

Phosphorus uptake and toxicity is delimited by mycorrhizal symbiosis in P-sensitive Eucalyptus marginata but not in P-tolerant Acacia celastrifolia

Article

Published Version

Creative Commons: Attribution 4.0 (CC-BY)

Open Access

Tibbett, M. ORCID: <https://orcid.org/0000-0003-0143-2190>, Daws, M. I. and Ryan, M. H. (2022) Phosphorus uptake and toxicity is delimited by mycorrhizal symbiosis in P-sensitive Eucalyptus marginata but not in P-tolerant Acacia celastrifolia. AoB PLANTS, 14 (5). plac037. ISSN 2041-2851 doi: 10.1093/aobpla/plac037 Available at <https://centaur.reading.ac.uk/107156/>

It is advisable to refer to the publisher's version if you intend to cite from the work. See [Guidance on citing](#).

To link to this article DOI: <http://dx.doi.org/10.1093/aobpla/plac037>

Publisher: Oxford Academic

All outputs in CentAUR are protected by Intellectual Property Rights law, including copyright law. Copyright and IPR is retained by the creators or other copyright holders. Terms and conditions for use of this material are defined in the [End User Agreement](#).

www.reading.ac.uk/centaur

CentAUR

Central Archive at the University of Reading

Reading's research outputs online

Studies

Phosphorus uptake and toxicity are delimited by mycorrhizal symbiosis in P-sensitive *Eucalyptus marginata* but not in P-tolerant *Acacia celastrifolia*

Mark Tibbett^{1,2,*}, Matthew I. Daws¹ and Megan H. Ryan³

¹Department of Sustainable Land Management and Soil Research Centre, School of Agricultural Policy and Development, University of Reading, Reading, Berkshire RG6 6AR, UK

²School of Biological Sciences, The University of Western Australia, 35 Stirling Highway, Crawley, WA 6009, Australia

³School of Agriculture and Environment, The University of Western Australia, 35 Stirling Highway, Crawley, WA 6009, Australia

*Corresponding author's e-mail address: m.tibbett@reading.ac.uk

Form & Function. Chief Editor: Kate McCulloh

Associate Editor: Mikio Nakazono

Abstract

Many plant species from regions with ancient, highly weathered nutrient-depleted soils have specialized adaptations for acquiring phosphorus (P) and are sensitive to excess P supply. Mycorrhizal associations may regulate P uptake at high external P concentrations, potentially reducing P toxicity. We predicted that excess P application will negatively impact species from the nutrient-depleted Jarrah forest of Western Australia and that mycorrhizal inoculation will reduce P toxicity by regulating P uptake. For seedlings of the N₂-fixing legume *Acacia celastrifolia* and the tree species *Eucalyptus marginata*, we measured growth at P concentrations of 0–90 mg kg⁻¹ soil and in relation to inoculation with the arbuscular mycorrhizal fungus (AMF) *Rhizophagus irregularis*. Non-inoculated *A. celastrifolia* maintained leaf P concentrations at <2 mg g⁻¹ dry mass (DM) across the range of external P concentrations. However, for non-inoculated *E. marginata*, as external P concentrations increased, leaf P also increased, reaching >9 mg g⁻¹ DM at 30 mg P kg⁻¹ soil. *Acacia celastrifolia* DM increased with increasing external P concentrations, while *E. marginata* DM was maximal at 15 mg P kg⁻¹ soil, declining at higher external P concentrations. Neither DM nor leaf P of *A. celastrifolia* was affected by inoculation with AMF. For *E. marginata*, even at 90 mg P kg⁻¹ soil, inoculation with AMF resulted in leaf P remaining <1 mg g⁻¹ DM, and DM being maintained. These data strengthen the evidence base that AMF may not only facilitate P uptake at low external P concentrations, but are also important for moderating P uptake at elevated external P concentrations and maintaining plant P concentrations within a relatively narrow concentration range.

Keywords: *Acacia*; fertilizer; mycorrhiza; P accumulation; P-use efficiency; P toxicity; rehabilitation; restoration.

Introduction

Ancient and highly weathered soils, such as those found in South West Western Australia, have naturally low phosphorus (P) concentrations. Many plants adapted to growth under these low P conditions have evolved a range of strategies for P acquisition, including cluster roots and exudation of carboxylates and phosphatases (Lambers *et al.* 2006, 2008). The majority of plant taxa from one of many mycorrhizal symbiotic types (Kariman *et al.* 2018), many of which serve to exploit soil nutrients through mycelia scavenging of resources not available to roots alone (Tibbett 2000; Tibbett and Sanders 2002; Smith *et al.* 2015).

Species adapted to naturally low soil P concentrations may display symptoms of P toxicity when supplied with P concentrations above those that they experience naturally in soil (Handreck 1991; Lambers *et al.* 2002; Shane *et al.* 2004b; Standish *et al.* 2007; Pang *et al.* 2010; de Campos *et al.* 2013;

Williams *et al.* 2019), due potentially to the loss of low-affinity transporter systems (Huang *et al.* 2011). P-sensitive species occur in a range of families, including the Fabaceae, Haemodoraceae, Myrtaceae, Proteaceae and Rutaceae. Symptoms of P sensitivity are highly species-specific and occur at shoot P concentrations less than 1 mg g⁻¹ dry mass (DM) to more than 40 mg P g⁻¹ DM (Shane *et al.* 2004a and references therein). Symptoms of P toxicity include a reduction in growth with increasing external P concentration (e.g. Standish *et al.* 2007; Williams *et al.* 2019), and visible symptoms including early leaf senescence and necrotic and chlorotic regions on leaves (e.g. Handreck 1991; Lambers *et al.* 2002; Shane *et al.* 2004a, b; Kariman *et al.* 2014a; Ye *et al.* 2021).

Mycorrhizal symbioses are well known for increasing P uptake in nutrient-deficient soils and increasing the P status of host plants (Bougher *et al.* 1990; Koide and Mosse 2004;

Received: 04 October 2021; Editorial decision: 2 July 2022; Accepted: 15 August 2022

© The Author(s) 2022. Published by Oxford University Press on behalf of the Annals of Botany Company.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<https://creativecommons.org/licenses/by/4.0/>), which permits unrestricted reuse, distribution, and reproduction in any medium, provided the original work is properly cited.

Smith *et al.* 2011; Kariman *et al.* 2018; Standish *et al.* 2022). However, they can also enable the growth of plants in soils containing toxic concentrations of heavy metals, or certain essential trace elements such as cadmium and zinc, by controlling the uptake of metal ions (Jentschke and Godbold 2000; Hildebrandt *et al.* 2007; de Oliveira *et al.* 2020; Yazici *et al.* 2021). Similarly, AMF can modify P uptake in the host plant by downregulating the expression of genes encoding high-affinity phosphate transporter proteins (Burleigh and Bechmann 2002). While mycorrhizal associations can increase plant shoot P concentrations by increasing uptake at low P availability and enabling exploitation of a greater soil volume (Tibbett 2000), there is increasing evidence for AMF and ectomycorrhizal fungi (ECM) (Kariman *et al.* 2014a) that they can also moderate shoot P at high P availability. For example, Nazeri *et al.* (2014) demonstrated, for a range of legume species, that inoculation with AMF could maintain shoot P concentrations within relatively narrow boundaries following the application of a single pulse of P. This effect was modulated by both mycorrhizal-related reductions in rhizosphere carboxylates and P transport from roots to shoots. Conversely, high P availability can itself inhibit both formation of arbuscules and intraradical colonization (e.g. Müller and Harrison 2019).

