

*Rapid development of a salivary calculus
in submandibular gland and its potential
causes in a young victim following
Russell's viper bite*

Article

Accepted Version

Creative Commons: Attribution-Noncommercial-No Derivative Works 4.0

Arathisenthil, S. V., Senthilkumaran, S., Vijayakumar, P., Savania, R., Williams, H. F., Elangovan, N., Bicknell, A. B., Patel, K., Trim, S. A., Thirumalaikolundusubramanian, P. and Vaiyapuri, S. ORCID: <https://orcid.org/0000-0002-6006-6517> (2022) Rapid development of a salivary calculus in submandibular gland and its potential causes in a young victim following Russell's viper bite. *Toxicon*, 206. pp. 85-89. ISSN 0041-0101 doi: 10.1016/j.toxicon.2021.12.018 Available at <https://centaur.reading.ac.uk/102088/>

It is advisable to refer to the publisher's version if you intend to cite from the work. See [Guidance on citing](#).

To link to this article DOI: <http://dx.doi.org/10.1016/j.toxicon.2021.12.018>

Publisher: Elsevier

All outputs in CentAUR are protected by Intellectual Property Rights law, including copyright law. Copyright and IPR is retained by the creators or other

copyright holders. Terms and conditions for use of this material are defined in the [End User Agreement](#).

www.reading.ac.uk/centaur

CentAUR

Central Archive at the University of Reading

Reading's research outputs online

1 **Rapid development of a salivary calculus in submandibular gland and its potential causes**
2 **in a young victim following Russell's viper bite**

3 S.V. Arathisenthil^{1*}, Subramanian Senthilkumaran^{1*}, Pradeep Vijayakumar^{2*}, Ravi Savania², Harry
4 F. Williams³, Namasivayam Elangovan⁴, Andrew B. Bicknell⁵, Ketan Patel⁵, Steven A. Trim⁶, Ponniah
5 Thirumalaikolundusubramanian^{7,8} and Sakthivel Vaiyapuri^{2§}

6 ¹Manian Medical Centre, Erode, Tamil Nadu, India

7 ²School of Pharmacy, University of Reading, Reading, UK

8 ³Toxiven Biotech Private Limited, Coimbatore, Tamil Nadu, India

9 ⁴Department of Biotechnology, School of Biosciences, Periyar University, Salem, Tamil Nadu, India

10 ⁵School of Biological Sciences, University of Reading, Reading, UK

11 ⁶Venomtech Limited, Sandwich, UK

12 ⁷Trichy SRM Medical College Hospital & Research Centre, Trichy, Tamil Nadu, India

13 ⁸The Tamil Nadu Dr MGR Medical University, Chennai, Tamil Nadu, India.

14
15 *These authors contributed equally.

16
17 §Correspondence to: s.vaiyapuri@reading.ac.uk

18
19 **Abstract**

20 Russell's viper bites are known to cause a range of haemotoxic, neurotoxic, myotoxic,
21 cytotoxic and nephrotoxic complications. However, the impact of Russell's viper bites as well as bites
22 from other venomous snakes on sialolithiasis has not been previously reported. Here, we present
23 an interesting case where a Russell's viper bite induced the rapid development of a calculus in
24 submandibular gland in a 10-year-old boy. Upon admission, the victim did not show any symptoms
25 of swelling and/or pain around his oral cavity. He received antivenom treatment to normalise his
26 coagulation parameters, however, on day three he developed swelling and extreme pain around his
27 right mandibular region. An ultrasound investigation revealed the presence of a calculus in his
28 submandibular gland, which was removed using a minor surgical procedure. The histopathological
29 examination revealed this as a poorly calcified salivary calculus, which is composed of cell debris,
30 mucopolysaccharides and lipids. The mechanisms behind its rapid development following a
31 snakebite are unclear although this could be linked to excessive inflammation or modifications to the
32 composition of saliva induced by venom toxins or other unknown factors. This report reveals an
33 unusual complication induced by a Russell's viper bite and alerts clinicians who treat snakebites to
34 be aware of such envenomation effects. Moreover, this will lead to novel research to explore the
35 relationship between venom toxins and functions of salivary glands.

36 **Key words**

37 Snakebite envenomation; Russell's viper; salivary calculi; salivary glands; sialolithiasis; inflammation

38 **Introduction**

39 Sialolithiasis is the formation of calculi in salivary glands, primarily in ducts connecting the
40 glands to the oral cavity (Delli et al., 2014; Hammett and Walker, 2021; Kraaij et al., 2014). This
41 occurs predominantly in adults (mostly in males) who are aged between 30 and 60 years old with
42 occurrence among children being rare (Kraaij et al., 2014). Between the three pairs of salivary glands
43 (parotid, submandibular, and sublingual), calculi formation has been predominantly (over 85%
44 cases) reported in submandibular glands (Grases et al., 2003; Kraaij et al., 2014), mostly in the
45 excretory ducts (Sigismund et al., 2015). There are no precise mechanisms or causative factors
46 established yet for the development of salivary calculi although they are broadly linked to various
47 anatomical features of salivary glands and composition of calculi (Grases et al., 2003). For example,
48 accumulation of bacteria, foreign bodies (Marchal et al., 2001), cellular debris (Harrison, 2009) and
49 excessive inflammation, which triggers the infiltration of neutrophils (Schapher et al., 2020) in ducts

