

Resilience of tropical invertebrate community assembly processes to a gradient of land use intensity

Article

Published Version

Creative Commons: Attribution 4.0 (CC-BY)

Open Access

Granville, N. R. ORCID: https://orcid.org/0000-0002-9031-9888, Barclay, M. V. L., Boyle, M. J. W., Chung, A. Y. C., Fayle, T. M. ORCID: https://orcid.org/0000-0002-1667-1189, Hah, H. E., Hardwick, J. L., Kinneen, L. ORCID: https://orcid.org/0000-0001-5502-8113, Kitching, R. L., Maunsell, S. C., Miller, J. A., Sharp, A. C., Stork, N. E., Wai, L., Yusah, K. M. and Ewers, R. M. (2024) Resilience of tropical invertebrate community assembly processes to a gradient of land use intensity. Oikos, 2024 (2). e10328. ISSN 1600-0706 doi: 10.1111/oik.10328 Available at https://centaur.reading.ac.uk/114707/

It is advisable to refer to the publisher's version if you intend to cite from the work. See <u>Guidance on citing</u>.

To link to this article DOI: http://dx.doi.org/10.1111/oik.10328

Publisher: Wiley

All outputs in CentAUR are protected by Intellectual Property Rights law, including copyright law. Copyright and IPR is retained by the creators or other copyright holders. Terms and conditions for use of this material are defined in



the End User Agreement.

www.reading.ac.uk/centaur

CentAUR

Central Archive at the University of Reading Reading's research outputs online



Research article

Resilience of tropical invertebrate community assembly processes to a gradient of land use intensity

Natasha R. Granville[®] 1, Maxwell V. L. Barclay², Michael J. W. Boyle^{1,3,4}, Arthur Y. C. Chung⁵, Tom M. Fayle[®] 6,7</sup>, Huai En Hah⁸, Jane L. Hardwick⁹, Lois Kinneen¹⁰, Roger L. Kitching¹¹, Sarah C. Maunsell⁹, Jeremy A. Miller¹², Adam C. Sharp¹, Nigel E. Stork¹¹, Leona Wai⁸, Kalsum M. Yusah¹³ and Robert M. Ewers¹

Correspondence: Natasha R. Granville (natasha.granville19@imperial.ac.uk)

Oikos 2023: e10328

doi: 10.1111/oik.10328

Subject Editor: Paul Caplat Editor-in-Chief: Pedro Peres-Neto Accepted 2 November 2023





www.oikosjournal.org

Understanding how community assembly processes drive biodiversity patterns is a central goal of community ecology. While it is generally accepted that ecological communities are assembled by both stochastic and deterministic processes, quantifying their relative importance remains challenging. Few studies have investigated how the relative importance of stochastic and deterministic community assembly processes vary among taxa and along gradients of habitat degradation. Using data on 1645 arthropod species across seven taxonomic groups in Malaysian Borneo, we quantified the importance of ecological stochasticity and of a suite of community assembly processes across a gradient of logging intensity. The relationship between logging and community assembly varied depending on the specific combination of taxa and stochasticity metric used, but, in general, the processes that govern invertebrate community assembly were remarkably robust to changes in land use intensity.

Keywords: community assembly, determinism, habitat degradation, logging, stochasticity

¹Georgina Mace Centre for the Living Planet, Imperial College London, Ascot, UK

²Natural History Museum, London, UK

³School of Biological Sciences, University of Hong Kong, Hong Kong

⁴Department of Biological Sciences, National University of Singapore, Singapore

⁵Forest Research Centre, Department of Sabah Forestry, Sandakan, Sabah, Malaysia

⁶School of Biological and Behavioural Sciences, Queen Mary University of London, London, UK

⁷Biology Centre of the Czech Academy of Sciences, Institute of Entomology, Ceske Budejovice, Czech Republic

⁸Institute for Tropical Biology and Conservation, Universiti Malaysia Sabah, Jalan UMS, Kota Kinabalu, Sabah, Malaysia ⁹Griffith University, Nathan, OLD, Australia

¹⁰Centre for Agri-Environmental Research, School of Agriculture, Policy and Development, University of Reading, Reading, UK

¹¹Centre for Planetary Health and Food Security, Griffith University, Nathan, QLD, Australia

¹²Understanding Evolution Research Group, Naturalis Biodiversity Center, Leiden, the Netherlands

¹³Royal Botanic Gardens, Kew, Richmond, London, UK

^{© 2023} The Authors. Oikos published by John Wiley & Sons Ltd on behalf of Nordic Society Oikos.

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

Introduction

Community assembly processes drive biodiversity patterns, and a key goal in community ecology is to quantify the relative importance of different community assembly processes. Currently, we have a strong awareness of the patterns of biodiversity change that are generated by land use change (Newbold et al. 2015), but little understanding of the extent to which the fundamental community assembly processes that create that change are impacted. Moreover, the specific assembly processes may vary among taxa due to differences in trait evolution (Weiher et al. 2011), meaning studies examining land use impacts on the assembly processes of mammals (Wearn et al. 2019) may provide little insight into the impacts on other taxa. Attempts to rely on natural ecological processes to restore biodiversity rely, by definition, on naturally occurring community assembly processes (Palmer et al. 1997, Hilderbrand et al. 2005). It is therefore of fundamental importance that we gain a deeper understanding of whether those assembly processes in modified habitats are the same or different to those observed in primary habitats (Hallet et al.

Community assembly involves a combination of determinism and stochasticity and there has been long-standing debate over their relative influences (Connor and Simberloff 1979, Chase and Myers 2011). Stochastic assembly generates community diversity patterns indistinguishable from those generated by random chance alone (Hubbell 2001, Chase and Myers 2011, Ning et al. 2019) and can involve random variation around species average demographic rates due to the probabilistic nature of demographic processes like birth, death and migration (Adler et al. 2007, Shoemaker et al. 2020). Deterministic community assembly, on the other hand, involves non-random, niche-based processes such as environmental filtering and biotic interactions (Chase and Myers 2011). Determinism and stochasticity are opposite ends of a continuum, with real-world communities existing somewhere between these extremes (Gravel et al. 2006, Kitching 2013). Previous studies have investigated the balance between stochasticity and determinism in different environments (Ellwood et al. 2009, Caruso et al. 2012, Shipley et al. 2012, Ortega-Martínez et al. 2020, Valdivia et al. 2021), but few have addressed the impact of land use change on this balance, despite the potential importance of this. It is often suggested that community assembly processes exhibit hysteresis (Beisner et al. 2003), implying that restoring an ecological community in a modified habitat could rely on different assembly processes than those that exist in primary habitats (Andersen et al. 2009, Suding and Hobbs 2009). Therefore, our understanding of primary community assembly may give information that at best is irrelevant, or at worst directly misleading, when planning the restoration of modified communities (Hallett et al. 2023).

Logging is a major driver of habitat degradation across many of the world's most productive and biodiverse tropical forest ecosystems (Laurance 2015). The tropical forests of Borneo have been subject to rapid and widespread logging

since the early 1970s. Between 1973 and 2010, there was an estimated 30% decline in the extent of Borneo's intact forests (Gaveau et al. 2014). Logging often results in a heterogeneous landscape, with habitat patches connected spatially but affected by different logging intensities (Berry et al. 2008). This can result in gradients of disturbance intensity, which are a frequent consequence of land use change in the tropics (Wearn et al. 2019).

