

Novel indices reveal that pollinator exposure to pesticides varies across biological compartments and crop surroundings

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1 **Novel indices reveal that pollinator exposure to pesticides varies**
2 **across biological compartments and crop surroundings**

3

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40

41 **Highlights**

42

- 43 • We use new indices to summarise big datasets on pesticide exposure of three species
44 of bees
- 45 • Novel indices are calculated using Item Response Theory (IRT) models
- 46 • The indices are linked to the number of pesticides rather than the active ingredients
- 47 • Matrices collected from apple orchards are exposed to a higher number of pesticides
48 than matrices collected from oilseed rape crops
- 49 • Pollen related matrices contained more pesticides than were found in nectar and on
50 the bees themselves

51

52

53 **Abstract.**

54 Declines in insect pollinators have been linked to a range of causative factors such as disease,
55 loss of habitats, the quality and availability of food, and exposure to pesticides. Here, we
56 analysed an extensive dataset generated from pesticide screening of foraging insects, pollen-
57 nectar stores/beebread, pollen and ingested nectar across three species of bees collected at
58 128 European sites set in two types of crop. In this paper, we aimed to (i) derive a new index
59 to summarise key aspects of complex pesticide exposure data and (ii) understand the links
60 between pesticide exposures depicted by the different matrices, bee species and apple
61 orchards *versus* oilseed rape crops. We found that summary indices were highly correlated
62 with the number of pesticides detected in the related matrix but not with which pesticides
63 were present. Matrices collected from apple orchards generally contained a higher number of
64 pesticides (7.6 pesticides per site) than matrices from sites collected from oilseed rape crops
65 (3.5 pesticides), with fungicides being highly represented in apple crops. A greater number of

66 pesticides were found in pollen-nectar stores/bee bread and pollen matrices compared with
67 nectar and bee body matrices. Our results show that for a complete assessment of pollinator
68 pesticide exposure, it is necessary to consider several different exposure routes and multiple
69 species of bees across different agricultural systems.

70

71 **Keywords**

72 Item Response Theory. Bumblebee. Osmia. Apple orchards. Oilseed rape.

73

74 **1. Introduction**

75 Declines in species of both managed and wild pollinators has been repeatedly documented
76 [1] in Europe [2], the US [3], Canada [4], Asia [5] and to some extent in South-America [6] and
77 Africa [7]. Managed bees such as honeybees (*Apis mellifera*) [8] and wild bees [9, 10] are the
78 most important group of pollinators in Europe and other regions of the world (IPBES 2016). A
79 range of factors have been suggested to explain losses of bees such as diseases [11, 12], loss
80 of habitats [13, 14], the quality and availability of food [15, 16] and exposure to pesticides [17,
81 18]. The way bees are exposed to pesticides is variable and depends mainly on the type of
82 pesticide [19, 20], their purpose of use (which is related to the application mode i.e. spray, soil
83 treatment, trunk injection), [21] and on the ecology of species [22, 23]. Application timing
84 (pre-bloom *versus* at-bloom) has logically dramatic impacts on exposure levels for pollinators
85 feeding on nectar and pollen from flowers [18]. Several techniques have been developed to
86 limit this exposure such as microencapsulated compounds and seed coated insecticides with
87 systemic properties [24]. Bees can also be exposed to pesticides through water consumption
88 [25, 26], pesticide contact [27], air [19, 28, 29] and, in the case of managed bees, the use of
89 veterinary products [30, 31]. However, dietary consumption is the major route of exposure
90 [18].

91 Honeybees produce large quantities of honey from collected nectar. In addition, for storage
92 purposes, after collection, pollen grains are processed into beebread. This term usually refers
93 to honeybee pollen stores, as beebread is pollen with added nectar and enzymes [32] and
94 stored in frames made of beeswax. For other bee species, however, any substance consisting
95 predominantly of stored pollen will be referred to as pollen-nectar stores in this paper.

96 Previously, pesticide residues have been documented in nectar [18], honey [33], pollen
97 collected on flowers [19], honeybee pollen pellets collected with traps [34], honeybee
98 beebread [35], wax [36] and honeybees themselves [37]. However, the majority of exposure
99 studies describe the contamination of one or two matrices at the same time [38]. To our
100 knowledge, our study is the first to present results across pesticides in pollen collected from
101 flowers and from pollen pellets, in pollen-nectar stores and beebread, in nectar regurgitated
102 from honeybees and from other bee species and from bee bodies, collected at the same time
103 in the same site. In an attempt to better understand the exposure route of three bee species
104 (*Apis mellifera*, *Bombus terrestris* and *Osmia bicornis*), we assessed pesticides in each of these
105 matrices at the same time in 128 sites set in two types of crops (apple orchards, oilseed rape)
106 across Europe. To our knowledge, this dataset is one of the most extensive datasets of bee
107 exposure to pesticides currently available.

108 As the number of pesticides measured in the different matrices and for each site was very
109 large, it was necessary to synthesise this complex information. The construction of such
110 indices, that are able to summarise information for all pesticides detected at a site, is of
111 paramount interest. Such an index can be used, for instance, for investigating the links
112 between the different matrices under study or in structuring model equations to explore the
113 role of stresses on bee population dynamics. A classic way to summarise pesticide information
114 is to calculate the richness (i.e., the number of pesticides detected in a given sample), or the
115 abundance (i.e., the total quantity of pesticides detected in a given sample) [39]. However,
116 these simple calculations do not capture information on pesticide variability across the
117 samples. In this paper, we propose to apply an original method, namely Item Response Theory
118 (IRT) models to calculate an index that includes as much variability as possible while being
119 easily interpretable.

120 The IRT models build such indices, each being associated with a matrix (i.e., pollen-nectar
121 stores or beebread, pollen, nectar and foragers from different species and flowers) and a crop
122 (i.e., apple orchards, oilseed rape). We also propose a method to interpret these indices
123 (section 3.1). In a second step, the links between all these indices are studied (section 3.2).
124 Results are discussed in the context of the existing literature (section 4).

125

126 **2. Materials and methods**

127 **2.1. Samples collection in PoshBee site network**

128 Within the H2020 project 'PoshBee' (www.PoshBee.EU), a site network for assessing exposure
129 of bees to chemical, nutritional, and pathogen stressors was established in 2019 [40]. Data
130 were collected at 128 sites across eight participating countries (Estonia, Germany, Ireland,
131 Italy, Spain, Sweden, Switzerland and the United Kingdom) situated in either apple orchards
132 or oilseed rape crops. At each site, three honeybee colonies, three trap nests seeded with
133 male and female cocoons of *Osmia bicornis* (solitary bee) and three *Bombus terrestris*
134 (bumblebee) colonies were installed following the PoshBee protocols [40].

135 At each site, various matrices were collected from all colonies and nests in equal proportions,
136 pooled per species and subsequently sent for pesticide residues analyses in different
137 laboratories [41]. If field constraints prevented the collection of equal proportions, acceptable
138 differences between colony/nest were limited to a maximum of 30%. If one colony/nest did
139 not produce the quantity required, the quantities from the remaining two were increased in
140 order to reach the total quantity required. The sampling of each matrix was performed only
141 once for each species at each site generally on the same day. Depending on the matrix,
142 sampling was performed either during or towards the end of the flowering period to be

143 consistent with biological cycles of bees (Figure A1 and Figure A2, in supplementary material).
144 At each site, *A. mellifera* and *B. terrestris* adults were collected alive. Bees were gently pressed
145 at the two first abdominal segments on the crop (honey sack) until a drop of nectar was
146 regurgitated between the bee mandibles. Nectar was collected and pooled for each species
147 to produce one sample per species for each site for pesticide analysis.