50 are likely to result in sialolithiasis. The presence of micro calculi (concrements) in the excretory duct
51 system in submandibular glands even in many healthy, asymptomatic individuals (Sigismund et al.,
52 2015) may act as nuclei for the deposition of various organic and inorganic molecules from saliva to
53 ultimately form large calculi over a long period (Grases et al., 2003; Sivasubramaniam et al., 2015).
54 Moreover, ducts in submandibular glands allow a slower salivary flow which may be affected when
55 the salivary thickness is increased leading to the development of sludge and ultimately calculi.
56 Similarly, increased ionic contents (specifically calcium) and modified amylase activities may induce
57 the formation of salivary calculi (Kraaij et al., 2014; Schröder et al., 2017).

58 Snakebite envenomation (SBE) has been reported to cause diverse neurotoxic, haemotoxic,
59 myotoxic, cytotoxic and nephrotoxic complications based on the type of offending snake and the
60 nature of their venom (Gutiérrez et al., 2017; Williams et al., 2019). Russell's viper (*Daboia russelii*)
61 is a key species that causes numerous bites and resulting deaths and disabilities in India (Samuel
62 et al., 2020; Vaiyapuri et al., 2013). While viper venoms generally induce haemotoxic effects,
63 Russell's viper venom causes both haemotoxic and neurotoxic effects (Silva et al., 2017; Silva et al.,
64 2016). In addition, Russell's viper bites are known to induce rare complications such as priapism
65 (Senthilkumaran et al., 2021) and bilateral blindness (Subasinghe et al., 2014). While Russell's viper
66 bite-induced swelling in parotid glands has been reported in few case reports (Deepak et al., 2013;
67 Sasidaran et al., 2017), the impact of SBE including bites from Russell's viper on salivary calculi
68 formation has not been previously documented. Hence, we report this unusual case of rapid salivary
69 calculus formation in submandibular gland in a 10-year-old boy following a Russell's viper bite. This
70 rare case is not only reporting an unusual complication of Russell's viper bite, but it also opens a
71 new avenue to explore the impacts of venom toxins on the pathophysiology of salivary glands.
72 Moreover, this will create awareness among clinicians who treat SBE to closely monitor the
73 development of this type of rare complications in SBE victims to provide better care.

74 **Case presentation**

75 A 10-year-old boy with no previous medical conditions was brought to our emergency
76 department following a snakebite on his left foot while walking in a field nearly two hours prior to the
77 admittance. A tourniquet had been placed above the bite site using a banana tree fibre by the victim's
78 father. They also brought the offending snake (killed), which was identified as a Russell's viper by a
79 herpetologist (**Figure 1A**). Upon admission, the boy was conscious, well oriented, afebrile, and mildly
80 anxious. Swelling was observed around the bite site although there was no excessive bleeding or
81 tissue damage (**Figure 1B**). His initial vital signs were measured as; blood pressure - 116/72 mm
82 Hg, heart rate - 129 beats/min, respiratory rate - 26 breaths/min, temperature - 36.9°C, and oxygen
83 saturation on room air - 100%. There were no obvious signs of swelling in his lymph nodes or around
84 the oral cavity. He exhibited typical features of Russell's viper bite-induced systemic (gum bleeding),
85 and local (swelling, developing wound and mild bleeding at bite site) envenomation effects. The 20-
86 minute whole blood clotting time was prolonged and therefore, he received 100 mL of polyvalent
87 antivenom (Bharat Serums and Vaccines Limited, India) intravenously according to the standard
88 protocols. The antivenom administration was well tolerated with no signs of adverse reactions. He
89 received another 100 mL of antivenom and four units of fresh frozen plasma to normalise his
90 coagulation parameters over the next twenty-four hours. His other haematologic, metabolic, and
91 biochemical parameters were within the normal limits. He was comfortable on the second day with
92 no obvious local inflammatory changes. However, on the third day (while staying at the hospital), he
93 complained of sudden onset of pain and swelling on the right mandibular region. On bimanual
94 palpation, right submandibular gland was warm and tender with swollen lymph node. Oral
95 examination has revealed a firm, tender, inflamed and erythematous duct opening with hard swelling
96 situated near the right premolars, which was extremely painful on palpation. There was no
97 associated discharge or bleeding from the affected area. The occlusal radiograph indicated the
98 absence of any salivary calculi although an ultrasound scan of his right cheek confirmed the
99 presence of a calculus measuring around 6 mm (**Figure 1C**). His salivary analysis confirmed
100 moderately increased thickness and threads in mucus, and a few epithelial and bacterial cells.
101 However, no crystals or major differences in biochemical parameters (including calcium levels) were
102 noticed in saliva (**Table 1**). Under local anaesthesia, an incision was made at the ductal orifice and
103 the calculus together with surrounding affected tissues was exposed and removed (**Figure 1D**).
104 Absorbable sutures were placed to close the incised area and his post-operative healing was smooth
105 and uneventful. Histopathological examination of salivary gland tissues extracted together with the