Jarrah (*Eucalyptus marginata*) is a dominant overstorey tree in the Jarrah forest of South West Western Australia and, based on pot experiments, is known to be sensitive to elevated external P (Kariman *et al.* 2014a, 2016). For example, in the absence of inoculation with AMF, Kariman *et al.* (2014a) reported the onset of leaf chlorosis and necrosis in the week following application of a pulse of P to Jarrah seedlings. When seedlings were inoculated with AMF, visible symptoms associated with P toxicity and shoot P concentrations were reduced. While studies have largely focused on visible phytotoxicity symptoms resulting from P application (e.g. Handreck 1991; Lambers *et al.* 2002; Shane *et al.* 2004a; Kariman *et al.* 2014a), a recent study by Williams *et al.* (2019) demonstrated that P toxicity can also be expressed as a significant reduction in growth rates at shoot P concentrations that do not necessarily result in visible symptoms. Consequently, there is a need to better understand both longer-term effects of applying P and potential interactions of AMF on shoot P concentrations and plant growth.

For Jarrah, these observations in relation to P may also have significant practical implications: large areas of Jarrah forest are cleared and restored each year following bauxite mining. Fertilizer application (especially P), to maximize early plant growth, is generally viewed as a key step in the rehabilitation process (Ward *et al.* 1990; Grant *et al.* 2007; Tibbett 2010). However, there may be the potential for negative impacts on plant growth from applying excess P, particularly given newly restored forest has low species diversity and abundance of AMF (Gardner and Malajkuk 1988; Glen *et al.* 2008). Our own unpublished observations have found very poor levels of colonization by AMF (near absent) in seedlings of acacias and eucalypts in recently restored sites (M. Tibbett and M. H. Ryan, unpublished).

In the light of potential P toxicities in tree seedlings, and the prospect for symbiotic mitigation of such effects, we investigated two linked hypotheses anticipating contrasting responses for two species with distinctive ecological strategies: *Acacia celastrifolia* and *E. marginata*. *Acacia celastrifolia* is a large understorey ruderal legume that exhibits a strong

growth response as a seedling in the field and Jarrah (*E. marginata*) is the dominant overstorey tree which constitutes around 80 % of stems in the native forest (Daws *et al.* 2015; Tibbett *et al.* 2020). Like many Australian *Acacia* spp., *A. celastrifolia* can form associations with AMF and N₂-fixing rhizobia and *E. marginata* can form associations with both AMF and ECM (Reddell and Milnes 1992; Kariman *et al.* 2012). We undertook two experiments, the first one to assess the effect of P-application rate on growth of both species and the second to assess the effect of AMF inoculation on the response to P application. Based on these experiments we tested firstly the hypothesis that *A. celastrifolia* would exhibit a strong growth response to P addition over a wide range of exogenous supply, whereas Jarrah would not, and would potentially show signs of growth depression and toxicity at high P-application rates. Our second hypothesis was that AMF inoculation would alter the response in terms of P supply, growth and uptake, leading to a suppression in acacia growth and offer a remedial effect on Jarrah growth response and P uptake at high amendment rates.

Materials and Methods

Plant and soil material

Seeds of *E. marginata* (Jarrah) and *A. celastrifolia* were collected in the northern Jarrah forest of Western Australia, ca. 130 km SSE of the state capital Perth (32°48'S, 116°28'E). Seeds were collected from a minimum of five individual plants per species. *Eucalyptus marginata* is the dominant tree species and *A. celastrifolia* is an abundant and widespread understorey shrub in the Jarrah forest. Prior to experimentation, seeds were surface-sterilized in a 1:3 dilution of sodium hypochlorite solution for 10 min followed by repeated rinsing in distilled water. The water-impermeable seed coat of *A. celastrifolia* was chipped at the end furthest from the axis using a scalpel and seeds of both species were soaked for 2 h in 1:10 'Seed Starter' smoke water (Kings Park Botanic Gardens and Parks Authority, Perth, WA). Seeds were sown on the surface of 10 % agar water and placed at 15 °C in the dark.

For both experiments, once seeds had commenced germination, seedlings were transplanted into sealed 9.6-L pots at a depth of 10 mm. The pots contained ca. 4 L of disinfested topsoil that had been steamed twice for 3 h at 80 °C, dried at 40 °C and then sieved to 4 mm. The topsoil used in the experiment was also obtained from the northern Jarrah forest of Western Australia (32°48'S, 116°28'E). Jarrah forest soils are gravelly with low concentrations of available N, P and K (Tibbett *et al.* 2020) and high rates of P fixation on amorphous iron and aluminium oxides. Soil available (Colwell) P concentrations are low, typically ~2–3 mg kg⁻¹ soil (Tibbett *et al.* 2020; Daws *et al.* 2021). Ten germinated seeds were placed into each pot at a depth of 10 mm. For the second, mycorrhizal inoculation experiment, inoculum of *Rhizophagus irregularis* (formerly *Glomus intraradices*) was placed beneath the seeds at the time of planting. The inoculum was created using single species pot cultures from the University of Western Australia culture collection, originally sourced from INVAM (<https://invam.wvu.edu/home>). The AM inoculum consisted of *R. irregularis* hyphae and colonized leek (*Allium porrum*) roots, grown in sterilized river sand. To balance any potential edaphic changes that might

occur from adding this culture material to the experimental pots containing Jarrah forest soil, autoclaved culture material was placed beneath the seeds in the non-mycorrhizal treatments. Pots were watered weekly to 50 % field capacity and seedlings were thinned to the two healthiest plants after 21 days, and then down to one plant after 31 days.

To ensure that only P was limiting in the experiment, and that no nutrient imbalances were induced by the addition of P, 10 mL of modified Long Ashton's nutrient solution (minus P) was added to each pot 15 days after seedlings were planted (Cavagnaro *et al.*, 2001). Macronutrients: K_2SO_4 (20 mM), $MgSO_4 \cdot 7H_2O$ (15 mM), $CaCl_2 \cdot 2H_2O$ (30 mM), FeEDTA (1 mM), $(NH_4)_2SO_4$ (40 mM), $NaNO_3$ (80 mM). Micronutrients: H_3BO_3 (28.6 mg L⁻¹), $MnCl_2 \cdot 4H_2O$ (18.1 mg L⁻¹), $ZnSO_4 \cdot 7H_2O$ (2.2 mg L⁻¹), $CuSO_4 \cdot 5H_2O$ (0.8 mg L⁻¹), $NaMoO_4 \cdot 2H_2O$ (0.25 mg L⁻¹).