148 The matrices listed in Figure A1 were sampled and subsequently analysed for determination
149 and quantification of pesticide residues. Due to the behavior and limited success of solitary
150 bees in the wild, it was not possible to obtain sufficient numbers of *O. bicornis* bees or
151 amounts of regurgitated nectar to perform analyses for pesticide residues on these matrices
152 (Table 1).

153

154 **2.2. Analytical methods for pesticide determination and quantification**

155 Four different laboratories analysed the samples to identify and quantify pesticide residues.
156 Each laboratory was in charge of a specific matrix and had specific developed and validated
157 methods with liquid chromatography tandem mass spectrometry (LC-MS/MS) and gas
158 chromatography tandem mass spectrometry (GC-MS/MS). The different analytical methods
159 were detailed for pollen-nectar stores and beebread [31], nectar (Martel et al, submitted),
160 bees [42] and pollen from flowers and from traps.

161 Sample preparation for residue analysis of 261 pesticides and their metabolites as well as 6
162 congeners of non-dioxin like polychlorinated biphenyls (ndl-PCB) in a very low mass beebread
163 or pollen-stores samples was based on modified QuEChERS protocol with all steps
164 miniaturized to enable multiresidue analysis. Sample of beebread (0.3 g) was extracted with
165 1 mL of acetonitrile containing 5% formic acid and ammonium formate salt was added for

166 partitioning. Then the supernatant was subjected to clean-up by freezing and two-step
167 dispersive solid phase extraction (dSPE) into a Supel™ QuE Verde mini tube with sorbents
168 (Supelclean™ ENVI-Carb™ Y, 10 mg; Supelclean™ PSA, 50 mg; Z-Sep+, 60 mg; magnesium
169 sulfate, 150 mg). After 1st step dSPE, a portion of extract was analysed by LC-MS/MS for 200
170 pesticide residues. Remaining extract was subjected to a 2nd step dSPE clean-up by another
171 Supel™ QuE Verde mini tube and then after evaporation to dryness and dissolved in hexane,
172 it was analysed by GC-MS/ MS for another 61 pesticide and 6 ndl-PCB residues. Method
173 enabled determination of residues of 101 insecticides, 72 herbicides, 67 fungicides, 10
174 acaricides, 6 growth regulators, 5 veterinary drugs and 6 ndl-PCB's. The limits of quantification
175 (LOQ) values have been established as follows: 0.001 mg/kg for 105, 0.005 mg/kg for 96, 0.01
176 mg/kg for 31, 0.05 mg/kg for 31 and 0.1 mg/kg for 4 compounds respectively.

177 For the determination of 85 pesticides in nectar regurgitated by honey bees and bumble bees
178 two multiresidue methods were applied. One method involved an extraction of 40 pesticides
179 in 10 µL of nectar sample and an analysis with LC-MS/MS. The other one for the determination
180 of 45 pesticides in 100 µL of nectar sample involved a liquid/liquid partitioning with an organic
181 solvent and an evaporation to dryness. Then the extract was recovered with appropriate
182 solvent for GC-MS/MS analysis. The LOQ were ranged between 5 and 10 pg/µL for LC-MS/MS
183 and between 10 and 521 pg/µL for GC-MS/MS.

184 In the bee bodies (honey bees and bumble bees) the total number of molecules that was
185 screened for, inclusive of isomers and metabolites, was 373. A simplified QuEChERS method
186 was used for sample preparation, which consisted of an extraction with water, acetonitrile
187 and salts (MgSO₄ and NaCl). After centrifugation, the supernatant was cleaned-up on PSA

188 (dSPE). The sample was again centrifuged, concentrated and a specific solvent was added for
189 the GC-MS/MS or LC-MS/MS analysis. The LOQ were between 0.002 and 0.05 mg/kg.

190 Sample preparation of pollen to quantify more than 336 compounds was based on modified
191 QuEChERS protocol. Water was added to the prepared homogeneous samples (1 g) before
192 extraction with acetonitrile. Samples below 300 mg could not be analysed. Magnesium
193 sulfate, sodium chloride and sodium citrate salts were added to the sample for liquid/liquid
194 partitioning. A portion of the organic phase was subjected to a step of freezing following by a
195 clean-up on a mixture of MgSO₄, PSA and C18 (dSPE). Then, the extract with 5% of formic acid
196 in acetonitrile was directly analysed by LC-MS/MS. The GC-MS analysis required the change of
197 solvent to hexane/acetone 4:1 (v/v). Limits of quantification were 0.005 to 0.01 mg/kg.

198 This resulted in five different lists of pesticides depending on matrices. However, 64 common
199 pesticides were selected at the beginning of PoshBee based on agrochemicals applied on crops
200 at the European level to enable comparison between matrices. The index calculation was not
201 restricted to these 64 pesticides. Indeed, if a pesticide was detected in only one matrix, it
202 contributed to increase the exposure in the site where it was detected. As a consequence, the
203 indices' values increased. At the end, 267 pesticides were screened for in pollen-nectar stores
204 and beebread, 373 pesticides in foragers, 85 pesticides in nectar, 336 pesticides in pollen from
205 *A. mellifera* traps and 300 pesticides in pollen from flowers.

206 A minimum quantity was required to perform laboratory analysis. This requirement was not
207 always met due to field constraints. Thus, results were missing for some sites or matrices. At
208 the end, 319 pollen-nectar store/beebread samples, 253 forager samples, 251 nectar samples,
209 117 *A. mellifera* pollen-trap samples and 60 flower pollen samples were analysed (Table 1).

210 Table 1 – Overview of the number of sites sampled and analysed, the number of pesticides screened and detected
211 in each matrix for each species and crop corresponding to the 18 datasets included in the indices calculation. The
212 percentages of sites with analysed samples were compared to the theoretical number of samples according to
213 the protocol (=64 samples for each matrix, i.e., 8 sites × 8 countries). *A. m*: *Apis mellifera*. *B. t*: *Bombus terrestris*.
214 *O. b*: *Osmia bicornis*. APP: apple. OSR: oilseed rape. *Apis*: pollen collected with pollen traps set up on *A. mellifera*
215 colonies.

216

217 The quality and consistency of all the analytical results was automatically controlled in a
218 database designed for this purpose (named Poshbase) enabling the collection of 18 datasets
219 corresponding to the matrices across the three bee species (Table 1).

220 The theoretical number of sites under study was 64 for a given matrix and crop (Table 1).
221 However for various reasons (i.e. quantity of sampled matrix not sufficient for subsequent
222 laboratory analysis, difficulty to retrieve matrix from the field due to weather conditions or
223 scarce quantity), the actual number of sites in the statistical analysis was reduced. The largest
224 reduction was observed for the pollen collected directly on flowers in apple orchards (N=26)
225 and oilseed rape (N=34). The number of sites with at least one pesticide detected in a matrix
226 varied from 100% in beebread from honeybee colonies in apple orchards or oilseed rape and
227 in pollen-nectar stores from solitary bees' nests in oilseed rape crops for instance, to 33% in
228 bumblebee foragers in oilseed rape crops. Between 11 (in bumblebees in oilseed rape crops)
229 and 98 (in honeybee beebread collected in colonies in apple orchards) pesticides were
230 detected in any given matrix, representing between 3% and 37% of the pesticides screened
231 for.