106 calculus displayed the infiltration of inflammatory cells and ductal dilation, and the composition of
107 calculus as glycoproteins, cellular debris, lipids and mucopolysaccharides (**Figure 1E**). The boy was
108 discharged on day seven, and his subsequent weekly reviews did not display any further
109 complications.

110 **Case discussion**

111 While SBE is known to induce a diverse spectrum of complications in humans (Williams et
112 al., 2019), to the best of our knowledge, its impact on sialolithiasis has not been previously reported.
113 Moreover, sialolithiasis is commonly encountered among adults who are aged between 30 and 60
114 years old (Grases et al., 2003; Hammett and Walker, 2021). Hence, this report presents an
115 interesting case of SBE (specifically Russell's viper bite)-induced sialolithiasis in a 10-year-old boy
116 with no previous history of any medical conditions. Here, we discuss various potential reasons for
117 sialolithiasis in this patient, although this could have independently developed without any impacts
118 by SBE. Notably in this patient, the salivary calculus has appeared to be formed rapidly following the
119 bite despite antivenom treatment. Similarly, the calculus could have formed already, but the area
120 became very sensitive and extremely painful following the bite. Although occlusal radiography has
121 been used to identify salivary calculi that are rich in calcium (Kim et al., 2016), it is not helpful in
122 identifying poorly calcified calculi as in this case. Hence, it is important to use alternative diagnostic
123 tools such as ultrasound and computed tomography (CT) scans, digital subtraction sialography and
124 sialendoscopy to corroborate the presence of calculi (Goncalves et al., 2017; Hammett and Walker,
125 2021; Kim et al., 2016). Salivary calculi normally form at the rate of 1 mm per year, and they are
126 noticeable only after several years (Grases et al., 2003). However, in this case, the incidence of pain
127 with associated swelling was discovered within three days following the bite. Therefore, further
128 investigation on the physical and biochemical nature of the calculi that are formed acutely (within a
129 short period of time as reported in this case) and chronically (formed over a longer time as reported
130 generally for sialolithiasis) should be thoroughly investigated to determine the mechanisms and
131 major differences between these two methods. While the mechanisms behind the development of
132 salivary calculi in this case are not entirely clear, here we discuss some plausible mechanisms for
133 this incident.

134 The production of saliva and its delivery to the oral cavity is a tightly regulated phenomenon
135 (Delli et al., 2014). Therefore, any adverse changes to the anatomical features of salivary glands
136 and ducts are likely to result in sialolithiasis. For example, inflammation in glands or ducts will lead
137 to infiltration of immune cells (specifically neutrophils) (Hammett and Walker, 2021; Marchal et al.,
138 2001; Schröder et al., 2017), stenosis or dilation and accumulation of minerals, foreign bodies, or
139 cellular debris, which will result in sialolithiasis (Delli et al., 2014; Grases et al., 2003; Harrison, 2009;
140 Kraaij et al., 2014; Marchal et al., 2001). Schapher et al. (Schapher et al., 2020) reported the impact
141 of inflammation in salivary glands by increasing the sudden flow of neutrophils in the ducts leading
142 to release of neutrophil extracellular traps (NETs), which subsequently induced the formation of
143 calculi by acting as a scaffold. Although NETosis is an inflammatory response elicited by neutrophils
144 mainly to remove the invading pathogens and cellular debris, it has been demonstrated to induce
145 the deposition of various organic and inorganic materials in different organs in addition to salivary
146 glands (Maueröder et al., 2015; Muñoz et al., 2019). In the present case, it was evident that
147 inflammatory cells were infiltrated in the ducts of submandibular glands. Moreover, the accumulation
148 of cellular debris, glycoproteins and mucopolysaccharides was noted during pathological
149 examination of extracted calculi. The venom of the saw-scaled viper (*Echis carinatus*) has been
150 reported to induce NETosis (Katkari et al., 2016), although no direct link to sialolithiasis has been
151 established. Russell's viper bites are known to induce inflammatory responses and cytotoxicity due
152 to the large proportion of phospholipase A₂ (PLA₂) present in this venom (Kalita et al., 2018). In
153 addition, the snake venom metalloproteases (Teixeira Cde et al., 2005) and serine proteases
154 (Menaldo et al., 2013) are able to contribute to the development of inflammation. Hence, the
155 components of Russell's viper venom could have induced the inflammation in salivary glands
156 resulting in infiltration of neutrophils and NETosis. The NETs released might have acted as a base
157 for the rapid accumulation of cellular debris, phospholipids and mucopolysaccharides to develop
158 poorly calcified calculus. However, it is unclear on why the toxic venom components have targeted
159 only salivary glands instead of any other organs although the damage to other organs could be
160 minimal and hence unnoticed.