Experimental design

Twenty-three days after planting, phosphate was added to the P treatments in the form of potassium dihydrogen phosphate (KH_2PO_4). To ensure a constant ionic background and balanced potassium levels, potassium chloride (KCl) was added in inverse proportions to KH_2PO_4 amendments. In Experiment 1, eight rates of P application were used (equivalent to 0, 0.9, 4.5, 13.5, 22.5, 31.5, 40.5 and 81 mg elemental P kg⁻¹ soil) to assess the susceptibility of the two plant species to different external P concentrations. In Experiment 2 there were four P-application rates (equivalent to 0, 4.5, 30 and 90 P kg⁻¹ soil) in a two-way factorial combination of P-application rate × mycorrhizal treatment to test the effect of AMF on P uptake at different external P concentrations. Both experiments were established in randomized blocks in a glasshouse, with each block containing all treatments. The temperature-controlled glasshouse was maintained at temperatures between 18 °C and 28 °C. Pots were regularly re-randomized throughout the growing period. All treatments were replicated three times for Experiment 1 and four times for Experiment 2. The levels of P-application rate were chosen to span the entire range of current and past fertilizer prescriptions applied in the rehabilitation of Jarrah forest ecosystems (Standish *et al.* 2015).

Mycorrhizal activity (Experiment 2)

Thirteen days after transplanting and at the end of the experiment (188 days), seedlings were screened for evidence of colonization by AMF. Roots were cleared using KOH then stained using lactic-glycerol blue and examined by light microscopy for evidence of colonization (Brundrett *et al.* 1996). At the end of the experiment, AM spores were also extracted from 150 g of soil sampled away from the middle of the pots following wet sieving and sucrose centrifugation (Walker *et al.* 1982). Spores were extracted and counted (in addition to direct measures of colonization) as sporulation provides an indication of the overall level of mycorrhizal abundance and reproduction over the duration of the experiment.

Plant measurements

For Experiment 1, plants were harvested 213 days after transplanting and roots and shoots dried separately at 70 °C for dry weight determination. For Experiment 2, plants were harvested for DM determination 53 or 188 days after transplanting. In both experiments, plants were carefully removed

from the growing medium, roots washed with water and the plants separated into roots and shoots.

Foliar P concentrations

Dried leaf material from the last sampling time points in both experiments was ground and then subsampled for digestion. Leaf material was digested using an $HNO_3/HClO_4$ mixture with the diluted digest approximately 10 % v/v with respect to $HClO_4$ (70 % w/w). Phosphorus content was determined using the molybdovanadophosphate method (yellow) and a spectrometer reading at a wavelength of 460 nm (modified from Simmons 1975, 1978).

Statistical analysis

For Experiment 1, two-way ANOVA implemented in Minitab 14 was used to assess, whether there were effects of (i) species and (ii) increasing external P concentration, on either plant DM or leaf P concentration. Data were log₁₀ transformed. For Experiment 2, two-way ANOVA was used to assess, for each of the two species, the effect of (i) inoculation with AMF and (ii) external P concentration on either plant DM or leaf P concentration. Finally, for Experiment 2, for the plants of each of the study species inoculated with AMF, two-way ANOVA was used to assess the interaction of (i) study species and (ii) external P concentration on spore count in the soil at the end of the experiment. Spore counts were log₁₀(*n* + 1) transformed to ensure normality.

Results

Experiment 1: Effect of external P concentration on DM and leaf P of non-inoculated plants

For non-inoculated plants, there was a significant effect of P-application rate on plant DM after 213 days (two-way ANOVA, $F_{1,32} = 18.50$, $P < 0.001$), but the two species (*A. celastriifolia* and *E. marginata*) responded differently to applied P (two-way ANOVA, $F_{1,32} = 107.20$, $P < 0.001$; Fig. 1). Across the range of concentrations from 0 to 15 mg P kg⁻¹ soil, *A. celastriifolia* DM increased rapidly with increasing P. At higher P-application rates, the rate of increase in DM declined. Nonetheless, total DM at the highest application rate (81 mg kg⁻¹ soil) was 10× higher than at 0 mg P kg⁻¹ soil (Fig. 1). For *E. marginata*, there was also an initial increase in DM as P-application rate increased from 0 to 15 mg kg⁻¹ soil (Fig. 1). However, at P-application rates greater than 15 mg kg⁻¹, DM declined: plant mass at a P-application rate of 15 mg kg⁻¹ was more than twice that at 81 mg kg⁻¹ (Fig. 1).

Leaf P concentrations of *A. celastriifolia* were ca. 0.65 mg g⁻¹ DM for plants at the nil P-application rate, then increasing to a maximum of ca. 3.2 mg g⁻¹ DM at the P-application rate of 81 mg kg⁻¹ soil (Fig. 2). For *E. marginata* leaf P was ca. 0.22 mg g⁻¹ DM at the nil P-application rate (Fig. 2) and increased rapidly with P application reaching ca. 9 mg g⁻¹ DM at 30 mg P kg⁻¹. As the P-application rate increased to 80 mg kg⁻¹, leaf P concentrations remained at ca. 9 mg g⁻¹ DM (Fig. 2). These responses were reflected in significant main effects of P-application rate (two-way ANOVA, $F_{7,32} = 19.58$, $P < 0.001$) and species (two-way ANOVA, $F_{1,32} = 26.28$, $P < 0.001$) as well as a significant P application × species interaction (two-way ANOVA, $F_{7,32} = 4.85$, $P = 0.001$). For *E. marginata* the leaf P concentration corresponding to maximum plant DM accumulation, and above which DM accumulation declined, was ca. 4 mg g⁻¹ DM (Figs 1 and 2).

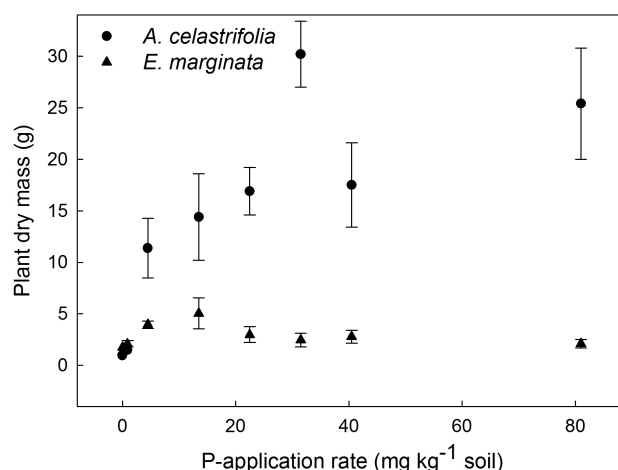


Figure 1. The effect of P-application rate on plant DM of non-AMF-inoculated *Acacia celastrifolia* and *Eucalyptus marginata* assessed 213 days after transplanting. Bars \pm 1 SE.

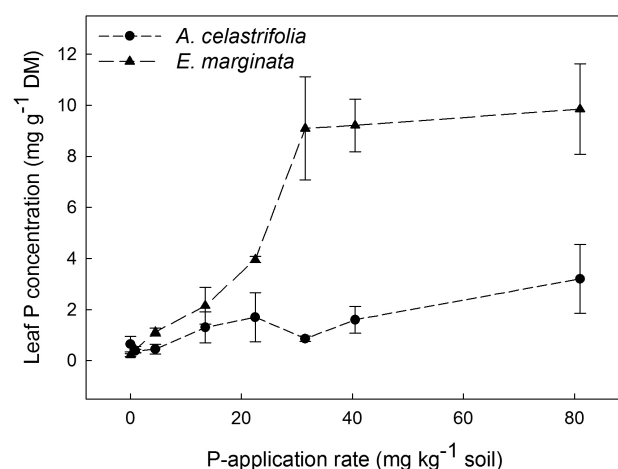


Figure 2. The effect of P-application rate on leaf P concentration for non-AMF-inoculated *Acacia celastrifolia* and *Eucalyptus marginata* assessed 213 days after transplanting. Bars \pm 1 SE.