There is uncertainty over how logging might affect stochastic and deterministic community assembly processes, with little in the way of direct evidence. Kitching et al. (2013) showed a distance-turnover relationship (decreasing community similarity with increasing geographic distance) for moth communities in primary forest, but such patterns were largely absent in logged forests, suggesting that stochastic turnover in plant species can drive deterministic changes in the niche dimensions available for moths in primary forests, but not in logged forests. Döbert et al. (2017) showed that understorey plant communities in tropical Bornean forests tend to be more stochastically assembled at higher logging intensities, whereas Wearn et al. (2019) showed that deterministic environmental control on mammal community assembly is higher in logged forest compared to old-growth forest. There is broad conceptual and empirical support for deterministic environmental filtering on community assembly becoming more important in harsher environmental conditions (Chase 2007, Lepori and Malmqvist 2009, Chase and Myers 2011, Ding et al. 2012, Wearn et al. 2019, Li et al. 2021, Hu et al. 2022). We therefore hypothesise that stochasticity will become less important with increasing logging intensity.

There is further uncertainty over how taxonomic groups might differ in their community assembly processes, with broad support for vastly different taxonomic groups showing variation in the balance between stochasticity and determinism. For example, Powell et al. (2015) found substantial differences in the assembly of soil bacterial and fungal communities, due to differences in dispersal capacity between these groups. Trophic position is also likely to play a role in mediating these differences between taxonomic groups; Keppeler et al. (2016) indicated differences in assembly processes between foraging bird and fish communities. Among comparatively more similar taxa, Thompson and Townsend (2006) showed that trophic groupings and species traits determine the relative importance of stochasticity and determinism in macroinvertebrate communities. Generalist taxa have been shown to contribute more to stochastic processes, and specialist taxa to contribute more to deterministic processes in microbial communities, because specialists tend to have narrower tolerance to environmental changes (Liao et al. 2016, Xu et al. 2022). Similarly, studies on microbial communities have also shown that rare taxa, which have narrower niche breadths, are more deterministically assembled while abundant taxa with wider niche breadths are more stochastically assembled (Gao et al. 2020, Yang et al. 2022). There has not, to our knowledge, been a similar study directly quantifying among-taxa variation in stochastic and deterministic assembly processes for invertebrate communities.

Stochasticity indices have been widely used in studies on microbial communities to quantify the importance of ecological stochasticity (Jiao et al. 2020, Le Moigne et al. 2020, Sun et al. 2021, Trego et al. 2021, Zhou et al. 2022, Wang et al. 2022). These indices compare observed community dissimilarity to the neutral expectation. Neutral models generally assume that all individuals within a feeding guild have equal chances of birth, death and migration, regardless of their taxonomic identity (Hubbell 2001). Significant deviations from neutral models of random community assembly can indicate that other processes, such as selection, are involved in structuring the community (Burns et al. 2016). The relative importance of classes of community assembly processes can be inferred from patterns of taxonomic and phylogenetic diversity (Ning et al. 2020). Phylogenetic and taxonomic turnover can indicate whether assembly processes are driving communities to be more heterogeneous (high turnover) or homogeneous (low turnover), and comparing these patterns with neutral expectation can indicate the relative importance of stochasticity (Ning et al. 2020).

There is a long-standing need to evaluate the relative importance of stochastic and deterministic processes along environmental gradients and among taxa (Weiher et al. 2011). Here, we address that knowledge gap by using a variety of indices to quantify the relative contribution of stochasticity to community assembly for seven invertebrate taxa across a gradient of logging intensity. We also quantify the relative importance of a suite of community assembly processes across the logging gradient. Our data encompass a comprehensive gradient of logging intensity, from areas that have never experienced logging to areas that have been salvage logged. We quantified community assembly for a range of invertebrate taxa including three groups of Coleoptera (beetles), along with Formicidae (ants), Lepidoptera (moths), Araneae (spiders) and Orthoptera. Together, these taxonomic groups encompass a range of feeding guilds and are of immense ecological importance (Barlow and Woiwod 1989, Didham et al. 1998, Grimaldi and Engel 2005, Nyffeler and Birkhofer 2017, Oumarou Ngoute et al. 2020). We use our data to test two hypotheses: 1) stochastic assembly will decrease in importance as logging intensity increases, as logged forest should have stronger environmental filtering; and 2) stochastic assembly will have a lower relative importance for trophic specialists, compared to trophic generalists, because trophic specialists are expected to be more strongly assembled by selective environmental filtering. Finally, we investigate whether the relative importance of a suite of different community assembly processes varies across a gradient of logging intensity for different invertebrate taxa.

Material and methods

Data collection

The study sites were located within the Stability of Altered Forest Ecosystems (SAFE) project (4°38′N–04°46′N,

116°57′E–117°42′E), a large-scale ecological experiment encompassing a gradient of land use intensities in the lowland tropical forests of Sabah, Malaysian Borneo (Ewers et al. 2011). We used data from 14 out of the 17 experimental sampling blocks at SAFE (Ewers et al. 2011), excluding three blocks located in oil palm plantation. Ten sampling blocks were located in twice-logged forests and four were located in protected areas. Two of these protected area blocks were in the Maliau Basin Conservation area and have never experienced logging, while the other two had experienced light logging through both legal and illegal processes. Each sampling block comprised a set of 4-43 sampling sites (mean = 19) and covered a spatial area of 4-229 ha (mean = 56). We grouped invertebrate samples collected within each block which we considered as one local community for analysis. The aggregation of all local communities across all sampling blocks was considered to represent the metacommunity.

Above-ground carbon density (ACD), calculated from LiDAR surveys and summarized at 1 ha resolution in 2014, was used to quantify logging intensity (Jucker et al. 2018, Swinfield et al. 2020): a higher ACD corresponds to a lower logging intensity. ACD was log-transformed to generate a more uniform spread of logging intensity values. The sampling blocks covered a wide range of logging intensities, with average above-ground carbon densities ranging from around 15 t C ha⁻¹ in heavily logged locations, to over 200 t C ha⁻¹ in the protected areas. Not all invertebrate taxa were sampled at the same subset of sampling points per block. For analysis, then, we calculated the average ACD per sampling block separately for each taxon, taking as inputs the ACD values for the specific subset of sampling sites where that taxon was collected.

We combined community composition data collected from seven invertebrate taxa: three groups of beetles, plus ants, moths, spiders and Orthoptera. Different groups had different sample sizes and not all groups were sampled in all 14 sampling blocks (Supporting information). Beetles were sampled between 2011 and 2013 using combination pitfall-malaise traps in all 14 sampling blocks. Three different groups of beetles were sampled: Curculionoidea (weevils), Staphylinidae (rove beetles) and Scarabaeoidea (scarabs) (Sharp et al. 2018, 2019). Because of differences in their feeding guilds, each group was considered a separate taxon and was analysed separately: weevils are predominantly herbivorous; most scarabs in our dataset are dung-feeders; and rove beetles can belong to several feeding guilds. Beetles were identified primarily to morphospecies, except some scarabs which were identified to species. Ants were sampled between December 2011 and June 2012 in 12 sampling blocks using 12 × 14 cm plastic cards, which were laid flat in the leaf litter and baited with 30 compressed dried earthworm pellets. The number of ants entering each card was observed and recorded for 40 min, and individuals were identified to morphospecies (Fayle et al. 2019). Moths were sampled in 2014 using UV light traps which were run overnight in eight sampling blocks. Moths were identified where possible and separated into morphospecies using morphology (Maunsell et al.