161 The saliva secreted from submandibular glands contains higher concentration of calcium
162 than from other salivary glands (Harrison et al., 1993). The submandibular calculi are formed with
163 over 70% inorganic materials, mostly calcium phosphates in the form of hydroxylapatite (Grases et
164 al., 2003; Hammett and Walker, 2021). The formation of salivary calculi has been demonstrated to
165 occur due to accumulation and deterioration of calcium rich saliva in the ductal area (Grases et al.,
166 2003). It was suggested that an organic molecule or particle forms a base micro calculus upon which
167 other salts, organic and inorganic matters accumulate to form large calculi over a long time (Harrison,
168 2009). Indeed, the presence of micro calculi has been reported even in many healthy individuals and
169 they can slowly induce congestion in ducts and stasis of saliva resulting in inflammation and further
170 growth even in asymptomatic people (Epivatianos and Harrison, 1989). In addition to calcium,
171 phospholipids and secreted glycoproteins will contribute to the development of calculi (Boskey et al.,
172 1981; Boskey et al., 1983; Harrison, 2009). Higher concentrations of bicarbonate in saliva are also
173 able to alter the ionic composition of saliva and induce NETosis (Leppkes et al., 2016). Therefore,
174 all these factors could have contributed to the rapid development of calculus in the present case
175 although the calculus was poorly calcified. Notably, in this patient, there were no major changes in
176 their biochemical parameters in saliva including calcium and bicarbonate levels compared to the
177 reference levels. PLA₂ present in Russell's viper venom causes cytotoxic effects (Chakrabarty and
178 Sarkar, 2015) and these may in turn induced the accumulation of cellular debris and associated
179 components to accumulate around the pre-existing base calculi. Acinar cells and duct cells are the
180 major cellular components of salivary glands. Acinar cells are involved in serous saliva secretion
181 while the duct cells secrete a more mucinous saliva (Kondo et al., 2015). The acinar cells produce
182 an isotonic fluid that is modified into a hypotonic fluid by duct cells before its release into the oral
183 cavity (Kondo et al., 2015). It has been reported that intracellular calcium plays a significant role in
184 the secretion of saliva (Ambudkar, 2016). Indeed, calcium acts as a regulator of various ion pathways
185 involved in salivary secretion. CRISPs (cysteine-rich secretary proteins) that are largely found in
186 venoms of vipers (including Russell's viper) and elapid snakes have been reported to block calcium
187 ion channels in various cell types (Tadokoro et al., 2020). Hence, these non-enzymatic proteins
188 could have affected calcium channels which might have affected the salivary production and its ionic
189 concentrations resulting in increased thickness of saliva. Therefore, studying the impact of CRISPs
190 of Russell's viper venom may demonstrate their influence on ion channels involved in salivary
191 production and their ultimate impact on the formation of salivary calculi. Moreover, an increase in
192 bicarbonate content in saliva precipitates more calcium and phosphate ions leading to formation of
193 calculi (Grases et al., 2003; Hammett and Walker, 2021). Hence, investigating the effects of Russell's
194 viper venom components in modulating bicarbonate synthesis and maintenance will also be useful.
195 Notably, the composition of Russell's viper venom differs based on their geographical locations
196 (Kalita et al., 2018). Therefore, it might be possible for Russell's vipers from specific regions to induce
197 this type of unusual complications in victims especially children.

198 Since Russell's viper bites are known to cause neurological complications (Kalita et al., 2018;
199 Silva et al., 2017), it is important to determine the impact of these venom toxins on neural networks
200 that regulate salivary secretion. Salivary neural networks involve autonomous, parasympathetic, and
201 sympathetic stimulations (Ekström, 1989; Proctor and Carpenter, 2007). The parasympathetic
202 signalling system in saliva secretion involves acetylcholine and noradrenaline release (Proctor and
203 Carpenter, 2014). Both acinar and duct cells express cholinergic receptors as acetylcholine is the
204 major neurotransmitter secreted by parasympathetic vagal fibres. Russell's viper venom consists of
205 pre-synaptically active neurotoxic, PLA₂ (Kalita et al., 2018). Therefore, further investigation into the
206 effects of this venom on salivary neural cells might shed light on the impact of Russell's viper bite on
207 the salivary neural systems and their association with the calculi formation. In addition, SBE including
208 from Russell's vipers is known to induce acute pain and long-term complications in multiple regions
209 other than the bite site (Waidyanatha et al., 2019). Therefore, it is possible that the calculus might
210 have formed already, but the area was sensitised and became painful due to the direct and/or indirect
211 actions of venom toxins.

