fig. S3 and Table 2). Our results highlighted the strong spatial variations in ebullitive CH₄ emission within the ponds, with the feeding areas being the ebullition hotspot areas. The extremely high rate of CH₄ ebullition in Zone F could be primarily ascribed to the large organic matter loading in sediment that enhanced CH₄ production. Over the two-year period, the mean sediment TC concentration in Zone F was higher than those in Zones N and A by 12–24% and 33–49%, respectively. Mean sediment TN concentration in Zone F was also higher than those in Zones N and A by 14–23% and 23–33%, respectively. Previous studies have shown that CH₄ in aquatic ecosystems is mainly generated from the organic-rich sediments (e.g., Bastviken et al., 2008; Grinham et al., 2018b; Xiao et al., 2017; Zhou et al., 2019). Organic matter in sediments provides the key carbon substrates for methanogenic activities, thereby providing favorable conditions for CH₄ production and bubble formation (Bastviken et al., 2008; Davidson et al., 2018; Walter et al., 2008; Zhu et al., 2016). The higher sediment organic C and N loads in Zone F would have contributed to larger CH₄ production (Fig. 3), and consequently larger CH₄ ebullition as compared to Zones A and

N. This was further confirmed by the significant and positive relationships observed (1) between ebullitive CH₄ fluxes and sediment CH₄ production rates (Fig. S7), and (2) between ebullitive CH₄ fluxes (or sediment CH₄ production rates) with sediment TC (Table S2, Fig. S4 and Fig. S8b) and TN (Table S2 and Fig. S5) concentrations.

Net CH₄ release from the aquatic systems to the atmosphere is determined by the processes of CH₄ production by methanogens, CH₄ oxidation by methanotrophs, and CH₄ transport. CH₄ oxidation can effectively reduce net CH₄ emissions to the atmosphere (Bastviken et al., 2008), and is governed by the DO level in the water column (Sundh et al., 2005; Yang et al., 2019b). CH₄ oxidation rates in the water column have been found to increase with the concentrations of dissolved CH₄ and DO (Bastviken et al., 2008; Matoušů et al., 2017; Yang et al., 2019c). Aquaculture ponds are aerated on a daily basis in Zone A (Yang et al., 2019a), which can improve the oxygen supply to the sediment surface and reduce the development of anaerobic conditions, ultimately leading to an inhibition of CH₄ production but an enhancement of CH₄ oxidation (Liu et al., 2015; Schrier-Uijl et al., 2011; Yang et al., 2019a). Although CH₄ oxidation rates in the present study were unavailable, our observation of the significant and negative relationship between sediment CH₄ production rates and water DO concentrations ($r=0.70$, $p<0.01$; Table S2) supported the influence of oxygen on methanogenesis. Furthermore, the aeration-induced physical disturbance of pond water might lead to the burst of CH₄ bubbles, which in turn promote the oxidation of a large proportion of the produced CH₄ in the water column before its release into the atmosphere. Therefore, aeration activity was considered as an important contributor to the lower ebullitive CH₄ fluxes detected in Zone A.