Experiment 2: Effect of AM colonization on DM and leaf P

Evidence of early colonization (6–29 %) by AM was found in 4 of the 12 samples taken (one *A. celastrifolia* and three *E. marginata*). Roots taken at the end of the experiment did not clear properly and were not able to be assessed reliably and consequently were discarded. There was no evidence of colonization by ECM for either plant species, nor was there evidence of nodulation in *A. celastrifolia*. Spores were found only in the mycorrhizal treatments, and levels of sporulation varied consistently with P-application rates, supporting the use of sporulation as an acceptable proxy for measuring mycorrhizal activity. Spore counts from mycorrhizal pots were low (under 100 per 150 g soil) for both *A. celastrifolia* and *E. marginata* at the nil P-application rate, whereas no spores were found in non-inoculated treatments. There was a significant effect of plant species on sporulation in the inoculated treatment (two-way ANOVA, $F_{1,24} = 11.64$, $P < 0.01$) with sporulation varying greatly between the species at the nil P-application rate, peaking at 4.5 mg kg⁻¹ P for *E. marginata* yet remaining fairly low at all P-application rates (Table 1).

Table 1. The effect of plant species, P-application rate and inoculating pots at the outset of the experiment with the AMF *Rhizophagus irregularis* on spore counts at the end of the experiment. Spore counts were taken at the end of Experiment 2 on Day 188. Error bars are \pm 1 SE of the mean.

Plant species	P-application rate (mg kg ⁻¹ soil)	Spore count (pots inoculated with AMF) (# spores 150 g ⁻¹ soil)	Spore count (pots not inoculated with AMF) (# spores 150 g ⁻¹ soil)
<i>Acacia celastrifolia</i>	0	45.3 \pm 4.4	0
	4.5	117.3 \pm 13.0	0
	30	191.8 \pm 27.0	0
	90	276.0 \pm 52.5	0
<i>Eucalyptus marginata</i>	0	80.5 \pm 3.5	0
	4.5	103.3 \pm 9.0	0
	30	58.5 \pm 5.8	0
	90	37.0 \pm 4.3	0

In contrast, sporulation continued increasing with P application for *A. celastrifolia*, peaking at 90 mg kg⁻¹ P with a mean of 276 spores per 150 g of soil. This difference in response between the two species was reflected in the P-application rate \times species interaction being highly significant (two-way ANOVA, $F_{3,24} = 9.34$, $P < 0.001$), but the main effect of P application being non-significant (two-way ANOVA, $F_{3,24} = 1.91$, $P = 0.154$).

For both plants inoculated and non-inoculated with AMF, for the two time periods that were measured (53 and 188 days), there was a significant effect of P-application rate on DM for *A. celastrifolia* (two-way ANOVA, $F_{3,24} = 11.0$, $P < 0.001$ and $F_{3,24} = 4.94$, $P < 0.01$ for 53 and 188 days, respectively) and *E. marginata* (two-way ANOVA, $F_{3,24} = 3.92$, $P < 0.05$ and $F_{3,24} = 4.32$, $P < 0.05$ for 53 and 188 days, respectively) (Fig. 3A and B). *Acacia celastrifolia* DM increased with P-application rate and the rate of increase declined above a P-application rate of 30 mg kg⁻¹ soil. For *A. celastrifolia*, there was no effect of AM inoculation on DM accumulation for either measurement interval (Fig. 3A and B; two-way ANOVA, $F_{1,24} = 1.96$, $P = 0.175$ and $F_{1,24} = 1.01$, $P = 0.324$ for 53 and 188 days, respectively).

At both measurement intervals, DM of non-inoculated *E. marginata* plants increased reaching a maximum at the P-application rate of 30 mg kg⁻¹ soil and declined thereafter (Fig. 3C and D). At a P-application rate of 90 mg kg⁻¹ soil, there were visible symptoms of P toxicity including leaf necrosis. Further, for the replicate sampled at 53 days after planting, only one of the four replicate plants was still alive. For plants inoculated with AMF, there was no effect of P-application rate on DM accumulation. Further, neither a decline in DM nor visible symptoms of P toxicity were observed at the highest P-application rate of 90 mg kg⁻¹ soil (Fig. 3C and D). At the second measuring interval these responses were reflected in a significant P-application rate \times AMF inoculation interaction (two-way ANOVA, $F_{3,24} = 3.62$, $P < 0.05$). Profound differences between inoculated and non-inoculated plants can be seen in Supporting Information—Fig. S1.

Leaf P concentration of *A. celastrifolia* increased from ca. 0.6 to 2 mg g⁻¹ DM as the P-application rate increased from 0 to 90 mg kg⁻¹ soil with this response being significant