212 We cannot rule out the effects of antivenom administered in this victim on sialolithiasis, as
213 the calculi formation was discovered post antivenom treatment. Although this patient did not display
214 any adverse reactions upon infusion of antivenom, the antigen-antibody (venom/antivenom)
215 complexes formed might have triggered inflammatory responses specifically in salivary glands
216 and/or ducts through inflammatory responses (de Silva et al., 2016; Morais and Massaldi, 2009).

217 The polyvalent antivenom used in this case is known to cause adverse reactions due to
218 immunoglobulin complexes and impurities. Among the adverse reactions caused by antivenom, IgE-
219 mediated and non-IgE mediated anaphylactic reactions are known to cause increased vascular
220 permeability and mucous secretion (de Silva et al., 2016; Morais and Massaldi, 2009). Similarly, the
221 fresh frozen plasma transfused in this patient could have triggered or contributed to this complication
222 as it is known induce hypersensitivity reactions (Pandey and Vyas, 2012) although it was not evident
223 in this case. These reactions could affect the rate of saliva production in submandibular glands and
224 its composition. Hence, this aspect should be further investigated.

225 In conclusion, it is not entirely clear if venom components alone induced the rapid formation
226 of poorly calcified salivary calculus in this case, or it was an aggravation of an already growing micro
227 calculi due to excessive inflammation. There is also a potential possibility that it was due to adverse
228 reactions to the antivenom or fresh frozen plasma. Further research into these types of unusual
229 cases can not only help us in understanding the molecular mechanisms of venom (and antivenom)
230 components but also enlighten us with the wide spectrum of envenomation complications which will
231 result in revising treatment guidelines for better clinical management of SBE. This case report will
232 assist clinicians in areas where SBE commonly occurs to be aware of such unusual clinical
233 manifestations specifically among children.

234 **Ethical statement**

235 The data collection, consent form, and information sheets were approved by the Institutional
236 Ethics Committee at Toxiven Biotech, Tamil Nadu, India (Reference number: ICMR-Toxiven Ethics
237 2021/1). A written consent was obtained from the patient's parents (since the patient was a minor)
238 to collect and publish the data presented in this article.

239 **References**

- 240 Ambudkar, I.S., 2016. Calcium signalling in salivary gland physiology and dysfunction. *J Physiol* 594,
241 2813-2824.
- 242 Boskey, A.L., Boyan-Salyers, B.D., Burstein, L.S., Mandel, I.D., 1981. Lipids associated with
243 mineralization of human submandibular gland sialoliths. *Arch Oral Biol* 26, 779-785.
- 244 Boskey, A.L., Burstein, L.S., Mandel, I.D., 1983. Phospholipids associated with human parotid gland
245 sialoliths. *Arch Oral Biol* 28, 655-657.
- 246 Chakrabarty, D., Sarkar, A., 2015. Cytotoxic Effects of Snake Venoms, in: Gopalakrishnakone, P.,
247 Inagaki, H., Mukherjee, A.K., Rahmy, T.R., Vogel, C.-W. (Eds.), *Snake Venoms*. Springer
248 Netherlands, Dordrecht, pp. 1-16.
- 249 de Silva, H.A., Ryan, N.M., de Silva, H.J., 2016. Adverse reactions to snake antivenom, and their
250 prevention and treatment. *Br J Clin Pharmacol* 81, 446-452.
- 251 Deepak, M., Basavaprabhu, A., Ramapuram, J.T., Nithyananda, C., Mahalingam, S., 2013. Bilateral
252 parotid enlargement following snake bite: a rare sign. *Asian Pac J Trop Biomed* 3, 154-155.
- 253 Delli, K., Spijkervet, F.K., Vissink, A., 2014. Salivary gland diseases: infections, sialolithiasis and
254 mucoceles. *Monogr Oral Sci* 24, 135-148.
- 255 Ekström, J., 1989. Autonomic control of salivary secretion. *Proc Finn Dent Soc* 85, 323-331;
256 discussion 361-323.
- 257 Epivatianos, A., Harrison, J.D., 1989. The presence of microcalculi in normal human submandibular
258 and parotid salivary glands. *Archives of Oral Biology* 34, 261-265.
- 259 Goncalves, M., Schapher, M., Iro, H., Wuest, W., Mantsopoulos, K., Koch, M., 2017. Value of
260 Sonography in the Diagnosis of Sialolithiasis: Comparison With the Reference Standard of Direct
261 Stone Identification. *Journal of Ultrasound in Medicine* 36, 2227-2235.
- 262 Grases, F., Santiago, C., Simonet, B.M., Costa-Bauzá, A., 2003. Sialolithiasis: mechanism of calculi
263 formation and etiologic factors. *Clin Chim Acta* 334, 131-136.
- 264 Gutiérrez, J.M., Calvete, J.J., Habib, A.G., Harrison, R.A., Williams, D.J., Warrell, D.A., 2017.
265 Snakebite envenoming. *Nature Reviews Disease Primers* 3, 17063.