4.3. Temporal variations in ebullitive CH₄ fluxes

Ebullitive CH₄ flux has been reported to vary temporally in inland waters (e.g., Natchimuthu et al., 2014; Wik et al., 2013; Wu et al., 2019). In our study, ebullitive CH₄ emissions from the estuarine aquaculture ponds also showed considerable variations between different seasons (Fig. 4 and Table 1). The seasonal mean ebullitive CH₄ emissions in 2017 were positively correlated with sediment CH₄ production rates (0–15 cm depth) ($r=0.79$, $p<0.01$). As such, the seasonal patterns of ebullitive CH₄ emissions were largely similar to those of sediment CH₄ production rates (0–15 cm depth) (Fig. 3). A high CH₄ production rate in sediments would increase the CH₄ concentrations in sediment porewater and subsequently the production and release of CH₄ bubbles to the atmosphere. Previous studies found that the seasonal patterns of CH₄ metabolism are governed by seasonal variations in temperature which drive the production of substrate precursors and microbial activity (Crawford et al., 2014; Rosentreter et al., 2017; Vizza et al., 2017). Our Spearman correlation (Table S2) and PCA (Fig. 6) results suggested that the seasonal pattern of ebullitive CH₄ fluxes (or CH₄ sediment production rates) generally followed the patterns of sediment temperature, TC and TN, but were opposite to the pattern of sediment SO₄²⁻. Therefore, the lower ebullitive CH₄ fluxes in the initial and final stages of aquaculture were primarily ascribed to low organic matter availability, high SO₄²⁻ in sediments and low water temperature, which reduced the substrate availability and metabolic activity of methanogens (Kelly and Chynoweth, 1981; Sun et al., 2013; Vizza et al., 2017). In contrast, high ebullitive CH₄ emissions observed in the middle stages of aquaculture were influenced, to a large extent, by the interactions between high sediment temperature, large organic matter load, and low sediment SO₄²⁻ content, which favored methanogenic activities (Davidson et al., 2018; Sun et al., 2013; Vizza et al., 2017). Further studies should be done to explore the exact

effects of microbial abundance and activity on CH₄ production and emission in aquaculture ponds.

We also observed substantial interannual variations in ebullitive CH₄ fluxes in this study (Fig. 4 and Table 2). The mean ebullitive CH₄ flux over the whole aquaculture period was $21.74 \pm 4.76 \text{ mg m}^{-2} \text{ h}^{-1}$ in 2017, which was significantly higher than that in 2018 ($8.09 \pm 4.57 \text{ mg m}^{-2} \text{ h}^{-1}$) ($p < 0.01$; Table 2). The interannual variations in ebullitive CH₄ emissions from the aquaculture ponds could possibly be ascribed to the variations in mean sediment temperature (25.0°C in 2017 vs 22.9°C in 2018), TC (19.8 vs 17.5 g kg^{-1}), and TN (2.1 vs 1.6 g kg^{-1}) between the two years, which was in line with the previous discussion regarding the effect of temperature and substrate availability on CH₄ metabolism. Furthermore, the mean sediment SO₄²⁻ content (1.4 g kg^{-1}) and EC (2.4 mS cm^{-1}) over the aquaculture period in 2017 were much lower than those in 2018 (SO₄²⁻: 1.9 g kg^{-1} ; EC: 6.4 mS cm^{-1}), as a result of freshwater dilution caused by the combination of high precipitation and large surface runoff. High salinity has been shown to support sulfate reduction that could lead to enhanced anaerobic CH₄ oxidation in sediments and intensified competition between sulfate reducing bacteria and methanogens (Chambers et al., 2013; van der Gon et al., 2001; Vizza et al., 2017). Subsequently, CH₄ fluxes decreased with the increase of SO₄²⁻ and salinity levels, as reported in coastal wetlands (Poffenbarger et al., 2011; Vizza et al., 2017; Wilson et al., 2015) and waters (Welti et al., 2017; Yang et al., 2018). The much lower sediment SO₄²⁻ content and salinity in 2017 (Fig. 2c and f) could have promoted methanogenic activities markedly, as supported by the higher sediment CH₄ production rates in 2017 than in 2018 ($1.09\text{--}6.34$ vs $0.03\text{--}0.53 \text{ } \mu\text{g g}^{-1} \text{ d}^{-1}$), which in turn enhanced CH₄ ebullition.

4.4. Implications for ebullitive CH₄ flux from aquaculture ponds

The seasonal integrated ebullitive CH₄ fluxes from the aquaculture ponds during the culture period in 2017 and 2018 were found to be $0.4\text{--}40.4 \text{ g CH}_4 \text{ m}^{-2}$ in Zone N, $1.1\text{--}64.5 \text{ g CH}_4 \text{ m}^{-2}$ in Zone F, and $0.2\text{--}24.9 \text{ g CH}_4 \text{ m}^{-2}$ in Zone A. In our ponds, Zones A, F and N covered 65%, 25%, and 10% of the total surface area, respectively. Zones F and N had a disproportionately high impact on CH₄ ebullition, contributing to 49.7–71.8% and 20.1–33.6% of the spatially averaged ebullitive fluxes (Fig. 7). Our results highlighted the importance of considering the spatial variations in ebullitive CH₄ emission within ponds during the upscaling of small-scale CH₄ fluxes to the whole pond level (Fig. 7).