2020b). Spider abundance data were collected in 2015 in 10 sampling blocks by beating plant foliage for 20 min at each site. Spiders were identified to family, then separated into morphospecies by genitalia dissections and DNA barcoding using the CO1 gene (Maunsell et al. 2020a). Finally, Orthoptera were sampled in 2015 by sweep netting along 100 m transects in 6 sampling blocks. Orthoptera were identified to family, then separated into morphospecies using identification guides (Hardwick et al. 2022).

Stochasticity metrics

There are different ways for communities to express stochasticity, so there is value in assessing multiple metrics of stochasticity on the same communities (Vellend 2010). We quantified stochasticity using three null-model based mathematical frameworks that summarise stochasticity both at the community level and at the level of individual species. All three stochasticity metrics were calculated from a separate community composition (site × species) matrix for each of the seven taxa. We calculated stochasticity metrics for all sites in the composition matrix, and grouped sites together by sampling block to calculate the mean of each stochasticity metric for each sampling block.

First, we employed the normalised stochasticity ratio (NST) (Ning et al. 2019) to assess the relative importance of ecological stochasticity. NST values are normalised on a scale from 0 to 1, with 0.5 as the boundary between more stochastic (> 0.5) and more deterministic (< 0.5) community assembly. NST is based on pairwise community dissimilarity measures, for which there are many competing metrics. We used Ružička dissimilarity which was shown to have the highest accuracy and precision when the NST was developed (Ning et al. 2019). NST compares the observed dissimilarity of the real community with the null expected dissimilarity for 1000 randomised communities (Ning et al. 2019). To generate the random metacommunities for the null expectation, the total number of species was fixed as observed, and within each local community, individuals were drawn at random from the metacommunity with probabilities proportional to their regional occurrence frequencies (Ning et al. 2019).

Second, to test the effect of different assumptions in the null modelling framework, we also used the modified sto-chasticity ratio (MST) which transforms NST under the assumption that observed community similarity is equal to the mean of the null expected community similarity under stochastic assembly. MST is also normalised from 0 to 1 with values closer to 1 indicating a higher importance of ecological stochasticity.

The NST and MST metrics assess ecological stochasticity at the community level, whereas our third metric, the neutral taxa percentage (NTP) assesses stochasticity at the level of individual species (Burns et al. 2016). NTP is the proportion of species with occurrence frequencies that could be predicted by Sloan's neutral model (Sloan et al. 2006, 2007). This model assumes that assembly is driven solely by chance and dispersal. It predicts the relationship between the occurrence

frequencies of taxa in a set of local communities and their abundances across the metacommunity (Sloan et al. 2006, 2007, Burns et al. 2016). The model is fitted to the frequency and abundance of species in a metacommunity by a single parameter which describes the migration rate (the probability that loss of an individual in a local community will be replaced by dispersal from the metacommunity rather than by reproduction within the local community) (Sloan et al. 2006, 2007, Burns et al. 2016). 'Neutral taxa' are those whose observed occurrence frequencies are within one confidence interval of that which would be expected by Sloan's neutral model (Burns et al. 2016). The proportion of neutral taxa was weighted according to the abundance of individuals in each taxon, and was used to assess the relative importance of ecological stochasticity at the level of individual species (Burns et al. 2016).

Community assembly processes

To further characterise the drivers of community assembly across the logging gradient, we quantified the relative importance of a suite of assembly processes: heterogeneous selection, homogeneous selection, dispersal limitation, homogenising dispersal and drift (Stegen et al. 2013, Ning et al. 2020). Heterogeneous selection is selection under heterogeneous abiotic and biotic conditions that leads to high phylogenetic compositional variation among local communities. Homogeneous selection takes place under homogeneous conditions and leads to low phylogenetic compositional variation. Dispersal limitation refers to the situation where low levels of dispersal among local communities constrains the exchange of organisms between these communities, leading to high levels of spatial turnover. Homogenising dispersal is the opposite situation where high levels of dispersal result in little turnover among communities. Communities that are not dominated by selection or dispersal are designated as being governed by drift (Stegen et al. 2013, Ning et al. 2020).

The proportional contribution of each of these five community assembly mechanisms was inferred through phylogenetic bin-based null model analysis (iCAMP) (Stegen et al. 2013, Ning et al. 2020). The iCAMP framework divides taxa into groups ('bins') based on their phylogenetic relationships, then identifies the dominant community assembly process in each bin. Phylogenies were not available for all of the taxa included in this study, so we obtained taxonomic hierarchies from the global biodiversity information facility (GBIF) using the safedata package (Aldersley and Orme 2023) which we used as proxies for phylogenies. The taxonomic hierarchies include the eight 'backbone' taxonomic ranks used in the GBIF database: Kingdom, Phylum, Class, Order, Family, Genus, Species and Subspecies. For the 1645 (morpho)species in our study, 13% were identified to species level, 31% were identified to genus level, 32% were identified to family level and 22% were identified to order level. Taxa were divided into bins based on taxonomic identity (Supporting information), and bins with fewer than the minimum number of taxa (n=9) were merged into the bin to which they were most closely related (Ning et al. 2020). The number of randomisations used for the null model analysis was 500. Details of the taxonomic hierarchies and bins can be found in the Supporting information.

To calculate the relative importance of community assembly processes, iCAMP uses null model analysis of phylogenetic diversity and taxonomic β-diversity to identify the process governing each bin. The phylogenetic and taxonomic diversity indices are calculated for pairwise comparisons between sites within phylogenetic bins. Phylogenetic diversity is quantified as the beta net relatedness index (βNRI), which is a measure of the mean pairwise phylogenetic distance between species in each site (Ning et al. 2020). Taxonomic β -diversity is quantified as the modified Raup-Crick index (RC) which measures the pairwise dissimilarity in species composition between sites (Chase et al. 2011, Ning et al. 2020). According to the framework developed by Ning et al. (2020); for each bin, the fraction of pairwise comparisons between sites with β NRI < -1.96 was considered as the percentage contribution of pairwise comparisons dominated by homogeneous selection, and those with $\beta NRI > +1.96$ as the percentage contribution of heterogeneous selection. Pairwise comparisons with $-1.96 \le \beta NRI \le +1.96$ were further partitioned using RC. The fraction of pairwise comparisons between sites with RC < -0.95 indicates the percentage contribution of homogenising dispersal, while those with RC > +0.95 indicates the percentage contribution of dispersal limitation. The remaining pairwise comparisons with $\beta NRI \leq |1.96|$ and $RC \leq |0.95|$ were considered to represent the contribution of drift, which includes ecological drift and other processes such as diversification, weak selection and weak dispersal (Ning et al. 2020). For each of the seven groups of taxa, we used iCAMP to calculate the relative importance of each process for all pairwise comparisons between all sites (Ning et al. 2020). We then grouped sites by sampling block and calculated the mean relative importance of each process for each taxa × sampling block combination. The methodological workflow is illustrated in the Supporting information.

Statistical analysis

All data analysis was conducted using R ver. 4.3.1(www.r-project.org). NST and MST stochasticity metrics were calculated using the 'NST' package (Ning et al. 2019), while NTP and iCAMP metrics were calculated using the 'iCAMP' package (Ning et al. 2020).

To see how species identity changed across the gradient of logging intensity, we compared the Bray–Curtis similarity of each sampling block to old-growth forest using the 'vegan' package (Oksanen et al. 2022), and used beta regression (Cribari-Neto and Zeileis 2010) to assess the effect of ACD on Bray–Curtis similarity.