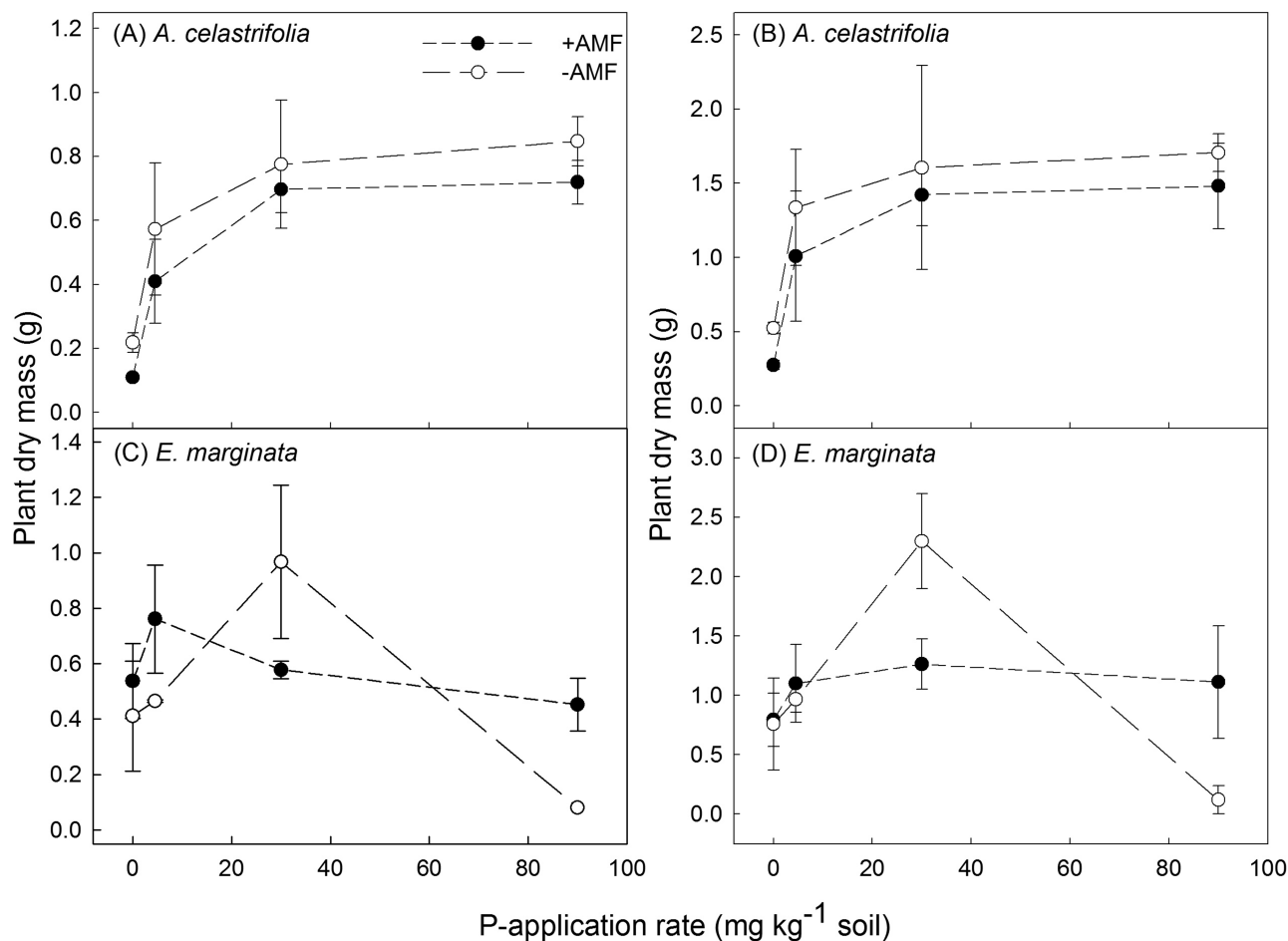


Figure 3. The effect of P-application rate and inoculation with the AMF *Rhizophagus irregularis* on plant DM assessed either 53 (A and C) or 188 days after transplanting (B and D) for *Acacia celastrifolia* and *Eucalyptus marginata*. Note that due to elevated mortality at high P, the 53-day data point for *E. marginata* at 90 mg P kg⁻¹ consists of data from one plant only. Bars ± 1 SE.

(two-way ANOVA, $F_{3,24} = 63.77$, $P < 0.001$). This response was independent of inoculation with AMF (Fig. 4A; two-way ANOVA, $F_{1,24} = 0.14$, $P = 0.71$). For non-inoculated *E. marginata* plants, the leaf P concentration increased from ca. 0.2 to 9 mg g⁻¹ DM as the P-application rate increased from 0 to 90 mg kg⁻¹ soil (Fig. 4B). However, for plants of *E. marginata* inoculated with AMF, leaf P concentration did not respond to an increasing P-application rate and remained at ca. 0.5 mg g⁻¹ DM across the entire range of P-application rates which was reflected in a significant P-application rate × AM inoculation interaction term (two-way ANOVA, $F_{3,22} = 7.66$, $P < 0.001$; Fig. 4B).

Discussion

For non-inoculated plants of *A. celastrifolia*, an increasing P-application rate increased DM up to high-level application rates (80 mg P kg⁻¹ soil). However, for non-inoculated plants of *E. marginata*, an increasing P supply increased growth only at relatively low external P concentrations: thereafter DM declined with increasing P supply. Similar contrasting patterns in response to increasing P supply have been observed previously for a range of Australian species from severely nutrient-impoor environments (Grundon 1972; Groves and Keraitis 1976; Handreck 1997; Pang et al. 2010; de Campos et al. 2013; Williams et al. 2019). For *E. marginata*, but not *A.*

celastrifolia, inoculation with AMF reduced growth at moderate P supply but facilitated growth at high P supply by regulating leaf P concentrations.

For non-inoculated *E. marginata*, maximum DM was observed at P-application rates of 15–30 mg kg⁻¹ soil, before declining at higher P-application rates. Maximum DM of a range of Australian natives has been reported to occur across a similar range of P supply (e.g. Bougher et al. 1990; Ryan et al. 2009; Pang et al. 2010; Williams et al. 2019). For example, for the 11 species studied by Pang et al. (2010), maximum growth occurred at P-application rates in the range 12–24 mg P kg⁻¹. Further, for 8 of the 11 species, growth declined at P-application rates greater than those required for maximum plant DM. Shane et al. (2004a) reported that, for a range of species, P toxicity occurred at leaf P concentrations of 0.9–47 mg g⁻¹ DM. The leaf P concentrations at which we observed negative effects on growth of *E. marginata* (>4 mg g⁻¹ DM) are at the lower end of these reported values. However, Williams et al. (2019) reported for *Eucalyptus torquata* that a reduction in growth occurred at leaf P concentrations > 2 mg g⁻¹ DM. Possible explanations for the differences between our current values and those reported by Shane et al. (2004a) are that either there are a wide range of species-specific thresholds for the onset of toxicity or that the values reported by Shane and co-workers are thresholds for the onset of visible symptoms

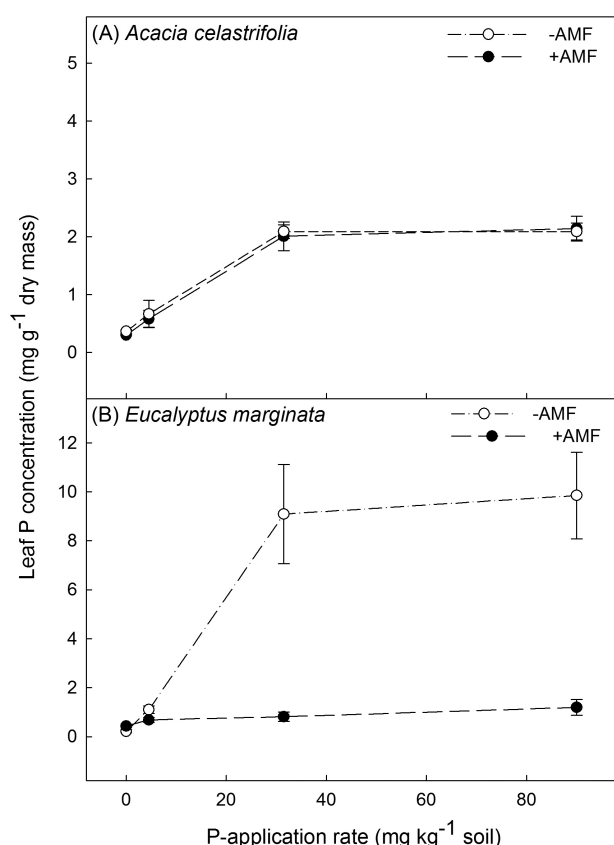


Figure 4. The effect of P-application rate and inoculation with the AMF *Rhizophagus irregularis* on leaf P concentration for *Acacia celastrifolia* and *Eucalyptus marginata* assessed 188 days after transplanting. Bars \pm 1 SE.

of P toxicity (e.g. necrosis). It is worth noting that our results and those of Williams *et al.* (2019) indicate the onset of a negative effect on plant growth and not necessarily the onset of visible symptoms.

Leaf P concentrations of non-inoculated *A. celastrifolia* initially increased with increasing P supply. However, even at higher P supply, leaf P concentrations did not exceed ca. 2 mg g⁻¹ DM. Similarly, Williams *et al.* (2019) reported that *Acacia acuminata* exhibited an initially increasing shoot P concentration in response to increasing P supply, but as P-application rates increased further, shoot P concentration was maintained at ca. 2 mg g⁻¹ DM. Conversely, *Acacia hemiteles* exhibited increasing shoot P with increasing P supply: shoot P reached concentrations of ca. 8 mg g⁻¹ DM, and growth reduced as shoot P concentration continued to increase (Williams *et al.* 2019). de Campos *et al.* (2013) also reported on the ability of two *Acacia* species (*Acacia truncata* and *Acacia xanthina*) to regulate internal P concentrations in relation to external P concentrations. *Acacia truncata* was unresponsive in terms of DM accumulation as the external P concentration increased, whilst *A. xanthina* exhibited declining DM as P increased. These results suggest that the growth response to elevated P, even within co-occurring members of a genus, can be unpredictable as also reported for the genus *Banksia* (de Campos *et al.* 2013).