232 As the calculation of the indices was intended to give the best discrimination between sites,
233 only pesticides detected in at least one site were taken into account. Thus, each dataset used

234 for the statistical analysis was of dimension $N \times P$ (Table 1; e.g. for *Beebread.Apis* and for apple
235 orchards, $P=98$ pesticides were detected and measured in $N=62$ sites) and included the
236 quantification of each pesticide in each site. More precisely for a given site, a given pesticide
237 and a given matrix, the following rules were applied: the LOQ (limit of quantification, the
238 pesticides detected below this value cannot be quantified) was used for values between the
239 LOD (limit of detection; below this value, the pesticides cannot be detected with sufficient
240 confidence) and the LOQ, and quantified values were kept in cases of values higher than LOQ.
241 As the data had many zeros (i.e., non-detected pesticides), the calculation of the indices was
242 based on binary data: 0 was used if the value was inferior to LOD and 1 was used otherwise.
243 However, the index's interpretation was based on raw quantified values.

244

245 **2.3. Statistical analyses**

246 Our aim was to summarise and interpret the large amount of information available in each
247 dataset. For this purpose and in a first step, 18 indices were built, one for each matrix and
248 each crop. The objective was to reduce the dimensionality of the datasets to characterise the
249 site exposure to pesticides in a unidimensional and interpretable index. Subsequently, each
250 index was interpreted according to the pesticides detected. Finally, and for each crop, the links
251 between the nine indices were studied with a Principal Component Analysis as a summary of
252 correlation matrix (Figure 1).

253 Figure 1 - The overall statistical procedure for a given crop (apple orchard or oilseed rape) for the nine matrices
254 across the three bee species (*Pollen.Flower*, *Nectar.Apis*, *Apis*, *Pollen.Apis*, *Beebread.Apis*, *Nectar.Bombus*,
255 *Bombus*, *Pollen-nectar stores.Bombus*, *Pollen-nectar stores.Osmia*). The map is from Hodge et al. 2022. IRT: Item
256 Response Theory. PCA: Principal Component Analysis.

257

258 **Calculation of indices.** Initially developed in the psychology framework, the Item Response
259 Theory (IRT) models aim at building a unidimensional scale (= latent trait = index), from
260 different items that measure this trait [43, 44]. The IRT concept was translated as to whether
261 a site exhibited a given pesticide or if the pesticide was absent from the site. The more
262 pesticide were recorded the i^{th} site was, the higher its index value, denoted θ_i .

263 For a given pesticide j , the two parameters to be estimated in the model were the mean
264 exposure level of a site (a_j) and the specific exposure level of a site (b_j), fitted with an EM
265 algorithm [45] (Chalmers, 2012). The exposure level (measured here as the number of
266 detected pesticides per site) was the level a site should have, to have 50% chance to exhibit a
267 pesticide. The specific exposure level represented how well the item (i.e. pesticide) separated
268 sites with high exposure scores from sites with low exposure scores. In theory, most, if not all
269 pesticides, should have a positive specific exposure level: the more exposed a site was, the
270 more likely it was to detect a given pesticide. For this purpose, the following two-parameter
271 logistic model was applied. Let $P(X_{i,j} | \theta_i)$ be the probability that the site i exhibited the pesticide
272 j given its exposure level, such as:

$$273 \quad P(X_{i,j} | \theta_i) = \frac{1}{1 + e^{-a_j(\theta_i - b_j)}} \quad \text{for the } j^{\text{th}} \text{ pesticide and the } i^{\text{th}} \text{ site } (i=1, \dots, 64)$$

274 With a_j the exposure level, b_j the site-discrimination and θ_i the level of exposure at site i .

275 For several pesticides under study, the previous model was adapted: all the pesticides were
276 included and then selected through a backward selection algorithm applied to filter out non-
277 interpretable pesticides. To maximize the statistical significance of the two parameters (a_j and
278 b_j), a double control on each step of the algorithms was implemented: (i) a stepwise loop
279 stopped if there were no more pesticides with a negative discrimination, or (ii) if the
280 performance criterion of the model (=Akaike information criterion, AIC) stopped decreasing.

281 At the end, only pesticides with a positive discrimination were retained. In addition, the
282 stability of the selection was tested with a leave-one-out cross validation, both on sites and
283 pesticides. In summary, using the index was relevant when the information on the pesticide
284 detection was fragmented between different pesticides (see the discussion for details).

285 **Interpretation of indices.** The index was calculated on pesticide presence or absence to have
286 robust calculations and deal with the many zeros. However, as the interpretation was not
287 based on more robust statistical tests, the quantities of pesticides from the raw quantified
288 data were used (Table 2). For a given matrix and a given crop, the pesticides, as well as
289 countries, that most contributed to the index were highlighted and interpreted. For this
290 purpose, all the available sites were clustered by means of a Hierarchical Clustering Analysis
291 applied to each index value [46]. Then, the pesticides that were significantly over-represented
292 in a cluster compared to the mean were highlighted [47]. Similarly, under-represented
293 pesticides compared to the mean could also be identified; they were detailed only in Table 2
294 for the example and interpretation. Two supplementary variables (i.e., number of pesticides
295 and country) were also taken into account. Sites of a given country that were over- or under-
296 represented in a cluster compared to the mean were also highlighted. Consequently, the
297 interpretation of presence/absence of sites from a given country compared to sites from other
298 countries was possible (see Table 2). It is worth noting that the number of sites per country
299 (N=8 sites) did not allow the extrapolation of results to the whole country. Indeed, the site
300 network was not designed to be representative of countries, but rather to be representative
301 of these crop landscapes in the European territory.

302 **Links between indices.** For a given crop (apple or oilseed rape), the links between the nine
303 indices – related to the different matrices – were studied with a Principal Component Analysis
304 (PCA) [48].

305 All the analyses were implemented in R software (version 4.1.3 <https://www.r-project.org/>).
306 The IRT models were estimated using the mirt R package with the ‘Rasch’ option. The
307 clustering was applied with the HCPC function of the FactoMineR package [49] and the
308 interpretation of the indices was made with the catdes (for categorical variable such as
309 country) or condes (for numeric variable such as the number of pesticides) functions of the
310 FactoMineR package. Principal Component Analyses were performed with the PCA function
311 of the FactoMineR package.

312

313 **3. Results**

314 **3.1 Indices: IRT results and interpretation**

315 **3.1.1 Detailed interpretation of indices related to beebread collected in *A. mellifera* colonies** 316 **in apple orchards**

317 As a proof of principle, we chose to interpret in detail the index of site characterisation for a
318 single dataset: the pesticide residues detected in beebread collected from *A. mellifera*
319 colonies in the 62 apple orchard sites (Table 2). The complete set of the indices’ values for
320 each site and the interpretation of the indices are given in Tables A.1 to A.4 (in supplementary
321 material).