- 266 Hammett, J.T., Walker, C., 2021. Sialolithiasis, StatPearls. StatPearls Publishing
 267 Copyright © 2021, StatPearls Publishing LLC., Treasure Island (FL).
- 268 Harrison, J.D., 2009. Causes, Natural History, and Incidence of Salivary Stones and Obstructions.
 269 Otolaryngologic Clinics of North America 42, 927-947.
- 270 Harrison, J.D., Triantafyllou, A., Baldwin, D., Schäfer, H., 1993. Histochemical and biochemical
 271 determination of calcium in salivary glands with particular reference to chronic submandibular
 272 sialadenitis. Virchows Arch A Pathol Anat Histopathol 423, 29-32.
- 273 Kalita, B., Mackessy, S.P., Mukherjee, A.K., 2018. Proteomic analysis reveals geographic variation
 274 in venom composition of Russell's Viper in the Indian subcontinent: implications for clinical
 275 manifestations post-envenomation and antivenom treatment. Expert Rev Proteomics 15, 837-849.
- 276 Katkar, G.D., Sundaram, M.S., NaveenKumar, S.K., Swethakumar, B., Sharma, R.D., Paul, M.,
 277 Vishalakshi, G.J., Devaraja, S., Girish, K.S., Kemparaju, K., 2016. NETosis and lack of DNase
 278 activity are key factors in Echis carinatus venom-induced tissue destruction. Nature Communications
 279 7, 11361.
- 280 Kim, J.H., Aoki, E.M., Cortes, A.R.G., Abdala-Júnior, R., Asaumi, J., Arita, E.S., 2016. Comparison
 281 of the diagnostic performance of panoramic and occlusal radiographs in detecting submandibular
 282 sialoliths. Imaging Sci Dent 46, 87-92.
- 283 Kondo, Y., Nakamoto, T., Jaramillo, Y., Choi, S., Catalan, M.A., Melvin, J.E., 2015. Functional
 284 differences in the acinar cells of the murine major salivary glands. J Dent Res 94, 715-721.
- 285 Kraaij, S., Karagozoglu, K.H., Forouzanfar, T., Veerman, E.C.I., Brand, H.S., 2014. Salivary stones:
 286 symptoms, aetiology, biochemical composition and treatment. British Dental Journal 217, E23-E23.
- 287 Leppkes, M., Maueröder, C., Hirth, S., Nowecki, S., Günther, C., Billmeier, U., Paulus, S., Biermann,
 288 M., Munoz, L.E., Hoffmann, M., Wildner, D., Croxford, A.L., Waisman, A., Mowen, K., Jenne, D.E.,
 289 Krenn, V., Mayerle, J., Lerch, M.M., Schett, G., Wirtz, S., Neurath, M.F., Herrmann, M., Becker, C.,
 290 2016. Externalized decondensed neutrophil chromatin occludes pancreatic ducts and drives
 291 pancreatitis. Nat Commun 7, 10973.
- 292 Marchal, F., Kurt, A.-M., Dulguerov, P., Lehmann, W., 2001. Retrograde Theory in Sialolithiasis
 293 Formation. Archives of Otolaryngology–Head & Neck Surgery 127, 66-68.
- 294 Maueröder, C., Kienhöfer, D., Hahn, J., Schauer, C., Manger, B., Schett, G., Herrmann, M.,
 295 Hoffmann, M.H., 2015. How neutrophil extracellular traps orchestrate the local immune response in
 296 gout. Journal of Molecular Medicine 93, 727-734.
- 297 Menaldo, D.L., Bernardes, C.P., Pereira, J.C., Silveira, D.S.C., Mamede, C.C.N., Stanziola, L.,
 298 Oliveira, F.d., Pereira-Crott, L.S., Faccioli, L.H., Sampaio, S.V., 2013. Effects of two serine proteases
 299 from Bothrops pirajai snake venom on the complement system and the inflammatory response.
 300 International Immunopharmacology 15, 764-771.
- 301 Morais, V., Massaldi, H., 2009. Snake antivenoms: Adverse reactions and production technology.
 302 Journal of Venomous Animals and Toxins including Tropical Diseases 15.
- 303 Muñoz, L.E., Boeltz, S., Bilyy, R., Schauer, C., Mahajan, A., Widulin, N., Grüneboom, A., Herrmann,
 304 I., Boada, E., Rauh, M., Krenn, V., Biermann, M.H.C., Podolska, M.J., Hahn, J., Knopf, J.,
 305 Maueröder, C., Paryzhak, S., Dumych, T., Zhao, Y., Neurath, M.F., Hoffmann, M.H., Fuchs, T.A.,
 306 Leppkes, M., Schett, G., Herrmann, M., 2019. Neutrophil Extracellular Traps Initiate Gallstone
 307 Formation. Immunity 51, 443-450.e444.
- 308 Pandey, S., Vyas, G.N., 2012. Adverse effects of plasma transfusion. Transfusion 52 Suppl 1, 65S-
 309 79S.
- 310 Proctor, G.B., Carpenter, G.H., 2007. Regulation of salivary gland function by autonomic nerves.
 311 Auton Neurosci 133, 3-18.
- 312 Proctor, G.B., Carpenter, G.H., 2014. Salivary secretion: mechanism and neural regulation. Monogr
 313 Oral Sci 24, 14-29.