We found low levels of early colonization by AMF in both species in 4 of the 12 samples examined. Similarly, low levels of colonization have been reported in previous studies with

Jarrah forest species. For example, Kariman *et al.* (2012) reported that although evidence of colonization by AMF of *E. marginata* in a pot experiment was found in as few as 2.3 % of samples, beneficial effects of AMF were still observed. Interestingly, even at low rates of intraradical colonization, AMF can still modulate the expression of phosphate transporters (Bulgarelli *et al.* 2020; Chu *et al.* 2020). Further, it should be noted that root colonization is not necessarily required for positive physiological responses in plant–fungus interactions (Neumann 1959; Kariman *et al.* 2014b). Similar results have also been reported for ECM colonization on seedlings of *Eucalyptus diversicolor* where in the absence of applied P, despite colonization rates on roots ranging from only 1 to 6 %, there was still a significant growth benefit for seedlings resulting from inoculation (Bougher *et al.* 1990).

For *A. celastrifolia*, plant DM was consistently, but not significantly, lower in the AM inoculation treatment across the entire range of P-application rates. While AMF can increase P uptake and result in increased growth (Smith *et al.* 2015), the transfer of carbon from the host plant to the AMF can also result in negative effects on plant growth. Indeed, this is further implied by the increased level of AMF sporulation observed at high P concentrations when in association with *A. celastrifolia*. In comparison, inoculation with AMF had three contrasting effects on DM of *E. marginata*.

First, at a low P-application rate (4.5 mg P kg⁻¹), at 53 days after planting, there was a positive classical growth effect of inoculation on DM. While inoculation with AMF did not increase leaf P concentrations it had increased total P uptake. Similarly, Kariman *et al.* (2014a) reported for 14-week-old *E. marginata* seedlings grown under P-deficient conditions that inoculation with the AMF *R. irregularis* did not increase leaf P concentration. In contrast, at low external P concentrations Jones *et al.* (1998) reported that inoculation with AMF both increased shoot P concentrations and growth of *Eucalyptus coccoifera*.

Second, at a moderate P-application rate (30 mg kg⁻¹), inoculation with AMF significantly depressed growth suggesting that at this supply, the association was parasitic rather than mutualistic. Indeed, for a range of species at elevated P concentrations, associations with AMF have been shown to move from mutualistic to parasitic (Johnson *et al.* 1997; Hoeksema *et al.* 2010; Johnson 2010).

Third, at high P-application rates, inoculation with AMF significantly increased DM, compared to non-inoculated plants, whilst maintaining leaf P concentrations within a similar range to that observed at lower external P concentrations. This is not a classical plant growth effect, rather a suppression of toxicity due to the symbiosis. We posit this mechanism is either related to a (down)regulation of root epidermal transporters which has been observed in AM plants for P, and at high concentrations for cadmium and putatively arsenic (De Oliveira *et al.* 2020; Kariman *et al.* 2014a; Nazeri *et al.* 2014; Kariman *et al.* 2016) or to a loss of function of the direct P-uptake pathway into roots colonized by AMF (Smith *et al.* 2003). Interestingly, for Jarrah, this effect of P-toxicity attenuation is occurring at high P availability when it is likely that there is increasing negative regulation of the AM symbiosis (e.g. Müller and Harrison 2019) which is supported by the declining spore counts we observed at high P. In nature, where plants are commonly mycorrhizal (Kariman *et al.* 2018), this may be a common mechanism whereby plants are protected from toxicities (at least to some extent) by mycorrhizal symbiosis.

Except for our control, nil P treatment, leaf P concentrations for both of our study species when non-inoculated were higher than concentrations previously reported in plants growing in relatively undisturbed and unfertilized Jarrah forest. For example, values of 0.4–0.45 mg P g⁻¹ DM have been reported for *E. marginata* (Hingston *et al.* 1981, M. I. Daws, unpublished data) and 0.3 mg P g⁻¹ DM for *A. celastriifolia* (M. I. Daws, unpublished data). For *E. marginata* inoculated with AMF, leaf P was maintained at concentrations similar to those observed in unfertilized forests across the full range of P-application rates. P is widely applied to newly established *E. marginata* stands following post-mining rehabilitation (Standish *et al.* 2015) and a recent study demonstrated significant ongoing impacts of applying P at the outset of rehabilitation on the re-establishing Jarrah forest plant community. Specifically, applied P shifts the balance between different plant functional groups favouring fast-growing legumes compared to long-lived understorey resprouter species (Daws *et al.* 2021) suggesting that P should be applied with caution. These results may have broader application to the use of fertilizer for rehabilitation of other biodiverse plant communities growing on P-deficient soils elsewhere (e.g. Tibbett *et al.* 2019). For rehabilitated Jarrah forest, investigating the role of AMF on plant growth and P uptake in the field would be of value, particularly since colonization of roots in newly established sites may be limited by the availability of propagules, e.g. spores, hyphae, colonized roots (Jasper *et al.* 2007). Arbuscular mycorrhizal fungus colonization could be manipulated, for example, by inoculating plants prior to planting in the field to act as nurse plants that accelerate the development of common mycorrhizal networks, facilitating fitness in plant species with high mycorrhizal dependency (Kariman *et al.* 2012; Montesinos-Navarro *et al.* 2019; Sortibrán *et al.* 2019; Standish *et al.* 2022).

Arbuscular mycorrhizal fungi are generally viewed as being important for increasing P uptake and facilitating growth at low external P concentrations. However, our data support a growing understanding that by regulating plant P concentration within a sufficient concentration range, AMF play an important role at high external P concentrations in enabling plant growth at concentrations that would otherwise result in reduced growth and P toxicity.

Supporting Information

The following additional information is available in the online version of this article—

Figure S1. Photograph of Jarrah (*Eucalyptus marginata*) seedlings from Experiment 2 (see Materials and Methods section) grown at a P-application rate of 90 mg kg⁻¹ soil. The pot on the left was not inoculated, whereas the pot on the right was inoculated with the arbuscular mycorrhizal fungus *Rhizophagus irregularis* at the commencement of the experiment. The photograph was taken at the end of the experiment (Day 188 after transplanting).

Acknowledgements

Our thanks to Matt Braimbridge, Russell Beazley, Anna Dudley, Tammy Edmonds-Tibbett, Bridget Kennedy, Henning Wallrabenstein, Chris Walker, Evonne Walker and Trudy Worthington for their contribution to this work.

Author Contributions

MT and MR conceived the study. MT designed the study. MD analysed the data. MT and MD interpreted the data and MD and MT wrote the manuscript. All authors contributed to the draft manuscript.

Conflict of Interest

The authors declare no conflict of interest.

Data Availability

Data are available at <https://doi.org/10.17864/1947.000399>.