322 According to their index values, the sites were separated into four clusters. The statistical
323 differences between clusters highlighted the unequal repartition of detected pesticides. In

324 other words, if a pesticide was detected (respectively not detected) in a limited number of
325 clusters, it was qualified as an over-represented (respectively under-represented) pesticide.
326 If a pesticide was present in all the clusters, it was not considered as over-represented.
327 Pesticides were less present in Cluster 1 (N=10 sites out of the 62) than the mean calculated
328 across all sites. It presented the lowest index value (-1.32). Only a few pesticides (mean of
329 3.90) were detected in samples and none were over-represented compared to the mean.
330 Estonian sites were the most frequent in this cluster. Cluster 2 (N=12) did not contain sites
331 over or under-represented compared to the mean. The index value was negative (-0.49) but
332 higher than cluster 1's, meaning than cluster 2's sites were exposed to fewer pesticides than
333 the mean calculated across all sites but exposed to a higher number of pesticides than the
334 sites in the cluster 1. Cluster 3 (index value of 0.16) contained most of the sites (N=21) though
335 no pesticide nor country was over- or under- represented. Cluster 4 (N=19, index value of 0.83)
336 included the sites exposed to a high number of pesticides with 30 pesticides over-represented
337 compared to the mean. One insecticide (flonicamid) and five herbicides were the most
338 significant pesticides ($p < 0.005$). The concentrations in this cluster ranged from 9 230 for the
339 dithianon to 78.2 $\mu\text{g}/\text{kg}$ for the flonicamid. The United Kingdom and German sites were over-
340 represented in this cluster and therefore hosted sites with higher number of detected
341 pesticides. Swiss, Irish and Swedish sites were significantly absent from Cluster 4. They were
342 present in Clusters 1, 2 and 3 but not over-represented in any of these clusters.

343

344 Table 2 – Field site characterisation based on the index calculated on pesticide residues detected in **beebread**
345 collected in *A. mellifera* colonies in the 62 **apple orchards** sites. CHE: Swiss sites. EST: Estonian sites. GER: German
346 sites. IRL: Irish sites. SWE: Swedish sites. UK: The United Kingdom sites.

347

348 3.1.2 Overall description of the indices

349 All 18 indices were highly positively correlated with the number of pesticides detected in the
350 matrices (mean correlation = 0.99; Table A.5, in supplementary material). This meant the
351 higher the value of an index, the more exposed to a high number of pesticides the site was
352 (details in Tables A.3 and A.4). Generally, matrices collected from apple orchards were
353 exposed to a higher number of pesticides than matrices collected from oilseed rape crops,
354 with respectively 7.6 [3.3-11.9] versus 3.5 [0.9-6.1] pesticides on average (details in Tables A.3
355 and A.4). Fungicides were highly present in the pesticides significant for the discrimination of
356 clusters: 70% and 43.4% in apple orchards sites and in oilseed rape crops, respectively (Table
357 A.6). Insecticides (20% and 33.9%, respectively) and herbicides (10% and 16.9%, respectively)
358 were the other pesticide families the most represented. The quantities of these pesticides
359 ranged from a minimum of 1.04 (insecticides) to a maximum of 9 230 µg/kg (fungicides) in
360 apple orchard sites; and from 0.47 (for insecticides and herbicides) to 2 880 µg/kg (fungicides)
361 in oilseed rape crop sites. Irrespective of the crop, pollen-nectar stores/beebread and pollen
362 matrices contained a higher number of pesticides than nectar and forager matrices (Tables
363 A.7 and A.8, in supplementary material). For apple orchards for instance, 15.1 and 10.4
364 pesticides were found respectively in beebread collected from *Apis* foragers and pollen from
365 flowers whereas only 2.2 and 1.3 were found in nectar regurgitated from *Apis* foragers and in
366 *Apis* foragers respectively. For oilseed rape, 14.9 and 7.7 pesticides were found in pollen-
367 nectar stores from *Bombus* foragers and pollen from flowers respectively, whereas only 1.2
368 were found in nectar regurgitated from *Bombus* foragers and 0.4 in *Bombus* foragers
369 themselves. It is worth noting that only 85 pesticides were screened for in nectar whereas
370 hundreds were screened in pollen-nectar stores/beebread, pollen and foragers. However,
371 despite the high number of pesticides screened for in foragers, only a few were found.

372 The pesticide residue presence in **pollen-nectar stores/beebread** collected from bees in apple
373 orchards was high in sites located in Italy for *Bombus* and *Osmia* species and in Germany and
374 the United Kingdom for *Apis* species. It was low in Estonian sites, irrespective of bee species
375 (Figure 3, Table A.3 and A.7). When looking at the pesticide residue presence in pollen-nectar
376 stores/beebread collected from bees in oilseed rape, the least exposed sites were in Estonia
377 for *Apis* and *Osmia* species and in Switzerland for *Bombus* species (Figure 3 and Table A.4). In
378 addition, sites located in Germany and Spain for *Apis* species and in Italy for *Osmia* species
379 were the most exposed according to the indices for pollen-nectar stores/beebread. No
380 country was over-represented in the exposed oilseed rape sites for *Bombus* species. Pesticides
381 that characterised the indices were different between the two crops. For a given crop,
382 different pesticides characterised the indices related to pollen-nectar stores/beebread from
383 the different bee species. In other words, pollen-nectar stores/beebread collected by the
384 three species did not contain the same type of pesticides irrespective of whether sampling
385 sites were in apple orchards or in oilseed rape crops. However, the characterisation of the
386 sites with a higher number of pesticides surrounded by oilseed rape included DMF (one
387 metabolite of the acaricide amitraz) for pollen-nectar stores/beebread collected from *Apis*
388 (3.49 µg/kg) and *Bombus* species (7.9 µg/kg) and the herbicide S-metolachlor for pollen-nectar
389 stores/beebread collected from *Apis* (3.93 µg/kg) and *Osmia* species (122.1 µg/kg).

390 Irrespective of the focal crop, the pesticide residue presence in **pollen collected from flowers**
391 was low in Spanish sites (Figure 3, Tables A.3 and A.4). The insecticide diflubenzuron (17.7 and
392 80 µg/kg, respectively) and the fungicide dimetomorph (15.6 and 58.3 µg/kg, respectively)
393 characterised the sites with a higher number of pesticides for pollen collected from apple
394 orchard and oilseed rape flowers. (Tables A.3 and A.4).

395 Looking at **pollen loads** collected from honeybee colonies in apple orchards, pesticide residue
396 presence was high in sites located in Germany and low in sites located in Spain (Figure 3 and
397 Table A.3). For honeybee pollen loads collected in oilseed rape sites, no sites were over-
398 represented in the highest cluster but Italian sites were over-represented in the lowest cluster
399 (Figure 3 and Table A.4). Different pesticides characterised the indices related to pollen loads
400 in the two crops. In other words, pollen loads collected from honeybee colonies did not
401 contain the same type of pesticides in apple orchards or in oilseed rape crops.

402 According to the indices, the **nectar** samples contained a higher number of pesticides when
403 collected in the United Kingdom sites in apple orchard, and fewer pesticides in Italian sites in
404 oilseed rape irrespective of the bee species (Figure 3, Tables A.3 and A.4). The characterisation
405 of the sites with a higher number of pesticides in apple orchards included the fungicide
406 epoxyconazole (2.43 µg/kg in nectar collected by honeybees). It was also present in nectar
407 (2.7 µg/kg) regurgitated from bumblebees collected in by oilseed rape sites and characterised
408 the sites with a higher number of pesticides.

409 When looking at pesticides present in **bees** collected from apple orchards sites, the indices
410 indicated that sites located in the United Kingdom had the highest number of pesticides and
411 those located in Estonia had the lowest, irrespective of the bee species (Figure 3, Tables A.3
412 and A.4). The pesticide residue presence in bees in oilseed rape crops was low in Irish sites for
413 *Apis* species and in Spanish sites for *Bombus* species (Figure 3, Tables A.3, A.4, A.7 and A.8).
414 No country was over-represented with respect to oilseed rape in the most exposed (in terms
415 of number of detected pesticides) sites. The characterisation of the most exposed sites in
416 apple orchards included the pesticide 1,2,3,6 tetrahydrophthalimide (metabolite of a foliar
417 fungicide Captan) for bees collected from both species (700.2 µg/kg in honeybees and 2 170

418 $\mu\text{g}/\text{kg}$ in bumblebees). It was also present in bumblebees collected in the most exposed sites
419 in oilseed rape crops ($197 \mu\text{g}/\text{kg}$). The insecticide tau-fluvalinate characterised the most
420 exposed sites in oilseed rape crops independently of the bee species. The fungicide boscalid
421 characterised the most exposed sites in both crops for bees collected from *Apis* species (176
422 $\mu\text{g}/\text{kg}$ in apple site and $275.2 \mu\text{g}/\text{kg}$ in oilseed rape sites).