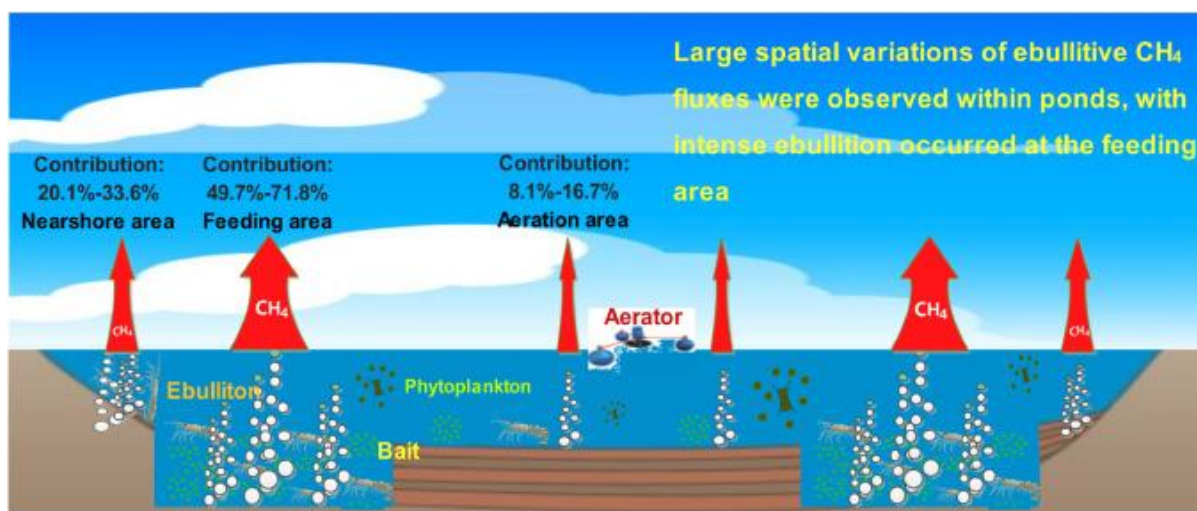


Fig. 7. Conceptual diagram illustrating the spatial variations in ebullitive CH_4 fluxes within aquaculture pond during the aquaculture period.

Due to the growing global demand for seafood, an increasing number of aquaculture ponds have been built and the total area of aquaculture ponds has increased continually over the last decades in the coastal areas in China and many other countries (Ren et al., 2019; Yang et al., 2020a). Thus, it is an imminent challenge to balance seafood production and greenhouse gas mitigation for the sustainable development of aquaculture sector (Yang, 2014). Our results indicated that high ebullitive CH_4 emission occurred in the feeding zone with high residue bait accumulation, while low CH_4 fluxes were maintained at the aeration zone (Fig. S3 and Fig. 7). Given the observed large within-pond variations in CH_4 flux, reducing the built-up of unconsumed feeds and increasing aeration are important strategies for mitigating CH_4 emissions from the mariculture ponds.