We calculated the mean of each stochasticity metric (NST, MST and NTP) and of each community assembly process (heterogeneous selection, homogeneous selection, dispersal limitation, homogenising dispersal and drift) for each taxa × sampling block combination. We estimated the 95%

quantiles of these means using bootstrapping; we sampled the values used to calculate the mean 1000 times with replacement, then took the 5th and 95th quantiles of this distribution. To compare the relative importance of stochasticity among taxa we used one-sample t-tests to compare the overall unweighted mean NST, MST and NTP across all taxa to 0.5, which we used as a boundary point separating stochastic (> 0.5) from deterministic (< 0.5) community assembly. We also used ANOVA to test for differences in NST, MST and NTP among taxa.

To test the hypothesis that trophic generalists (ants and rove beetles) would be more stochastic than trophic specialists (moths, Orthoptera, spiders, scarabs and weevils), we used beta regression with a single categorical predictor that describes whether the taxon is considered a trophic generalist or specialist. We fitted three separate beta generalised mixed effect models, each with a stochasticity metric as the response variable. We included random intercepts for species richness and number of samples (number of sites in each sampling block) to account for variation in sampling among taxa. We broadly categorised each taxonomic group into trophic generalists or specialists based on trophic level. Ants were considered generalists because their diets can range from almost herbivorous to omnivorous and fully predatory (Blüthgen et al. 2004). Many rove beetles are generalist predators, though some can belong to other feeding guilds (Méndez-Rojas et al. 2021), so we also classified rove beetles as trophic generalists. The Orthoptera and weevils included in our data were mainly herbivorous (Sharp et al. 2019, Hardwick et al. 2022), so we classified them as trophic specialists. Spiders were all predatory (Russell-Smith and Stork 1995), scarabs were primarily coprophagous (Sharp et al. 2018) and moths were herbivorous as larvae and nectarivorous as adults (Romeis et al. 2005), so we classified these groups as trophic specialists for our analysis.

To investigate how each of the three metrics of stochasticity were affected by logging intensity, and how this relationship varies among the seven taxonomic groups, we employed beta generalised mixed effect models with logit link functions using the 'glmmTMB' package (Brooks et al. 2023). The models included the mean ACD of each block and the taxonomic groups as interacting predictors of the stochasticity metric (e.g. NST \sim ACD \times taxa). The community assembly metrics are based on taxonomic and phylogenetic diversity, so they could be influenced by changes in species richness, which can change systematically among taxa and along logging gradients (Burivalova et al. 2014). Therefore, the beta generalised mixed effect models included the random effects (1|richness) and (1|number of samples) to account for variation in species richness and sampling effort of different taxa X sampling block combinations. To gain an overall, taxa-independent indication of how logging affected the stochasticity metrics, we also fitted another model for each stochasticity metric with ACD as a single predictor of the stochasticity metric, with the same random effects as described above.

We used iCAMP to identify the dominant community assembly processes for each taxonomic group, then we used

the same beta generalised mixed effect model structure as described above, with the assembly process as a response variable, to investigate how each dominant process varied with ACD for different taxa, accounting for variation in sampling.

To gain a taxa-independent metric reflecting the overall relative importance of the five community assembly processes across the logging gradient, we combined the mean relative importance of each of the five processes and each of the three stochasticity metrics to give a weighted summary mean for all taxa using the following method adapted from Borenstein et al. (2010):

For each group of taxa (*i*), the weight (*w*) is the inverse of the variance for that group $w_i = \frac{1}{v_i}$. The variance (*v*) is the range of the 95% confidence intervals. \overline{M} is the overall weighted mean for all groups of taxa combined, it is analo-

gous to the combined effect size in a meta-analysis.
$$\overline{M}$$
 is calculated as $\overline{M} = \frac{\sum_{i=1}^{k} w_i m_i}{\sum_{i=1}^{k} w_i}$, where m is the mean relative importance for each group of taxa, analogous to the effect size of each study in a meta-analysis, and w is the weight assigned

of each study in a meta-analysis, and w_i is the weight assigned to group i, and k is the number of groups of taxa (k=7).

Results

Across all datasets, our analysis included 32 294 individuals belonging to 1645 species or morphospecies (Supporting information). In general, community similarity between sampling blocks was low. Each of the three beetle taxa, as well as moths, showed a decrease in community similarity compared to old growth forest as logging intensity increased, whereas the remaining three taxa (ants, Orthoptera and spiders) did not (Supporting information).

Stochasticity metrics

The overall average NST across all taxa was 0.49 (95%) CI = 0.44, 0.54), which was not significantly different from 0.5 (one-sample t-test, t = -0.30, df = 76, p = 0.76). However, the MST metric indicated that determinism plays a greater role in structuring invertebrate communities than ecological stochasticity (MST: mean = 0.28; CI = 0.22, 0.33; t=-8.19, df=76, p < 0.0001). In contrast, NTP indicated a greater role of stochasticity (NTP: mean = 0.68; CI = 0.64, 0.71; t=10.1, df=63, p < 0.0001). When the overall average NST was weighted by the variance of each taxon, it reduced to 0.39 (CI = 0.07, 0.83) and MST reduced to 0.10 (CI = 0.01, 0.75). The unweighted average NTP was not significantly different to the weighted average NTP (0.69, CI = 0.43, 0.89) (Fig. 1, Supporting information).

There were significant differences in the role of stochasticity among taxa (ANOVA, NST: $F_{70.6} = 19.7$, p < 0.0001; MST: $F_{70.6} = 33.6$, p < 0.0001; NTP: $F_{58.5} = 2.6$, p = 0.04). On average, ants had the highest NST (0.67), whereas scarabs had the lowest NST (0.16). Scarabs also had the lowest MST (0.02), while moths had the highest MST (0.65). Rove beetles had the highest NTP (0.77) and Orthoptera had the lowest NTP (0.59) (Supporting information). Trophic generalists had higher NTP than trophic specialists (beta regression, coefficient = -0.08, z = -2.24, p = 0.03), however there was no significant difference in NST or MST between trophic generalists and trophic specialists (NST: coefficient = -0.08, z=-1.42, p=0.15. MST: coefficient=-0.10, z=-2.34, p = 0.02) (Supporting information).

The relative importance of stochasticity was calculated within each sampling block then compared among blocks. Each block had a different level of logging intensity. When all taxa were combined, there was no significant effect of logging on stochasticity (Fig. 1. NST: slope = 0.06, z = 0.22, p = 0.83. MST: slope = 0.27, z = 1.12, p = 0.26. NTP: slope = -0.03,

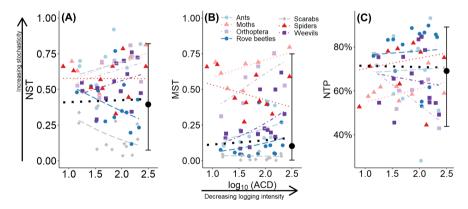


Figure 1. Variation in three different metrics of ecological stochasticity across a gradient of logging intensity for seven invertebrate taxa. Logging is quantified as log₁₀-transformed above-ground carbon density (ACD), where a higher ACD corresponds to a lower logging intensity. The thick black dotted regression lines indicate that there was no significant effect of logging on stochasticity when all taxa were combined. The black circles show the weighted average stochasticity metrics for all taxa combined, and the error bars represent 95% quantiles of these averages. The three stochasticity metrics are: (A) normalised stochasticity ratio (Ning et al. 2019); (B) modified stochasticity ratio (Ning et al. 2019); and (C) proportion of taxa with observed occurrence frequencies predicted by Sloan's neutral model (Sloan et al. 2006, 2007, Burns et al. 2016).

z=-0.14, p=0.89). The relationship between logging and stochasticity varied among taxa (Fig. 1). But, in general, the slopes for each taxon were not significant, with the only significant relationships being a decrease in NST with decreasing logging intensity for scarabs (slope=-1.03, p=0.046), and an increase in MST with decreasing logging intensity for Orthoptera (Orthoptera=1.04, p=0.027). Sloan's neutral model could not be fitted to the scarabs dataset, so scarabs were excluded from the NTP analysis.