314 Samuel, S.P., Chinnaraju, S., Williams, H.F., Pichamuthu, E., Subharao, M., Vaiyapuri, M.,
315 Arumugam, S., Vaiyapuri, R., Baksh, M.F., Patel, K., Trim, S.A., Duncombe, T.E., Vaiyapuri, S.,
316 2020. Venomous snakebites: Rapid action saves lives-A multifaceted community education
317 programme increases awareness about snakes and snakebites among the rural population of Tamil
318 Nadu, India. *PLoS Negl Trop Dis* 14, e0008911.

319 Sasidaran, M.N., Samuel, S.P., ChinnaRaju, S., Antonysamy, M., Bulfone, T.C., Lewin, M.R., 2017.
320 Parotid swelling after Russell's viper envenomation: an unusual and poor prognostic sign. *Clin Case*
321 *Rep* 6, 262-266.

322 Schapher, M., Koch, M., Weidner, D., Scholz, M., Wirtz, S., Mahajan, A., Herrmann, I., Singh, J.,
323 Knopf, J., Leppkes, M., Schauer, C., Grüneboom, A., Alexiou, C., Schett, G., Iro, H., Muñoz, L.E.,
324 Herrmann, M., 2020. Neutrophil Extracellular Traps Promote the Development and Growth of Human
325 Salivary Stones. *Cells* 9.

326 Schrøder, S.A., Homøe, P., Wagner, N., Bardow, A., 2017. Does saliva composition affect the
327 formation of sialolithiasis? *The Journal of Laryngology & Otology* 131, 162-167.

328 Senthilkumaran, S., Williams, H.F., Patel, K., Trim, S.A., Thirumalaikolundusubramanian, P.,
329 Vaiyapuri, S., 2021. Priapism following a juvenile Russell's viper bite: An unusual case report. *PLOS*
330 *Neglected Tropical Diseases* 15, e0009242.

331 Sigismund, P.E., Zenk, J., Koch, M., Schapher, M., Rudes, M., Iro, H., 2015. Nearly 3,000 salivary
332 stones: Some clinical and epidemiologic aspects. *The Laryngoscope* 125, 1879-1882.

333 Silva, A., Kuruppu, S., Othman, I., Goode, R.J., Hodgson, W.C., Isbister, G.K., 2017. Neurotoxicity
334 in Sri Lankan Russell's Viper (*Daboia russelii*) Envenoming is Primarily due to U1-viperitoxin-Dr1a,
335 a Pre-Synaptic Neurotoxin. *Neurotox Res* 31, 11-19.

336 Silva, A., Maduwage, K., Sedgwick, M., Pilapitiya, S., Weerawansa, P., Dahanayaka, N.J., Buckley,
337 N.A., Siribaddana, S., Isbister, G.K., 2016. Neurotoxicity in Russell's viper (*Daboia russelii*)
338 envenoming in Sri Lanka: a clinical and neurophysiological study. *Clin Toxicol (Phila)* 54, 411-419.

339 Sivasubramaniam, R., Choroomi, S., Stone, H., 2015. Sialolithiasis in a Remnant Wharton s Duct: a
340 Case Study and Discussion. *Journal of Otolaryngology-ENT Research* 2.

341 Subasinghe, C.J., Sarathchandra, C., Kandeepan, T., Kulatunga, A., 2014. Bilateral blindness
342 following Russell's viper bite - a rare clinical presentation: a case report. *Journal of Medical Case*
343 *Reports* 8, 99.

344 Tadokoro, T., Modahl, C.M., Maenaka, K., Aoki-Shioi, N., 2020. Cysteine-Rich Secretory Proteins
345 (CRISPs) From Venomous Snakes: An Overview of the Functional Diversity in A Large and
346 Underappreciated Superfamily. *Toxins* 12, 175.

347 Teixeira Cde, F., Fernandes, C.M., Zuliani, J.P., Zamuner, S.F., 2005. Inflammatory effects of snake
348 venom metalloproteinases. *Mem Inst Oswaldo Cruz* 100 Suppl 1, 181-184.

349 Vaiyapuri, S., Vaiyapuri, R., Ashokan, R., Ramasamy, K., Nattamaisundar, K., Jeyaraj, A.,
350 Chandran, V., Gajjeraman, P., Baksh, M.F., Gibbins, J.M., Hutchinson, E.G., 2013. Snakebite and
351 Its Socio-Economic Impact on the Rural Population of Tamil Nadu, India. *PLOS ONE* 8, e80090.