423 For indices related to the matrices collected in apple orchards, the clusters of sites with the
424 highest rank of exposure included sites from either Germany, Italy or the United Kingdom
425 (Figure 2). The clusters with the lowest rank of exposure included sites from either Estonia or
426 Spain. Irish and Swiss sites were never over-represented in clusters for these indices. For the
427 indices related to the matrices from sites in by oilseed rape crops, the clusters of sites with
428 the highest rank of exposure included sites from either Germany, Italy or Spain. The clusters
429 with the lowest rank of exposure included sites from either Estonia, Ireland, Italy, Spain or
430 Switzerland. The United Kingdom and Swedish sites were never over-represented in clusters
431 for these indices.

432

433 Figure 2 – Summary of the sites that were most over-represented compared to the mean ($p\text{-value} < 0.05$) in the
434 clusters with low (yellow) and high (blue) number of pesticides based on IRT index values for the nine matrices.
435 Sites in apple orchards are at the top of the figure, whereas those in oilseed rape are below. The bars mean that
436 no sites were over-represented compared to the mean in a cluster.

437

438 **3.2 Links between the indices**

439 The links between indices were illustrated by means of a PCA for matrices collected in apple
440 orchards and in oilseed rape crops (Figure 3). The PCA correlation circles of variables (left
441 plots) represented the link between the nine indices related to each matrix for a given crop.

442 The plots on the right represent the 64 sites, the country being considered as a supplementary
443 information. In data from apple orchard sites, 74.8% of the overall inertia was explained.
444 Inertia is the overall information contained in the data. The remaining 15.6% of missing values
445 were imputed. In data from oilseed rape sites, 51.3% of the overall inertia was explained. The
446 remaining 10.8% of missing values were imputed.

447 Irrespective of the crop (Figure 3), the positive correlations between the nine indices meant
448 that the number of pesticides measured in the various matrices varied in the same way. As
449 indices and number of pesticides were highly correlated (section 3.2.2), the more detected
450 pesticides there were in any given matrix, the more there were in related matrices. However
451 detected pesticides were hardly the same.

452 Figure 3 – Graphical display of the first two components of the Principal Component Analysis of the nine indices
453 (left) from the 64 sites (right) in **apple orchards** (A) or **oilseed rape crops** (B), the country being considered as a
454 supplementary information. The interpretation arrows indicate the nature of the matrices regarding their
455 content of fat (lipophilic, they attract molecules that dissolve in fats) and water (hydrophilic, they attract
456 molecules soluble in water – see discussion for details) and their level pesticide content (low or high number of
457 pesticides – details are given in the text).

458

459 In the apple orchard sites (Figure 3A left), two bundles of variables were highlighted: on one
460 hand, indices related to nectar regurgitated from *Apis* and *Bombus* foragers and to *Apis* and
461 *Bombus* foragers themselves, and on the other hand, indices related to pollen-nectar
462 stores/beebeard collected from colonies and nests, pollen collected from flowers and pollen
463 loads from *Apis* traps. The indices related to nectar were highly correlated with each other
464 ($cor=0.69$) as well as with bumblebees ($cor=0.47$ for *Nectar.Apis/Bombus* and $cor=0.60$ for
465 *Nectar.Bombus/Bombus*). The indices related to pollen-nectar stores/beebeard collected in

466 honeybee or in bumblebee colonies were highly correlated with each other ($cor=0.83$) and, to
467 a lesser extent, to the one collected in solitary bee nests ($cor=0.79$ for *Pollen-nectar*
468 *stores.Osmia/Beebread.Apis* and $cor=0.83$ for *Pollen-nectar stores.Osmia/Pollen-nectar*
469 *stores.Bombus*). These three indices related to pollen-nectar stores/beebead were also linked
470 with the pollen collected from flowers ($cor=0.72$ to 0.75) and with the pollen loads collected
471 from *Apis* traps ($cor=0.65$ to 0.72).

472 Some Italian apple orchard sites were the most exposed for pollen collected from flowers and
473 from *Apis* traps, pollen-nectar stores/beebead collected in colonies and nests from the three
474 bee species and honeybee foragers, whereas some the United Kingdom sites were the most
475 exposed for nectar regurgitated from both bee species and bumblebee foragers (Figure 3A
476 right). In Estonian, Spanish and Swedish sites, pesticide were less found in the matrices in
477 general. In some countries (Ireland, Italy and Sweden), the levels of exposure were highly
478 variable, whereas in others (Estonia, Spain) the levels were homogeneous.

479 In the oilseed rape sites (Figure 3B left), three bundles of variables were highlighted: (i) indices
480 related to pollen-nectar stores/beebead and pollen from flowers, (ii) indices related to *Apis*
481 and *Bombus* foragers, and (iii) indices related to nectar regurgitated from foragers and pollen
482 from *Apis* traps. The indices were less correlated than indices from the apple orchard sites. In
483 the oilseed rape sites, the indices related to nectar were correlated with each other ($cor=0.63$
484 for *Nectar.Apis* and *Nectar.Bombus*). The indices related to pollen-nectar stores/beebead
485 (*Beebread.Apis*, *Pollen-nectar stores.Bombus* and *Pollen-nectar stores.Osmia*) were
486 moderately correlated with each other ($cor=0.31$ to 0.45). These three indices related to
487 pollen-nectar stores/beebead were also slightly correlated to the pollen collected from

488 flowers ($\text{cor}=0.11$ with *Beebread.Apis*, $\text{cor}=0.23$ with *Pollen-nectar stores.Bombus* and
489 $\text{cor}=0.41$ with *Pollen-nectar stores.Osmia*).

490 Italian sites, and to a lesser extent, the German, Spanish and Swiss sites contained the highest
491 number of pesticides for pollen from flowers and pollen-nectar stores/bee bread. In Estonian
492 and Irish sites the matrices contained the lowest number of pesticides in general (Figure 3B
493 right). In some countries (Germany and Sweden) the number of detected pesticides was highly
494 variable whereas in some others (Italy and Spain), it was rather homogeneous.

495

496 **4. Discussion and conclusions**

497 While several surveys have explored the presence of pesticides at the same time in different
498 matrices [19, 34, 50], none proposed an index to characterise the exposure to pesticides. In
499 this paper, we presented a highly novel statistical method using the IRT models to summarise
500 complex information on pesticide presence into a single, yet interpretable, index.

501

502 **4.1 Indices from IRT models: strengths, adaptation and limits**

503 This index illustrated the exposure to pesticides. It was more informative than a classic
504 assessment of richness or abundance because it took into account the overall repartition of
505 pesticides between samples together with quantities of pesticides. This index made possible
506 the calculation of clusters based on similarity or dissimilarity of samples in terms of pesticide
507 detection. As a consequence, comparison between sites (based on pesticide detection in the
508 different samples collected in a given site) was possible.