Community assembly processes

Dispersal limitation was the dominant community assembly process when all taxa were combined and weighted by sample size (overall relative importance = 73, 95% quantiles = 4, 96). Dispersal limitation was the dominant process for four out of the seven taxonomic groups (rove beetles, scarabs, weevils and ants), with a relative importance ranging from 94 to 97% (quantiles = 87, 99). Drift was the dominant process for spiders, Orthoptera and moths; 89% of spider community assembly (quantiles = 78, 96) was estimated to be underpinned by drift, while this was estimated to be 75% for Orthoptera (quantiles = 55, 86) and 67% for moths (quantiles = 54, 78) (Fig. 2A, Supporting information).

We analysed the relative importance of the two dominant community assembly processes among sampling blocks. Each block had a different level of logging intensity. There was a weak decrease in the relative importance of dispersal limitation with decreasing logging intensity when all taxa were combined (slope=-0.66, z=-2.34, p=0.02) (Fig. 2B). The relative importance of drift was not significantly altered by logging when all taxa were combined (slope=0.05, z=0.21, p=0.83) (Fig. 2C). The only significant relationships for individual taxonomic groups were a decrease in the relative importance of dispersal limitation with decreasing logging intensity for moths, Orthoptera and rove beetles (moths: slope=-1.10, p=0.002, Orthoptera: slope=-2.01, p=0.002, rove beetles: slope=-1.24, p=0.01) (Fig. 2B).

Discussion

While the relative importance of stochasticity varied among taxa and metrics, in general, the balance between stochasticity and determinism appeared robust to a gradient of land use intensity. At a finer resolution, dispersal limitation, inferred from community patterns, was the dominant assembly process overall. Two out of the three stochasticity metrics (NST

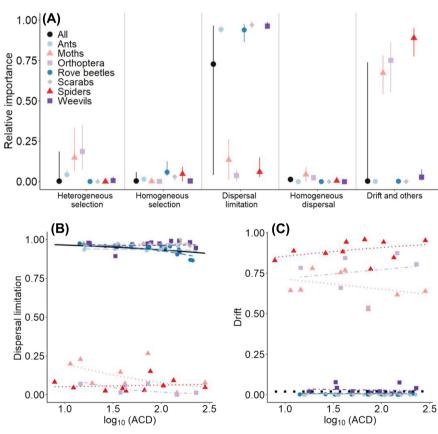


Figure 2. (A) Mean relative importance of five community assembly processes for generating the community structure of seven invertebrate taxa (coloured points), and weighted averages for all taxa combined (black circles). Error bars show bootstrapped 95% quantiles. (B–C) The effect of above-ground carbon density (ACD) on the two dominant community assembly processes: (B) dispersal limitation (C) drift for each taxon (coloured points and lines), and the overall estimates for all taxonomic groups combined (black lines: solid lines indicate significant relationships and dotted lines indicate non-significant relationships). Higher ACD corresponds to a lower logging intensity.

and MST) were not significantly different between trophic generalists and trophic specialists. Only the NTP metric supported our hypothesis that trophic generalists would be more stochastic than trophic specialists. In general, land use change had little impact on the relative importance of a suite of community assembly processes. Together, this suggests that, while logging has profoundly negative impacts on biodiversity, it tends to have little impact on the main community assembly drivers for most invertebrate taxa studied.

Overall, there was at best a very weak effect of land use intensity on the role of ecological stochasticity in structuring arthropod communities. The direction of the relationship between logging and stochasticity varied among taxa, but, in general, these relationships were not statistically significant (Fig. 1). We might expect to find little evidence of a change in the assembly processes governing invertebrate communities if those communities do not exhibit turnover in species composition across the logging gradient. For three of the taxa (ants, Orthoptera and spiders), this assumption holds true: we found no evidence of changing taxonomic identities across the logging gradient (Supporting information) which aligns well with a lack of change in the assembly processes governing those taxa (Fig. 2B). However, all three beetle taxa (rove beetles, scarabs and weevils) and moths did exhibit significant turnover in taxonomic identity as logging intensity increased (Supporting information), which is consistent with previous studies (Summerville and Crist 2002, Hamer et al. 2003, Cleary et al. 2007, Sharp et al. 2019). Yet these taxa generally did not exhibit significant changes in community assembly metrics across the logging gradient, suggesting that the species turnover was generated by the same ecological processes, regardless of logging intensity. This leads to the general conclusion that, regardless of whether taxonomic identities change, community assembly processes remain robust to changes in ACD.

One possible explanation for why community assembly processes appear to be strongly conserved across the gradient of logging intensity is that our data were collected after logging had taken place. The sites that experienced logging were initially logged in the 1970s, then logged between one and three times again between 1990 and 2008 (Riutta et al. 2018). While our study landscape encompasses a very wide range of historic logging intensity, the time delay between the logging event itself and our description of the invertebrate communities means any transitory impacts of logging on community assembly processes would not have been detectable. Mahayani et al. (2020) showed that phylogenetic diversity and community structure of tree communities had recovered 10 years after a single logging cycle in Bornean tropical forest. Therefore, it could be possible that the ecological communities we sampled might have recovered their basic, pre-logging structures so that the community assembly mechanisms in logged forests now resemble those of unlogged forests.

When inferring assembly processes from community patterns using coarse taxonomic hierarchies, dispersal limitation was the most important driver of community assembly for ants and beetles, whereas drift was the main driver of assembly for spiders, Orthoptera and moths (Fig. 2A). Spiders, Orthoptera and moths were sampled at fewer sites than ants and beetles (Supporting information), so we cannot definitively rule out the possibility that this result may represent a sampling effect. We do note, however, that the scarab community had a lower total number of individuals and a higher number of species than the Orthoptera, suggesting that an undersampling-driven effect should have exerted a greater impact on them than on the Orthoptera. Further, when we conducted a sensitivity analysis by grouping sites within the same block, drift was still the dominant community assembly process for spiders, moths and Orthoptera (Supporting information).

The overall importance of dispersal limitation as the dominant community assembly process, especially for ants and beetles highlights the importance of maintaining and, if necessary, restoring landscape connectivity in logged forests (Wearn et al. 2019). We emphasise that dispersal limitation was not directly measured in this study but was inferred from community patterns. Dispersal limitation can result in high spatial turnover in community composition due to low levels of exchange of organisms among local communities (Stegen et al. 2013). The increase in dispersal limitation as logging intensity increased (Fig. 2B) is in a direction that is consistent with previous studies suggesting that animal communities in logged tropical forests may experience lower levels of dispersal compared to primary forests (Stratford and Robinson 2005, Laurance et al. 2009, Edwards et al. 2014).

This study has quantified community assembly mechanisms across a gradient of logging intensity for seven groups of invertebrate taxa in Bornean rainforests. The effect of logging on stochasticity, and on different community assembly processes, varied among the different taxa and different metrics of stochasticity, but painted a general picture in which the dominant community assembly mechanisms are not impacted by logging disturbance. Although logging did not alter the balance between stochastic and deterministic community assembly processes for most taxa, we emphasise that logging, and in particular severe logging, profoundly reduces species richness and changes community composition (Thorn et al. 2018). The robustness of invertebrate communities to logging disturbance in our study suggests that knowledge of primary community assembly can be useful in planning the restoration of modified communities as there were generally not significant changes in assembly processes despite changes in land use intensity.