Literature Cited

- Bougher NL, Grove TS, Malajczuk N. 1990. Growth and phosphorus acquisition of Karri (*Eucalyptus diversicolor* F. Muell.) seedlings inoculated with ectomycorrhizal fungi in relation to phosphorus supply. *New Phytologist* 114:77–85.
- Brundrett MC, Ashwath N, Jasper DA. 1996. Mycorrhizas in the Kakadu region of tropical Australia. II. Propagules of mycorrhizal fungi in disturbed habitats. *Plant and Soil* 184:173–184.
- Bulgarelli RG, De Oliveira VH, de Andrade SAL. 2020. Arbuscular mycorrhizal symbiosis alters the expression of PHT1 phosphate transporters in roots and nodules of P-starved soybean plants. *Theoretical and Experimental Plant Physiology* 32:243–253.
- Burleigh SH, Bechmann IE. 2002. Plant nutrient transporter regulation in arbuscular mycorrhizas. *Plant and Soil* 244:247–251.
- Cavagnaro T, Smith F, Lorimer M, Haskard K, Ayling S, Smith S. 2001. Quantitative development of Paris-type arbuscular mycorrhizas formed between *Asphodelus fistulosus* and *Glomus coronatum*. *New Phytologist* 149:105–113.
- Chu Q, Zhang L, Zhou JW, Yuan LX, Chen FJ, Zhang FS, Feng G, Rengel Z. 2020. Soil plant-available phosphorus levels and maize genotypes determine the phosphorus acquisition efficiency and contribution of mycorrhizal pathway. *Plant and Soil* 449:357–371.
- Daws MI, Standish RJ, Koch JM, Morald TK, Tibbett M, Hobbs RJ. 2015. Phosphorus fertilisation and large legume species affect Jarrah forest restoration after bauxite mining. *Forest Ecology and Management* 354:10–17.
- Daws MI, Walters SJ, Harris RJ, Tibbett M, Grigg AH, Morald TK, Hobbs RJ, Standish RJ. 2021. Nutrient enrichment diminishes plant diversity and density, and alters long-term ecological trajectories, in a biodiverse forest restoration. *Ecological Engineering* 165:106222.
- de Campos MCR, Pearse SJ, Oliveira RS, Lambers H. 2013. Downregulation of net phosphorus-uptake capacity is inversely related to leaf phosphorus-resorption proficiency in four species from a phosphorus-impooverished environment. *Annals of Botany* 111:445–454.
- De Oliveira VH, Ullah I, Dunwell JM, Tibbett M. 2020. Mycorrhizal symbiosis induces divergent patterns of transport and partitioning of Cd and Zn in *Populus trichocarpa*. *Environmental and Experimental Botany* 171:103925.
- Gardner JH, Malajczuk N. 1988. Recolonization of rehabilitated bauxite mine sites in Western Australia by mycorrhizal fungi. *Forest Ecology and Management* 24:27–42.
- Glen M, Bougher NL, Colquhoun IJ, Vlahos S, Loneragan WA, O'Brien PA, Hardy GE. 2008. Ectomycorrhizal fungal communities of rehabilitated bauxite mines and adjacent, natural Jarrah forest in Western Australia. *Forest Ecology and Management* 255:214–225.
- Grant CD, Ward SC, Morley C. 2007. Return of ecosystem function to restored bauxite mines in Western Australia. *Restoration Ecology* 15:S94–S103.
- Groves RH, Keraitis K. 1976. Survival of seedlings of three sclerophyll species at high levels of phosphorus and nitrogen. *Australian Journal of Botany* 24:681–690.