509 Before choosing IRT models, different statistical methods were considered to reduce the
510 complexity of the 18 datasets that originated from bee exposure to apple orchards and oilseed
511 rape crops including the Multiple Correspondence Analysis (MCA) [51] applied on the overall
512 distance matrix [52]. Contrary to the indices summarising the exposure to infectious and
513 parasitic agents (IPAs) [53], the MCA was not adapted to deal with the multidimensionality of
514 our data, as there was a very slow decay of eigenvalues due to the strong association between
515 sites and pesticides. The proposed indices revealed a structure related to the number of
516 pesticides detected on the sites, illustrated by the linear link between the number of
517 pesticides detected and the exposure level of the sites (the index). The clustering of the sites
518 based on the indices showed a clear separation between the clusters (Tables A.3 and A.4).

519

520 **4.2 Links between matrices and species**

521 When designing the site network, one goal was to explore land-use management across
522 countries and across agroecosystems, resulting in a gradient of exposure to pesticides [40].
523 The land-use management data will be used in forthcoming statistical analyses. Eight countries
524 from four biogeographic zones and two crops were included in the site network. The country
525 of origin was not considered for the index calculation. However, this additional information
526 was very useful to explain the different exposure levels at the sites. Applied to our dataset,
527 the indices showed that in general, matrices collected in apple orchards contained a higher
528 number of pesticides than matrices collected in oilseed rape crops. For a given matrix and a
529 given country, different pesticides characterised the exposure at the sites according to crop
530 exposure. These differences resulted from the crop treatments that were also different from
531 country to country, most probably because of weather constraints and the blooming stage

532 when sampling was performed. However, other factors may explain the diversity of pesticide
533 uses across European countries such as the type of soils, the cultural habits and the
534 commercial strategies from the pesticide industry.

535 In all cases, further statistical analysis is needed to compare the pesticide residue results to
536 the real use of pesticides in the different countries. In other words, it would be worth
537 investigating if, in the example of bees, the 1,2,3,6 tetrahydrophthalimide was more applied
538 on apple orchards in the United Kingdom sites than in Estonian sites. Statistical analysis could
539 focus on field treatments recorded during PoshBee; and on the theoretical number of
540 formulations with a market authorisation in these countries. To our knowledge, such
541 comparison has never been made.

542 In general, the same countries had the most exposed (Germany and Italy) or the least exposed
543 sites (Estonia, Spain) irrespective of the analysed matrix and the crop. However, there was
544 some variation in pesticide detection between matrices for example between beebread
545 collected in *Apis* bees and nectar regurgitated from *Apis* bees in oilseed rape sites located in
546 Italy and Spain. These results show the difference of use and application of pesticides between
547 European countries. This could be further explored with analyses including additional data on
548 pesticide availability in the European countries. Our results also give first insights in the
549 pathway of the contamination chain to understand the source and effect of pesticide residues
550 on bees as aimed at by the site network [40]. For a given site, all matrices contained similar
551 number of pesticides but not necessarily by the same pesticides.

552 At apple orchard sites, the PCA highlighted the discrimination between pollen-nectar
553 stores/beebread and pollen indices from nectar and bee indices. This separation was expected
554 due to the high fat content of pollen-nectar stores/beebread and pollen and the high water

555 content of nectar. This matrix discrimination was independent of country. To our surprise, the
556 indices from the bee matrices (honeybees and bumblebees) were associated with the
557 hydrophilic matrix (regurgitated nectars) rather than lipophilic matrix. It should be noted that
558 this discrimination is based on pesticide numbers, as mentioned before. To further understand
559 the matrix partition, it would be worth looking at the type of pesticides found in the sites, and
560 checking if their chemical characteristics (lipophilicity, use of pKa) are in accordance with the
561 discrimination of the matrices.

562 Consistently across bee species, sites were exposed at the same level for a given matrix. Some
563 pesticides were in common, but in general the detected pesticides were different between
564 the bee species. The three focal bee species selected in this study differ in foraging distances
565 from <1 km for solitary bees [54] up to 6 km for honeybees [55] and foraging preferences.
566 Thus, they probably foraged to different extents on the two focal crops, other flowering crops
567 and wild plants, contributing to different detected pesticide exposure levels. This question will
568 be further explored with the palynological data analysis of pollen-nectar stores/beebread and
569 published in future papers.

570 The number of samples collected from *Osmia* bees were either reduced (for the pollen-nectar
571 stores) or absent (for the regurgitated nectar and for the bee bodies). This was an unfortunate
572 side-effect of the ecology and biology of this species. If the difficulty to retrieve this matrix
573 could be overcome, it would be worth examining the characteristics of pesticides (family,
574 active ingredients and quantities) found in *Osmia* pollen-nectar stores compared to the ones
575 found in pollen-nectar stores/beebread from the other two bee species.

576 Although there was a tendency for the UK, German, and Italian sites to be the most exposed
577 and the Spanish and Estonian sites the least exposed, there were exceptions according to

578 matrices. For example, sites located in Italy were the least exposed when looking at the
579 pesticide residue presence in nectar regurgitated from *Apis* and *Bombus* foragers and pollen
580 loads collected from *Apis* traps following oilseed rape exposure (Tables A.1 to A.4).

581

582 **4.3 Chemicals analysis as a key point to compare results on pesticide detection**

583 The four laboratories involved in the analyses used different methods with large variation of
584 screened pesticides depending on the extraction procedures and the analytical devices used
585 [31, 56]. Ring tests between the different analytical laboratories could be implemented to
586 produce comparable results. This preliminary work should be taken into consideration in
587 future surveys. Usually, stock standard solutions are used to calibrate the analytical devices,
588 with ready-to-use solutions containing several active ingredients. The non-availability of these
589 stock standard solutions depending on the countries was a key point, preventing from having
590 a common list of active ingredients screened for across the four laboratories. However, the
591 list of 64 common pesticides to be screened in all the matrices defined before analyses
592 enabled statistical comparisons when looking at analytical results. Many pesticides were
593 included in the lists of screened pesticides and of those relatively few were found in the
594 matrices – maximum 37% in beebread collected from honeybee colonies (Table 1). These
595 results show that more reflection should be made on targeting analyses to reduce the number
596 of screened pesticides without impairing analytical relevancy. Indeed chemical analyses have
597 potentially important economic and ecological costs.

598

599 **4.4 Risk posed by pesticide residue presence in various matrices**

600 The IRT-based indices focused on bee exposure, not on risk assessment. However, considering
601 the toxicity of detected pesticides is key for the assessment of pesticide risks for different bee
602 species [57] and is linked to the quantities of pesticides in the different matrices. The
603 pesticides significant for discrimination (Table A.6) were mainly fungicides (70% in matrices
604 collected in apple orchard sites, and 43.4% in those surrounded by apple). The proportion was
605 the other way around for insecticides, more frequently found in apple orchard sites compared
606 to oilseed rape. Being more toxic to bees, the exposure to insecticides puts bees more at risk
607 than fungicide exposure. However, quantities and exposure scenarios are also important and
608 should be integrated in the calculation of risk indicators. It would be interesting to explore
609 whether the sites would be similarly clustered for pesticide risk, e.g., assessment based on
610 hazard quotients [34, 50, 58, 59] as regards to exposure, and if correlation between matrices
611 would be similar. In other words, would the risk posed by pollen-nectar stores consumption
612 to bumblebees be positively correlated to the risk posed by beebread consumption to
613 honeybees? Such statistical work should be further explored. Another way to look at these
614 data would be to explore the correlation between the cumulative concentrations of pesticides
615 and the IRT-based indices for each site. If there was a correlation, we could discuss the notion
616 of toxicity. It would be very interesting to have a comparison between cumulative
617 concentrations and added toxic units such as toxicity-weighted concentration [60, 61].