Acknowledgements – Tom Swinfield, David Milodowski, Tommaso Jucker, Michele Dalponte and David Coomes generated the aboveground carbon density layer from LiDAR data.

Funding – TMF was supported by a Czech Science Foundation Standard Grant (19-14620S). Field data collection was funded by Australian Research Council (ARC) Discovery Project grants (DP140101541 and DP160102078).

Author contributions

Natasha R. Granville: Conceptualization (equal); Formal analysis (lead); Visualization (lead); Writing - original draft (equal); Writing - review and editing (lead). Maxwell V. L. Barclay: Data curation (equal); Investigation (equal); Writing - review and editing (equal). Michael J. W. Boyle: Data curation (equal); Investigation (equal); Writing review and editing (equal). Arthur Y. C. Chung: Data curation (equal); Investigation (equal); Writing - review and editing (equal). Tom M. Fayle: Data curation (equal); Investigation (equal); Writing - review and editing (equal). Huai En Hah: Data curation (equal); Investigation (equal); Writing - review and editing (equal). Jane L. Hardwick: Data curation (equal); Investigation (equal); Writing – review and editing (equal). Lois Kinneen: Data curation (equal); Investigation (equal); Writing - review and editing (equal). Roger L. Kitching: Data curation (equal); Investigation (equal); Writing - review and editing (equal). Sarah C. Maunsell: Data curation (equal); Investigation (equal); Writing - review and editing (equal). Miller Jeremy: Data curation (equal); Investigation (equal); Writing - review and editing (equal). Adam C. Sharp: Data curation (equal); Investigation (equal); Writing – review and editing (equal). Nigel E. Stork: Data curation (equal); Investigation (equal); Writing – review and editing (equal). Leona Wai: Data curation (equal); Investigation (equal); Writing - review and editing (equal); Kalsum M. Yusah: Data curation (equal); Investigation (equal); Writing - review and editing (equal). Robert M. Ewers: Conceptualization (equal); Data curation (equal); Investigation (equal); Supervision (lead); Writing – original draft (equal); Writing – review and editing (equal).

Data availability statement

The data associated with this article are openly available on Zenodo at:

https://doi.org/10.5281/zenodo.1323504 (beetles) (Sharp et al. 2018)

https://doi.org/10.5281/zenodo.7011354 (Orthoptera) (Hardwick et al. 2022)

https://doi.org/10.5281/zenodo.4247169 (moths) (Maunsell et al. 2020b)

https://doi.org/10.5281/zenodo.7252200 (spiders) (Maunsell et al. 2020a)

https://doi.org/10.5281/zenodo.3876227 (ants) (Fayle et al. 2019)

https://doi.org/10.5281/zenodo.4020697 (above-ground carbon density)(Swinfield et al. 2020)

Supporting information

The Supporting information associated with this article is available with the online version.

References

Adler, P. B., Hillerislambers, J. and Levine, J. M. 2007. A niche for neutrality. – Ecol. Lett. 10: 95–104.

- Aldersley, A. and Orme, D. 2023. safedata: interface to data from the SAFE project. R package ver. 1.1.3 https://imperialcollegelondon.github.io/safedata/.
- Andersen, T., Carstensen, J., Hernández-García, E. and Duarte, C. M. 2009. Ecological thresholds and regime shifts: approaches to identification. Trends Ecol. Evol. 24: 49–57.
- Barlow, H. S. and Woiwod, I. P. 1989. Moth diversity of a tropical forest in Peninsular Malaysia. J. Trop. Ecol. 5: 37–50.
- Beisner, B., Haydon, D. and Cuddington, K. 2003. Alternative stable states in ecology. Front. Ecol. Environ. 1: 376–382.
- Berry, N. J., Phillips, O. L., Ong, R. C. and Hamer, K. C. 2008. Impacts of selective logging on tree diversity across a rainforest landscape: the importance of spatial scale. Landscape Ecol. 23: 915–929.
- Blüthgen, N., E. Stork, N. and Fiedler, K. 2004. Bottom-up control and co-occurrence in complex communities: honeydew and nectar determine a rainforest ant mosaic. Oikos 106: 344–358.
- Borenstein, M., Hedges, L. V., Higgins, J. P. and Rothstein, H. R. 2010. A basic introduction to fixed-effect and random-effects models for meta-analysis. Res. Synth. Methods 1: 97–111.
- Brooks, M., Bolker, B., Kristensen, K., Maechler, M., Magnusson, A., McGillycuddy, M., Skaug, H., Nielsen, A., Berg, C., van Bentham, K., Sadat, N., Lüdecke, D., Lenth, R., O'Brien, J., Geyer, C. J., Jagan, M., Wiernik, B. and Stouffer, D. B. 2023. glmmTMB: generalized linear mixed models using template model builder https://github.com/glmmTMB/glmmTMB
- Burivalova, Z., Sekercioğlu, C. H. and Koh, L. P. 2014. Thresholds of logging intensity to maintain tropical forest biodiversity. Curr. Biol. 24: 1893–1898.
- Burns, A. R., Stephens, W. Z., Stagaman, K., Wong, S., Rawls, J. F., Guillemin, K. and Bohannan, B. J. 2016. Contribution of neutral processes to the assembly of gut microbial communities in the zebrafish over host development. ISME J. 10: 655–664.
- Caruso, T., Taormina, M. and Migliorini, M. 2012. Relative role of deterministic and stochastic determinants of soil animal community: a spatially explicit analysis of oribatid mites. J. Anim. Ecol. 81: 214–221.
- Chase, J. M. 2007. Drought mediates the importance of stochastic community assembly. – Proc. Natl Acad. Sci. USA 104: 17430–17434.
- Chase, J. M. and Myers, J. A. 2011. Disentangling the importance of ecological niches from stochastic processes across scales. Phil. Trans. R. Soc. B 366: 2351–2363.
- Chase, J. M., Kraft, N. J. B., Smith, K. G., Vellend, M. and Inouye, B. D. 2011. Using null models to disentangle variation in community dissimilarity from variation in α-diversity. – Ecosphere 2: 1–11.
- Cleary, D. F. R., Boyle, T. J., Setyawati, T., Anggraeni, C. D., Van Loon, E. E. and Menken, S. B. 2007. Bird species and traits associated with logged and unlogged forest in Borneo. Ecol. Appl. 17: 1184–1197.
- Connor, E. F. and Simberloff, D. 1979. The assembly of species communities: chance or competition? Ecology 60: 1132–1140.
- Cribari-Neto, F. and Zeileis, A. 2010. Beta regression in R. J. Stat. Softw. 34: 1–24.
- Didham, R. K., Lawton, J. H., Hammond, P. M. and Eggleton, P. 1998. Trophic structure stability and extinction dynamics of beetles (Coleoptera) in tropical forest fragments. – Phil. Trans. R. Soc. B 353: 437–451.
- Ding, Y., Zang, R., Letcher, S. G., Liu, S. and He, F. 2012. Disturbance regime changes the trait distribution, phylogenetic structure and community assembly of tropical rain forests. Oikos 121: 1263–1270.