- Grundon NJ. 1972. Mineral nutrition of some Queensland heath plants. *Journal of Ecology* 60:171–181.
- Handreck KA. 1991. Interactions between iron and phosphorus in the nutrition of *Banksia ericifolia* L. f. var. *ericifolia* (Proteaceae) in soil-less potting media. *Australian Journal of Botany* 39:373–384.
- Handreck KA. 1997. Phosphorus requirements of Australian native plants. *Australian Journal of Soil Research* 35:241–289.
- Hildebrandt U, Regvar M, Bothe H. 2007. Arbuscular mycorrhiza and heavy metal tolerance. *Phytochemistry* 68:139–146.
- Hingston FJ, Dimmock GM, Turton AG. 1981. Nutrient distribution in a Jarrah (*Eucalyptus marginata* Donn ex Sin.) ecosystem in south-west Western Australia. *Forest Ecology and Management* 3:183–207.
- Hoeksema JD, Chaudhary VB, Gehring CA, Johnson NC, Karst J, Koide RT, Pringle A, Zabinski C, Bever JD, Moore JC, Wilson GWT, Klironomos JN, Umbanhowar J. 2010. A meta-analysis of context dependency in plant response to inoculation with mycorrhizal fungi. *Ecology Letters* 13:394–407.
- Huang CY, Shirley N, Genc Y, Shi S, Langridge P. 2011. Phosphate utilization efficiency correlates with expression of low-affinity phosphate transporters and noncoding RNA, *IPS1*, in barley. *Plant Physiology* 156:1217–1229.
- Jasper DA. 2007. Beneficial soil microorganisms of the Jarrah forest and their recovery in bauxite Southwestern Australia. *Restoration Ecology* 15:S74–S84.
- Jentschke G, Goldbold DL. 2000. Metal toxicity and ectomycorrhizas. *Physiologia Plantarum* 109:107–116.
- Johnson NC. 2010. Resource stoichiometry elucidates the structure and function of arbuscular mycorrhizas across scales. *New Phytologist* 185:631–647.
- Johnson NC, Graham JH, Smith FA. 1997. Functioning of mycorrhizal associations along the mutualism–parasitism continuum. *New Phytologist* 135:575–585.
- Jones MD, Durall DM, Tinker PB. 1998. A comparison of arbuscular and ectomycorrhizal *Eucalyptus coccifera*: growth response, phosphorus uptake efficiency and external hyphal production. *New Phytologist* 140:125–134.
- Kariman K, Barker SJ, Finnegan PM, Tibbett M. 2012. Dual mycorrhizal associations of Jarrah (*Eucalyptus marginata*) in a nurse-pot system. *Australian Journal of Botany* 60:661–668.
- Kariman K, Barker SJ, Finnegan PM, Tibbett M. 2014a. Ecto- and arbuscular mycorrhizal symbiosis can induce tolerance to toxic pulses of phosphorus in Jarrah (*Eucalyptus marginata*) seedlings. *Mycorrhiza* 24:501–509.
- Kariman K, Barker SJ, Jost R, Finnegan PM, Tibbett M. 2014b. A novel plant–fungus symbiosis benefits the host without forming mycorrhizal structures. *New Phytologist* 201:1413–1422.
- Kariman K, Barker SJ, Jost R, Finnegan PM, Tibbett M. 2016. Sensitivity of Jarrah (*Eucalyptus marginata*) to phosphate, phosphite, and arsenate pulses as influenced by fungal symbiotic associations. *Mycorrhiza* 26:401–415.
- Kariman K, Barker SJ, Tibbett M. 2018. Structural plasticity in root–fungal symbioses: diverse interactions lead to improved plant fitness. *PeerJ* 6:e6030.
- Koide RT, Mosse B. 2004. A history of research on arbuscular mycorrhiza. *Mycorrhiza* 14:145–163.
- Lambers H, Juniper D, Cawthray GR, Veneklaas EJ, Martinez-Ferri E. 2002. The pattern of carboxylate exudation in *Banksia grandis* (Proteaceae) is affected by the form of phosphate added to the soil. *Plant and Soil* 238:111–122.
- Lambers H, Raven JA, Shaver GR, Smith SE. 2008. Plant nutrition-acquisition strategies change with soil age. *Trends in Ecology and Evolution* 23:95–103.
- Lambers H, Shane MW, Cramer MD, Pearse SJ, Veneklaas EJ. 2006. Root structure and functioning for efficient acquisition of phosphorus: matching morphological and physiological traits. *Annals of Botany* 98:693–713.
- Montesinos-Navarro A, Valiente-Banuet A, Verdú M. 2019. Mycorrhizal symbiosis increases the benefits of plant facilitative interactions. *Ecography* 42:447–455.
- Müller LM, Harrison MJ. 2019. Phytohormones, miRNAs, and peptide signals integrate plant phosphorus status with arbuscular mycorrhizal symbiosis. *Current Opinion in Plant Biology* 50:132–139.
- Nazeri NK, Lambers H, Tibbett M, Ryan MH. 2014. Moderating mycorrhizas: arbuscular mycorrhizas modify rhizosphere chemistry and maintain plant phosphorus status within narrow boundaries. *Plant, Cell and Environment* 37:911–921.
- Neumann R. 1959. Relationships between *Pisolithus tinctorius* (Mich. ex Pers.) Coker et Couch. and *Eucalyptus camaldulensis* [rostrata] Dehn. *Bulletin of the Research Council of Israel* 7D:116.
- Pang JY, Tibbett M, Denton MD, Lambers H, Siddique KHM, Bolland MDA, Revell CK, Ryan MH. 2010. Variation in seedling growth of 11 perennial legumes in response to phosphorus supply. *Plant and Soil* 328:133–143.
- Reddell P, Milnes AR. 1992. Mycorrhizas and other specialized nutrient-acquisition strategies: their occurrence in woodland plants from Kakadu and their role in rehabilitation of waste rock dumps at a local uranium mine. *Australian Journal of Botany* 40:223–242.
- Ryan MH, Ehrenberg S, Bennett RG, Tibbett M. 2009. Putting the P in *Ptilotus*: a phosphorus-accumulating herb native to Australia. *Annals of Botany* 103:901–911.
- Simmons W. 1975. Determination of low concentrations of cobalt in small samples of plant material by flameless atomic absorption spectrophotometry. *Analytical Chemistry* 47:2015–2018.
- Simmons W. 1978. Background absorption error in determination of copper in plants by flame atomic absorption spectrometry. *Analytical Chemistry* 50:870–873.
- Shane MW, McCully ME, Lambers H. 2004a. Tissue and cellular phosphorus storage during development of phosphorus toxicity in *Hakea prostrata* (Proteaceae). *Journal of Experimental Botany* 55:1033–1044.
- Shane MW, Szota C, Lambers H. 2004b. A root trait accounting for the extreme phosphorus sensitivity of *Hakea prostrata* (Proteaceae). *Journal of Experimental Botany* 27:991–1004.
- Smith SE, Anderson IC, Smith FA. 2015. Mycorrhizal associations and phosphorus acquisition: from cells to ecosystems. In: Plaxton WC, Lambers H, eds. *Annual plant reviews*, vol. 48. Hoboken, NJ: John Wiley & Sons Inc., 409–439.
- Smith SE, Jakobsen I, Gronlund M, Smith FA. 2011. Roles of arbuscular mycorrhizas in plant phosphorus nutrition: interactions between pathways of phosphorus uptake in arbuscular mycorrhizal roots have important implications for understanding and manipulating plant phosphorus acquisition. *Plant Physiology* 156:1050–1057.
- Smith SE, Smith FA, Jakobsen I. 2003. Mycorrhizal fungi can dominate phosphate supply to plants irrespective of growth responses. *Plant Physiology* 133:16–20.
- Sortibrán L, Verdú M, Valiente-Banuet A. 2019. A nurse plant benefits from facilitative interactions through mycorrhizae. *Plant Biology* 21:670–676.
- Standish RJ, Daws MI, Gove AD, Didham RK, Grigg AH, Koch JM, Hobbs RJ. 2015. Long-term data suggest Jarrah-forest establishment at restored mine sites is resistant to climate variability. *Journal of Ecology* 103:78–89.
- Standish RJ, Daws MI, Morald TK, Speijers J, Koch JM, Hobbs RJ, Tibbett M. 2022. Phosphorus supply affects seedling growth of mycorrhizal but not cluster-root forming Jarrah-forest species. *Plant and Soil* 472:577–594.
- Standish RJ, Stokes BA, Tibbett M, Hobbs RJ. 2007. Seedling response to phosphate addition and inoculation with arbuscular mycorrhizas and the implications for old-field restoration in Western Australia. *Environmental and Experimental Botany* 61:58–65.
- Tibbett M. 2000. Roots, foraging and the exploitation of soil nutrient patches: the role of mycorrhizal symbiosis. *Functional Ecology* 14:397–399.
- Tibbett M. 2010. Large-scale mine site restoration of Australian eucalypt forests after bauxite mining: soil management and ecosystem development. In: Batty LC, Hallberg K, eds. *Ecology of industrial pollution*. Cambridge, UK: Cambridge University Press, 309–326.

- Tibbett M, Daws MI, George SJ, Ryan MH. 2020. The where, when and what of phosphorus fertilisation for seedling establishment in a biodiverse Jarrah forest restoration after bauxite mining in Western Australia. *Ecological Engineering* 153:105907.
- Tibbett M, O'Connor R, Daws MI. 2019. Too much of a good thing: phosphorus over-fertilisation in rehabilitated landscapes of high biodiversity value. In: Fourie AB, Tibbett M, eds. *Proceedings of the 13th International Conference on Mine Closure*. Perth, Australia: Australian Centre for Geomechanics, 651–666.
- Tibbett M, Sanders FE. 2002. Ectomycorrhizal symbiosis can enhance plant nutrition through improved access to discrete organic nutrient patches of high resource quality. *Annals of Botany* 89:783–789.
- Walker C, Mize CW, McNabb HS Jr. 1982. Populations of endogonaceous fungi at two locations in central Iowa. *Canadian Journal of Botany* 60:2518–2529.
- Ward SC, Koch JM, Nichols OG. 1990. Bauxite mine rehabilitation in the Darling Range, Western Australia. *Proceedings of the Ecology Society of Australia* 16:557–565.
- Williams A, George S, Birt HWG, Daws MI, Tibbett M. 2019. Sensitivity of seedling growth to phosphorus supply in six tree species of the Australian Great Western Woodlands. *Australian Journal of Botany* 67:390–396.
- Yazici MA, Asif M, Tutus Y, Ortas I, Ozturk L, Lambers H, Cakmak I. 2021. Reduced root mycorrhizal colonization as affected by phosphorus fertilization is responsible for high cadmium accumulation in wheat. *Plant and Soil*. doi:[10.1007/s11104-021-05041-5](https://doi.org/10.1007/s11104-021-05041-5).
- Ye D, Clode PL, Hammer TA, Pang J, Lambers H, Ryan MH. 2021. Accumulation of phosphorus and calcium in different cells protects the phosphorus-hyperaccumulator *Ptilotus exaltatus* from phosphorus toxicity in high-phosphorus soils. *Chemosphere* 264:128438.