618 Future studies could further assess whether pesticide residue exposure was related to bee
619 population traits recorded in the field [40] along with further potential stressors of bee health
620 [62]. In a previous study, we proposed an index calculation to summarise the exposure to IPAs
621 [53]. The two kinds of indices (IPA and pesticide exposure) could be related to each other or
622 used in structural modeling equations to understand the drivers of bee health. PoshBee data
623 from the site network made it possible to assess pollinator development under field

624 conditions, which is likely more informative for real world scenarios than tests conducted in
625 laboratory conditions [63]. Comparing the pesticides found in the different matrices is also of
626 importance and should be conducted in future statistical works.

627 To conclude, the index calculation based on the IRT methodology presented in this paper is
628 reliable and offers many applications. The characterisation of sampling sites based on the
629 number of detected pesticides across different matrices enabled us to summarise information
630 from complex samples into a single and interpretable index. Our results show that although
631 pesticide numbers were similar in matrices from any given country irrespective of bee species,
632 some important variations could be observed. Therefore, for a complete assessment of
633 pollinator pesticide exposure, it is necessary to consider several different exposure routes and
634 multiple species of bees across different agricultural systems. Other parameters should be
635 considered such as bee population traits, different pesticide and application use between
636 countries, other potential stressors of bee health. However all these information are usually
637 lacking in field studies.

638 These results highlight the variation in the use and application of pesticides across European
639 countries. This could be further explored with analyses including additional data on pesticide
640 availability in the European countries. Our results also give first insights in the pathway of the
641 contamination chain to understand the source and effect of pesticide residues on bees as
642 aimed at by the site network [40]. For a given site, all matrices experienced similar number of
643 pesticides but not by the same pesticides or in comparable quantities.

644 Beyond such summarisation of complex data, the indices can be used in many ways, e.g. to
645 compare and explore the correlation between matrices. Our datasets and matrices offer

646 important opportunities for statistical analyses to examine relationships of the presented IRT
647 indices with risks posed by pesticides to pollinators or their influence on bee health.

648

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663

664 **References**

- 665 1 Potts, S. G., Biesmeijer, J. C., Kremen, C., Neumann, P., Schweiger, O., Kunin, W. E. 2010
666 Global pollinator declines: trends, impacts and drivers. *Trends Ecol Evol.* **25**, 345-353.
667 (<https://doi.org/10.1016/j.tree.2010.01.007>)
668 2 IPBES, Potts, S. G., Imperatriz-Fonseca, V. L., Ngo, H. T., Biesmeijer, J. C., breeze, T. D.,
669 Dicks, L. V., Garibaldi, L. A., Hill, R., Settele, J., *et al.* 2016 *Summary for policymakers of*

670 *the assessment report of the Intergovernmental Science-Policy Platform on Biodiversity and*
671 *Ecosystem Services on pollinators, pollination and food production.* Bonn, Germany:
672 Secretariat of the Intergovernmental Science-Policy Platform on Biodiversity and Ecosystem
673 Services.

674 3 Kulhanek, K., Steinhauer, N., Rennich, K., Caron, D. M., Sagili, R. R., Pettis, J. S., Ellis, J.
675 D., Wilson, M. E., Wilkes, J. T., Tarpy, D. R., *et al.* 2017 A national survey of managed
676 honey bee 2015–2016 annual colony losses in the USA. *J. Apic. Res.* **56**, 328-340.
677 (<https://doi.org/10.1080/00218839.2017.1344496>)

678 4 CAPA. 2022 CAPA statement on Colony Losses in Canada. 24 pages.
679 (<https://capabees.com/capa-statement-on-honey-bees/>)

680 5 van der Zee, R., Pisa, L., Andonov, S., Brodschneider, R., Charrière, J.-D., Chlebo, R.,
681 Coffey, M. F., Crailsheim, K., Dahle, B., Gajda, A., *et al.* 2012 Managed honey be colony
682 losses in Canada, China, Europe, Israel and Turkey, for the winters of 2008-9 and 2009-10. *J.*
683 *Apic. Res.* **51**, 100-114. (<https://doi.org/10.3896/IBRA.1.51.1.12>)

684 6 Maggi, M., Antunez, K., Invernizzi, C., Aldea, P., Vargas, M., Negri, P., Brasesco, C., de
685 Jong, D., Message, D., Teixeira, E. W., *et al.* 2016 Honeybee health in South America.
686 *Apidologie.* (<https://doi.org/10.1007/s13592-016-0445-7>)

687 7 Pirk, C. W., Human, H., Crewe, R. M., Vanengelsdorp, D. 2014 A survey of managed
688 honey bee colony losses in the Republic of South Africa–2009 to 2011. *J. Apic. Res.* **53**, 35-
689 42. (10.3896/IBRA.1.53.1.03)

690 8 Moritz, R. F. A., Erler, S. 2016 Lost colonies found in a data mine: Global honey trade but
691 not pests or pesticides as a major cause of regional honeybee colony declines. *Agric. Ecosyst.*
692 *Environ.* **216**, 44-50. (<https://doi.org/10.1016/j.agee.2015.09.027>)

693 9 Soroye, P., Newbold, T., Kerr, J. 2020 Climate change contributes to widespread declines
694 among bumble bees across continents. *Science.* **367**, 685-688.
695 (<https://doi.org/10.1126/science.aax8591>)

696 10 Powney, G. D., Carvell, C., Edwards, M., Morris, R. K. A., Roy, H. E., Woodcock, B. A.,
697 Isaac, N. J. B. 2019 Widespread losses of pollinating insects in Britain. *Nature*
698 *communications.* **10**, 1018. (<https://doi.org/10.1038/s41467-019-08974-9>)

699 11 Pritchard, Z. A., Hendriksma, H. P., St Clair, A. L., Stein, D. S., Dolezal, A. G., O’Neal,
700 M. E., Toth, A. L. 2021 Do Viruses From Managed Honey Bees (Hymenoptera: Apidae)
701 Endanger Wild Bees in Native Prairies? *Environ. Entomol.*,
702 (<https://doi.org/10.1093/ee/nvaa181>)

703 12 Oddie, M. A. Y., Burke, A., Dahle, B., Le Conte, Y., Mondet, F., Locke, B. 2021
704 Reproductive success of the parasitic mite (*Varroa destructor*) is lower in honeybee colonies
705 that target infested cells with recapping. *Sci. Rep.* **11**, ([https://doi.org/10.1038/s41598-021-](https://doi.org/10.1038/s41598-021-88592-y)
706 [88592-y](https://doi.org/10.1038/s41598-021-88592-y))

707 13 Rollin, O., Pérez-Méndez, N., Bretagnolle, V., Henry, M. 2019 Preserving habitat quality
708 at local and landscape scales increases wild bee diversity in intensive farming systems. *Agric.*
709 *Ecosyst. Environ.* **275**, 73-80. (<https://doi.org/10.1016/j.agee.2019.01.012>)

710 14 Ollerton, J., Erenler, H., Edwards, M., Crockett, R. 2014 Pollinator declines. Extinctions of
711 aculeate pollinators in Britain and the role of large-scale agricultural changes. *Science.* **346**,
712 1360-1362. (<https://doi.org/346/6215/1360>)

713 15 Somme, L., Moquet, L., Quinet, M., Vanderplanck, M., Michez, D., Lognay, G.,
714 Jacquemart, A. L. 2016 Food in a row: urban trees offer valuable floral resources to
715 pollinating insects. *Urban Ecosyst.* (<https://doi.org/10.1007/s11252-016-0555-z>)