- Döbert, T. F., Webber, B. L., Sugau, J. B., Dickinson, K. J. M. and Didham, R. K. 2017. Logging increases the functional and phylogenetic dispersion of understorey plant communities in tropical lowland rain forest. – J. Ecol. 105: 1235–1245.
- Edwards, D. P., Tobias, J. A., Sheil, D., Meijaard, E. and Laurance, W. F. 2014. Maintaining ecosystem function and services in logged tropical forests. Trends Ecol. Evol. 29: 511–520.
- Ellwood, M. D. F., Manica, A. and Foster, W. A. 2009. Stochastic and deterministic processes jointly structure tropical arthropod communities. Ecol. Lett. 12: 277–284.
- Ewers, R. M., Didham, R. K., Fahrig, L., Ferraz, G., Hector, A., Holt, R. D., Kapos, V., Reynolds, G., Sinun, W., Snaddon, J. L. and Turner, E. C. 2011. A large-scale forest fragmentation experiment: the stability of altered forest ecosystems project. – Phil. Trans. R. Soc. B 366: 3292–3302.
- Fayle, T. M. ., Yusah, K. M., Ewers, R. M. and Boyle, M. J. W. 2019. How does forest conversion and fragmentation affect ant communities and the ecosystem processes that they mediate? [Data set].—Zenodo, https://doi.org/10.5281/zenodo.3876227.
- Gao, G. F., Peng, D., Tripathi, B. M., Zhang, Y. and Chu, H. 2020. Distinct community assembly processes of abundant and rare soil bacteria in coastal wetlands along an inundation gradient. – mSystems 5: e01150-20.
- Gaveau, D. L. A., Sloan, S., Molidena, E., Yaen, H., Sheil, D., Abram, N. K., Ancrenaz, M., Nasi, R., Quinones, M., Wielaard, N. and Meijaard, E. 2014. Four decades of forest persistence, clearance and logging on Borneo. – PLoS One 9: e101654.
- Gravel, D., Canham, C. D., Beaudet, M. and Messier, C. 2006. Reconciling niche and neutrality: the continuum hypothesis. – Ecol. Lett. 9: 399–409.
- Grimaldi, D. and Engel, M. S. 2005. Evolution of the insects. Cambridge Univ. Press.
- Hallett, L. M., Aoyama, L., Barabás, G., Gilbert, B., Larios, L., Shackelford, N., Werner, C. M., Godoy, O., Ladouceur, E. R., Lucero, J. E., Weiss-Lehman, C. P., Chase, J. M., Chu, C., Harpole, W. S., Mayfield, M. M., Faist, A. M. and Shoemaker, L. G. 2023. Restoration ecology through the lens of coexistence theory. – Trends Ecol. Evol. 38: 1085–1096.
- Hamer, K. C., Hill, J. K., Benedick, S., Mustaffa, N., Sherratt, T. N., Maryati, M. and K., C. V. 2003. Ecology of butterflies in natural and selectively logged forests of northern Borneo: the importance of habitat heterogeneity. J. Appl. Ecol. 40: 150–162.
- Hardwick, J., Kinneen, L., Maunsell, S., Stork, N., Yusah, K. M. and Kitching, R. 2022. The effects of habitat modification on the distribution and feeding ecology of Orthoptera 2015. [Data set]. Zenodo, https://doi.org/10.5281/zenodo.7011354.
- Hilderbrand, R. H., Watts, A. C. and Randle, A. M. 2005. The myths of restoration ecology. Ecol. Soc. 10: 19.
- Hu, X., Shu, Q., Shang, Z., Guo, W. and Qi, L. 2022. Secondary succession in the tropical lowland rainforest reduced the stochasticity of soil bacterial communities through the stability of plant communities. Forests 13: 348.
- Hubbell, S. P. 2011. The unified neutral theory of biodiversity and biogeography. – Princeton Univ. Press.
- Jiao, C., Zhao, D., Zeng, J., Guo, L. and Yu, Z. 2020. Disentangling the seasonal co-occurrence patterns and ecological sto-chasticity of planktonic and benthic bacterial communities within multiple lakes. Sci. Total Environ. 740: 140010.
- Jucker, T. et al. 2018. Estimating aboveground carbon density and its uncertainty in Borneo's structurally complex tropical for-

- ests using airborne laser scanning. Biogeosciences 15: 3811–3830.
- Keppeler, F. W., Cruz, D. A., Dalponti, G. and Mormul, R. P. 2016. The role of deterministic factors and stochasticity on the trophic interactions between birds and fish in temporary floodplain ponds. – Hydrobiologia 773: 225–240.
- Kitching, R. L. 2013. Niches and neutrality: community ecology for entomologists. – Aust. J. Entomol. 52: 1–7.
- Laurance, W. F., Goosem, M. and Laurance, S. G. 2009. Impacts of roads and linear clearings on tropical forests. Trends Ecol. Evol. 24: 659–669.
- Laurance, W. F. 2015. Emerging threats to tropical forests. Ann. Miss. Bot. Gard. 100: 159–169.
- Le Moigne, A., Bartosiewicz, M., Schaepman-Strub, G., Abiven, S. and Pernthaler, J. 2020. The biogeochemical variability of Arctic thermokarst ponds is reflected by stochastic and nichedriven microbial community assembly processes. Environ. Microbiol. 22: 4847–4862.
- Lepori, F. and Malmqvist, B. 2009. Deterministic control on community assembly peaks at intermediate levels of disturbance.
 Oikos 118: 471–479.
- Li, B., Wang, Y., Tan, W., Saintilan, N., Lei, G. and Wen, L. 2021.
 Land cover alteration shifts ecological assembly processes in floodplain lakes: consequences for fish community dynamics.
 Sci. Total Environ. 782: 146724.
- Liao, J., Cao, X., Zhao, L., Wang, J., Gao, Z., Wang, M. C. and Huang, Y. 2016. The importance of neutral and niche processes for bacterial community assembly differs between habitat generalists and specialists. – FEMS Microbiol. Ecol. 92: fiw174.
- Mahayani, N. P. D., Slik, F. J. W., Savini, T., Webb, E. L. and Gale, G. A. 2020. Rapid recovery of phylogenetic diversity, community structure and composition of Bornean tropical forest a decade after logging and post-logging silvicultural interventions. – For. Ecol. Manage. 476: 118467.
- Maunsell, S., Miller, J., Fayle, T., Yusah, K. M., Stork, N. and Kitching, R. 2020a. Variation in arthropod responses to tropical landscape transformation: spiders 2015. [Data set]. Zenodo, https://doi.org/10.5281/zenodo.7252200.
- Maunsell, S., Fayle, T., Yusah, K. M., Wai, L, Hah, H. E., and, Stork, N. and Kitching, R. 2020b. Variation in arthropod responses to tropical landscape transformation: moths 2014. [Data set].—Zenodo, https://doi.org/10.5281/zenodo.4247169.
- Méndez-Rojas, D. M., Cultid-Medina, C. and Escobar, F. 2021. Influence of land use change on rove beetle diversity: a systematic review and global meta-analysis of a mega-diverse insect group. Ecol. Indic. 122: 107239.
- Newbold, T. et al. 2015. Global effects of land use on local terrestrial biodiversity. Nature 520: 45–50.
- Ning, D., Deng, Y., Tiedje, J. M. and Zhou, J. 2019. A general framework for quantitatively assessing ecological stochasticity.
 Proc. Natl Acad. Sci. USA 116: 16892–16898.
- Ning, D., Yuan, M., Wu, L., Zhang, Y., Guo, X., Zhou, X., Yang, Y., Arkin, A. P., Firestone, M. K. and Zhou, J. 2020. A quantitative framework reveals ecological drivers of grassland microbial community assembly in response to warming. Nat. Commun. 11: 4717.
- Nyffeler, M. and Birkhofer, K. 2017. An estimated 400–800 million tons of prey are annually killed by the global spider community. Naturwissenschaften 104: 30.
- Oksanen, J. et al. 2022. vegan: community ecology package. R package, ver. 2.6.4,https://github.com/vegandevs/vegan.