Some efforts have been made to estimate large-scale CH_4 emissions from aquaculture systems by extrapolating short-term field measurements (< 1 year) to longer periods in a given area (e.g., Hu et al., 2016; Yang et al., 2018; Yuan et al., 2019). However, our results showed that ebullitive CH_4 emissions from the estuarine aquaculture ponds had strong inter-annual variation, with the average fluxes differing by more than two-fold between the two years, which implied a large uncertainty of CH_4 flux estimation based on short-term measurements. Therefore, it is crucial to make long-term observations in order to minimize the potential bias of CH_4 flux estimates and improve our understanding of CH_4 budget. Furthermore, ebullitive CH_4 emissions from the estuarine aquaculture ponds varied considerably between different months (Fig. 4), with the coefficients of variation ranging between 110%–117%. These findings indicated the potential problem of extrapolating flux estimates from a single month to seasonal or annual scales, exacerbating the uncertainty of CH_4 emission estimates. Therefore, it is suggested that flux measurements should be conducted in as many sites as practicable over multiple months in order to achieve a more accurate CH_4 flux estimate and enhance our understanding of CH_4 dynamics in aquaculture ponds.

4.5. Limitation and future research

Similar to many studies, there are some limitations in the current study. First, ebullitive CH_4 fluxes were determined in the aquaculture ponds in MRE, southeast China only. Care

should be taken in upscaling our ebullitive CH₄ fluxes to broader spatial scales. To further improve the accuracy of estimates of regional or global ebullitive CH₄ emissions from mariculture ponds, more works on the effects of other factors, such as regional climate, aquaculture types, and management practice, on CH₄ flux are needed. Second, while sediment TC and TN were analyzed in the current study, other chemical and biological variables, such as autochthonous and allochthonous dissolved organic matter (Zhou et al., 2019), and the abundance and activity of methanogens and methanotrophs, can also influence the spatiotemporal variations in ebullitive CH₄ emission in aquatic ecosystems. Therefore, future studies should further explore the influence of a wider range of physio-chemical and biological variables on CH₄ ebullition. Third, previous studies have shown that CH₄ ebullition events were often coincided with low atmospheric pressure (Mattson and Likens, 1990; Tokida et al., 2007; Zhu et al., 2016). Tropical cyclones are extreme perturbations that can cause a rapid drop in atmospheric pressure, which in turn can trigger substantial CH₄ ebullition in tropical coastal areas. Therefore, more direct measurement of CH₄ ebullition should be done during the period of typhoon passage in coastal mariculture ponds (Yang et al., 2020b). Fourth, direct measurement of the diel pattern of ebullitive CH₄ fluxes was unavailable in the current study, although diurnal variations in CH₄ emission have been reported in lakes (e.g., Erkkilä et al., 2018; Hirota et al., 2007; Xing et al., 2004). Future research should quantify the diel variations in ebullitive CH₄ fluxes from aquaculture ponds. Fifth, our floating chambers might fail to completely capture all the ebullition events, leading to a gross under-estimation of annual CH₄ emission. In future, more advanced measurement techniques should be applied to more accurately measure the ebullitive CH₄ emission from aquaculture ponds. Lastly, our study did not consider the influence of water velocity in the estimation of diffusive CH₄ fluxes from the aquaculture ponds. A gas transfer coefficient model that consider both wind speed and water velocity could be used in further studies to improve the accuracy of diffusive CH₄ flux estimates in aquatic ecosystems.

5. Conclusions

This study was the first to characterize the spatiotemporal variations in ebullitive CH₄ fluxes within aquaculture ponds. Our results showed that ebullitive CH₄ emissions contributed to over 90% of the total CH₄ emission from aquaculture ponds, indicating the importance of ebullition in transporting CH₄ from the ponds to the atmosphere. Ebullitive CH₄ fluxes varied significantly between the two measurement years and within the ponds. Intense CH₄ ebullition from the ponds occurred in the middle stage of aquaculture (July to September), which could be attributed to the large organic matter loading and high temperature during this period in the subtropical estuarine environment. The feeding area was a hotspot of CH₄ ebullition, contributing 49.7–71.8% to the spatially averaged ebullitive fluxes. Our results showed that flux measurements based on a single point in space or time would introduce marked biases in estimating whole-pond ebullitive CH₄ emissions. Field measurements of CH₄ ebullition should be done in as many sites as practicable over a longer term in order to produce more accurate estimates of ebullitive CH₄ fluxes at the whole-pond or even larger scales.

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