352 Waidyanatha, S., Silva, A., Siribaddana, S., Isbister, G.K., 2019. Long-term Effects of Snake
353 Envenoming. *Toxins* 11, 193.

354 Williams, H.F., Layfield, H.J., Vallance, T., Patel, K., Bicknell, A.B., Trim, S.A., Vaiyapuri, S., 2019.
355 The Urgent Need to Develop Novel Strategies for the Diagnosis and Treatment of Snakebites. *Toxins*
356 (Basel) 11.

357
358
359
360
361

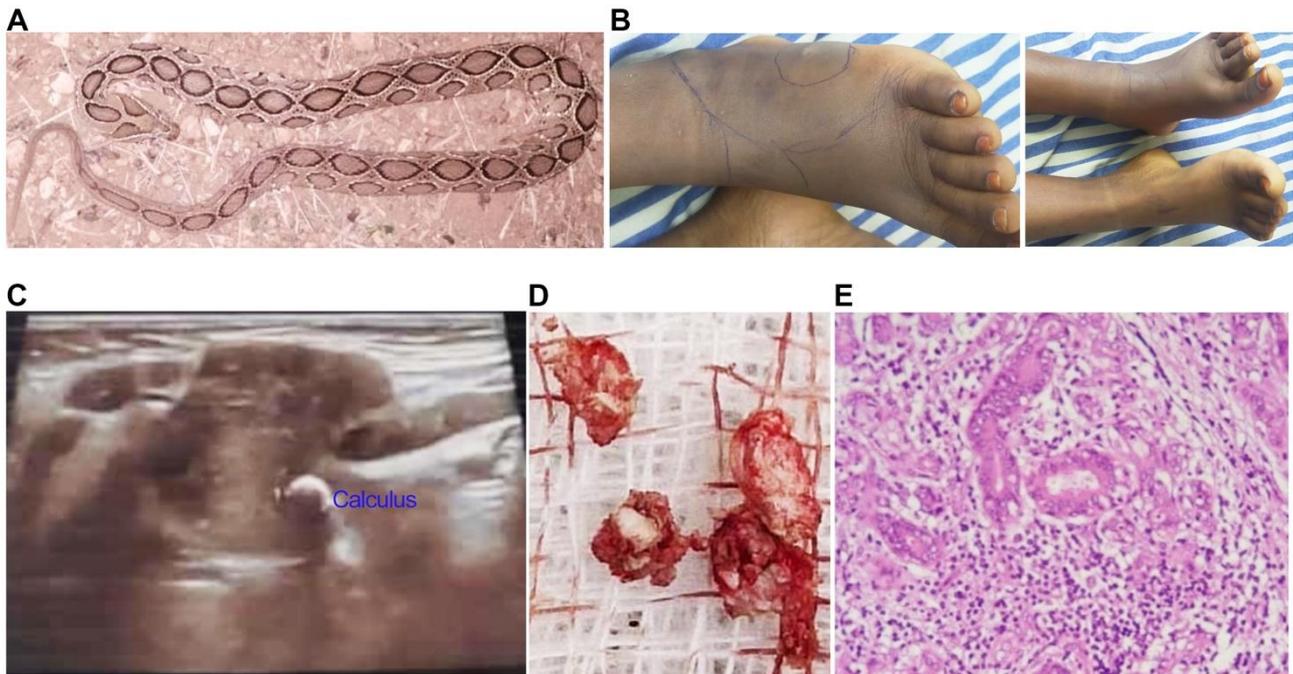
362 **Figure Legnds**

363

364 **Figure 1:** **A**, the offending snake was identified as a Russell's viper. **B**, bite area shows mild swelling
365 and developing wound. **C**, an ultrasound scan reveals the presence of a calculus at the duct of
366 submandibular gland. **D**, extracted calculus with affected tissues using a minor surgery. **E**,
367 haemotoxylin and eosin staining of extracted salivary tissues demonstrates the excessive infiltration
368 of leukocytes.

369

370 **Figure 1**



371

372

373

374

375

376

377

378

379

380

381

382

383

384

385

386

387

388

389 **Table 1:** Salivary analysis report

390

Parameter analysed	Results	Unit	Reference range
Colour	White	-	-
Appearance	Turbid	-	-
Reaction	Alkaline	-	-
pH	7.12	-	-
Specific gravity	1.53	-	-
Cast	Mucus ++	-	-
Crystals	None	-	-
Mucus threads	++	-	-
Others	None	-	-
Sodium	130	mmol/L	135 to 150
Potassium	4.0	mmol/L	3.5 to 5.0
Chloride	112	mmol/L	98 to 107
Bicarbonate	21	mmol/L	22 to 29
Calcium	7.1	mg/dL	8.4 to 10.2
Pus cells	0	-	0-1/HPF
Red blood cells	0	-	0-1/HPF
Epithelial cells	4-6	-	2-3/HPF
Bacteria	Present	-	-

391

392

393