- Ortega-Martínez, I. J., Moreno, C. E., Rios-Díaz, C. L., Arellano, L., Rosas, F. and Castellanos, I. 2020. Assembly mechanisms of dung beetles in temperate forests and grazing pastures. Sci. Rep. 10: 391.
- Oumarou Ngoute, C., Kekeunou, S., Lecoq, M., Nzoko Fiemapong, A. R., Um Nyobe, P. C. A. and Bilong Bilong, C. F. 2020. Effect of anthropogenic pressure on grasshopper (Orthoptera: Acridomorpha) species diversity in three forests in southern Cameroon. J. Orthopt. Res. 29: 25–34.
- Palmer, M. A., Ambrose, R. F. and Poff, N. L. 1997. Ecological theory and community restoration ecology. – Restor. Ecol. 5: 291–300.
- Powell, J. R., Karunaratne, S., Campbell, C. D., Yao, H., Robinson, L. and Singh, B. K. 2015. Deterministic processes vary during community assembly for ecologically dissimilar taxa. – Nat. Commun. 6: 8444.
- Riutta, T., Malhi, Y., Kho, L. K., Marthews, T. R., Huaraca Huasco, W., Khoo, M., Tan, S., Turner, E., Reynolds, G., Both, S., Burslem, D. F. R. P., Teh, Y. A., Vairappan, C. S., Majalap, N. and Ewers, R. M. 2018. Logging disturbance shifts net primary productivity and its allocation in Bornean tropical forests. Global Change Biol. 24: 2913–2928.
- Romeis, J., Städler, E. and Wäckers, F. 2005. Nectar- and pollenfeeding by adult herbivorous insects. – In: Wäckers, F. L., van Rijn, P. C. J. and Bruin, J. (eds), Plant-provided food for carnivorous insects: a protective mutualism and its applications. Cambridge Univ. Press, pp. 178–220.
- Russell-Smith, A. and Stork, N. E. 1995. Composition of spider communities in the canopies of rainforest trees in Borneo. J. Trop. Ecol. 11: 223–235.
- Sharp, A., Barclay, M., Chung, A., Ewers, R. 2018. Beetle diversity. [Data set]. Zenodo, https://doi.org/10.5281/zenodo.1323504.
- Sharp, A. C., Barclay, M. V. L., Chung, A. Y. C. and Ewers, R. M. 2019. Tropical logging and deforestation impacts multiple scales of weevil beta-diversity. – Biol. Conserv. 234: 172–179.
- Shipley, B., Paine, C. E. and Baraloto, C. 2012. Quantifying the importance of local niche-based and stochastic processes to tropical tree community assembly. Ecology 93: 760–769.
- Shoemaker, L. G. et al. 2020. Integrating the underlying structure of stochasticity into community ecology. Ecology 101: e02922.
- Sloan, W. T., Lunn, M., Woodcock, S., Head, I. M., Nee, S. and Curtis, T. P. 2006. Quantifying the roles of immigration and chance in shaping prokaryote community structure. – Environ. Microbiol. 8: 732–740.
- Sloan, W. T., Woodcock, S., Lunn, M., Head, I. M. and Curtis, T. P. 2007. Modeling taxa-abundance distributions in microbial communities using environmental sequence data. Microb. Ecol. 53: 443–455.
- Stegen, J. C., Lin, X., Fredrickson, J. K., Chen, X., Kennedy, D. W., Murray, C. J., Rockhold, M. L. and Konopka, A. 2013. Quantifying community assembly processes and identifying features that impose them. ISME J. 7: 2069–2079.
- Stratford, J. A. and Robinson, W. D. 2005. Gulliver travels to the fragmented tropics: geographic variation in mechanisms of avian extinction. Front. Ecol. Environ. 3: 85–92.

- Suding, K. N. and Hobbs, R. J. 2009. Threshold models in restoration and conservation: a developing framework. Trends Ecol. Evol. 24: 271–279.
- Summerville, K. S. and Crist, T. O. 2002. Effects of timber harvest on forest Lepidoptera: community, guild, and species responses. Ecol. Appl. 12: 820–835.
- Sun, Y., Zhang, M., Duan, C., Cao, N., Jia, W., Zhao, Z., Ding, C., Huang, Y. and Wang, J. 2021. Contribution of stochastic processes to the microbial community assembly on field-collected microplastics. – Environ. Microbiol. 23: 6707–6720.
- Swinfield, T., Milodowski, D., Jucker, T., Michele, D. and Coomes, D. 2020. LiDAR canopy structure 2014 [dataset]. – Zenodo, https://doi.org/10.5281/zenodo.4020697.
- Thompson, R. and Townsend, C. 2006. A truce with neutral theory: local deterministic factors, species traits and dispersal limitation together determine patterns of diversity in stream invertebrates. J. Anim. Ecol. 75: 476–484.
- Thorn, S. et al. 2018. Impacts of salvage logging on biodiversity: a meta-analysis. J. Appl. Ecol. 55: 279–289.
- Trego, A. C., McAteer, P. G., Nzeteu, C., Mahony, T., Abram, F., Ijaz, U. Z. and O'Flaherty, V. 2021. Combined stochastic and deterministic processes drive community assembly of anaerobic microbiomes during granule flotation. – Front. Microbiol. 12: 666584.
- Valdivia, N., Garcés-Vargas, J., Garrido, I., Gómez, I., Huovinen, P., Navarro, N. P., Macaya, E. C. and Pardo, L. M. 2021. Beta diversity of Antarctic and sub-Antarctic benthic communities reveals a major role of stochastic assembly processes. – Front. Mar. Sci. 8: 780268.
- Vellend, M. 2010. Conceptual synthesis in community ecology. Q. Rev. Biol. 85: 183–206.
- Wang, D., Huang, K., He, X., Zhang, X. X. and Meng, Y. 2022. Varied interspecies interactions between anammox and denitrifying bacteria enhanced nitrogen removal in a single-stage simultaneous anammox and denitrification system. – Sci. Total Environ. 813: 152519.
- Wearn, O. R., Carbone, C., Rowcliffe, J. M., Pfeifer, M., Bernard,
 H. and Ewers, R. M. 2019. Land-use change alters the mechanisms assembling rainforest mammal communities in Borneo.
 J. Anim. Ecol. 88: 125–137.
- Weiher, E., Freund, D., Bunton, T., Stefanski, A., Lee, T. and Bentivenga, S. 2011. Advances, challenges and a developing synthesis of ecological community assembly theory. Phil. Trans. R. Soc.B 366: 2403–2413.
- Xu, Q., Vandenkoornhuyse, P., Li, L., Guo, J., Zhu, C., Guo, S., Ling, N. and Shen, Q. 2022. Microbial generalists and specialists differently contribute to the community diversity in farmland soils. – J. Adv. Res. 40: 17.
- Yang, Y., Cheng, K., Li, K., Jin, Y. and He, X. 2022. Deciphering the diversity patterns and community assembly of rare and abundant bacterial communities in a wetland system. Sci. Total Environ. 838: 156334.
- Zhou, Z., Zheng, M., Xia, J. and Wang, C. 2022. Nitrogen addition promotes soil microbial beta diversity and the stochastic assembly. Sci. Total Environ. 806: 150569.