716 16 Di Pasquale, G., Alaux, C., Le Conte, Y., Odoux, J. F., Pioz, M., Vaissiere, B. E.,
717 Belzunces, L. P., Decourtye, A. 2016 Variations in the Availability of Pollen Resources
718 Affect Honey Bee Health. *PLoS One.* **11**, e0162818.
719 (<https://doi.org/10.1371/journal.pone.0162818>)

- 720 17 Siviter, H., Richman, S. K., Muth, F. 2021 Field-realistic neonicotinoid exposure has sub-
721 lethal effects on non-Apis bees: A meta-analysis. *Ecol Lett.* **24**, 2586-2597.
722 (<https://doi.org/10.1111/ele.13873>)
- 723 18 Zioga, E., Kelly, R., White, B., Stout, J. C. 2020 Plant protection product residues in plant
724 pollen and nectar: A review of current knowledge. *Environ. Res.* **189**, 109873.
725 (<https://doi.org/10.1016/j.envres.2020.109873>)
- 726 19 Ward, L. T., Hladik, M. L., Guzman, A., Winsemius, S., Bautista, A., Kremen, C., Mills,
727 N. J. 2022 Pesticide exposure of wild bees and honey bees foraging from field border flowers
728 in intensively managed agriculture areas. *Sci. Total Environ.* **831**, 154697.
729 (<https://doi.org/10.1016/j.scitotenv.2022.154697>)
- 730 20 Straw, E. A., Brown, M. J. F. 2021 Co-formulant in a commercial fungicide product causes
731 lethal and sub-lethal effects in bumble bees. *Sci. Rep.* **11**, 21653.
732 (<https://doi.org/10.1038/s41598-021-00919-x>)
- 733 21 Graham, K. K., Milbrath, M. O., Zhang, Y., Baert, N., McArt, S., Isaacs, R. 2022 Pesticide
734 risk to managed bees during blueberry pollination is primarily driven by off-farm exposures.
735 *Sci. Rep.* **12**, 7189. (<https://doi.org/10.1038/s41598-022-11156-1>)
- 736 22 Rundlof, M., Andersson, G. K., Bommarco, R., Fries, I., Hederstrom, V., Herbertsson, L.,
737 Jonsson, O., Klatt, B. K., Pedersen, T. R., Yourstone, J., *et al.* 2015 Seed coating with a
738 neonicotinoid insecticide negatively affects wild bees. *Nature.* **521**, 77-80.
739 (<https://doi.org/10.1038/nature14420>)
- 740 23 Knapp, J. L., Nicholson, C. C., Jonsson, O., de Miranda, J. R., Rundlöf, M. 2023
741 Ecological traits interact with landscape context to determine bees' pesticide risk. *Nature*
742 *Ecology & Evolution.* **7**, 547-556. (<https://doi.org/10.1038/s41559-023-01990-5>)
- 743 24 Wisk, J. D., Pistorius, J., Beevers, M., Bireley, R., Browning, Z., Chauzat, M. P.,
744 Nikolakis, A., Overmyer, J. P., Rose, R., Sebastien, R., *et al.* 2014 Assessing the exposure of
745 pesticides to bees. In *Pesticide risk assessment for pollinators*. (ed. ^eds. D. Fischer, T.
746 Moriarty), pp. 45-74: Wiley Blackwell.
- 747 25 Carter, L. J., Agatz, A., Kumar, A., Williams, M. 2020 Translocation of pharmaceuticals
748 from wastewater into beehives. *Environ. Int.* **134**, 105248.
749 (<https://doi.org/10.1016/j.envint.2019.105248>)
- 750 26 McCune, F., Samson-Robert, O., Rondeau, S., Chagnon, M., Fournier, V. 2021 Supplying
751 honey bees with waterers: a precautionary measure to reduce exposure to pesticides.
752 *Environmental Science and Pollution Research.* **28**, 17573-17586.
753 (<https://doi.org/10.1007/s11356-020-12147-3>)
- 754 27 Arena, M., Sgolastra, F. 2014 A meta-analysis comparing the sensitivity of bees to
755 pesticides. *Ecotoxicology.* **23**, 324-334. (<https://doi.org/10.1007/s10646-014-1190-1>)
- 756 28 Negri, I., Mavris, C., Di Prisco, G., Caprio, E., Pellicchia, M. 2015 Honey Bees (*Apis*
757 *mellifera*, L.) as Active Samplers of Airborne Particulate Matter. *PLoS One.* **10**, e0132491.
758 (<https://doi.org/10.1371/journal.pone.0132491>)
- 759 29 Pochi, D., Biocca, M., Fanigliulo, R., Pulcini, P., Conte, E. 2012 Potential exposure of
760 bees, *Apis mellifera* L., to particulate matter and pesticides derived from seed dressing during
761 maize sowing. *Bull. Environ. Contam. Toxicol.* **89**, 354-361.
- 762 30 Mahefarisoa, K. L., Simon Delso, N., Zaninotto, V., Colin, M. E., Bonmatin, J. M. 2021
763 The threat of veterinary medicinal products and biocides on pollinators: A One Health
764 perspective. *One Health.* **12**, 100237. (<https://doi.org/10.1016/j.onehlt.2021.100237>)
- 765 31 Kiljanek, T., Niewiadowska, A., Malysiak, M., Posyniak, A. 2021 Miniaturized
766 multiresidue method for determination of 267 pesticides, their metabolites and
767 polychlorinated biphenyls in low mass beebread samples by liquid and gas chromatography
768 coupled with tandem mass spectrometry. *Talanta.* **235**, 122721.
769 (<https://doi.org/10.1016/j.talanta.2021.122721>)

770 32 Pavlova, D., Atanassova, J., Karadjova, I., Bani, A. 2022 Pollen and Chemical Content of
771 Beebreads from Serpentine Areas in Albania and Bulgaria. *Biol. Trace Elem. Res.* **200**, 413-
772 425. (<https://doi.org/10.1007/s12011-021-02638-w>)

773 33 Kavanagh, S., Henry, M., Stout, J. C., White, B. 2021 Neonicotinoid residues in honey
774 from urban and rural environments. *Environmental Science and Pollution Research*.
775 (<https://doi.org/10.1007/s11356-021-12564-y>)

776 34 Favaro, R., Bauer, L. M., Rossi, M., D'Ambrosio, L., Bucher, E., Angeli, S. 2019
777 Botanical Origin of Pesticide Residues in Pollen Loads Collected by Honeybees During and
778 After Apple Bloom. *Front. Physiol.* **10**, 1069. (<https://doi.org/10.3389/fphys.2019.01069>)

779 35 Raimets, R., Bontsutsnaja, A., Bartkevics, V., Pugajeva, I., Kaart, T., Puusepp, L., Pihlik,
780 P., Keres, I., Viinalass, H., Mand, M., *et al.* 2020 Pesticide residues in beehive matrices are
781 dependent on collection time and matrix type but independent of proportion of foraged oilseed
782 rape and agricultural land in foraging territory. *Chemosphere.* **238**, 124555.
783 (<https://doi.org/10.1016/j.chemosphere.2019.124555>)

784 36 Mullin, C. A., Frazier, M., Frazier, J. L., Ashcraft, S., Simonds, R., Vanengelsdorp, D.,
785 Pettis, J. S. 2010 High levels of miticides and agrochemicals in North American apiaries:
786 implications for honey bee health. *PLoS One.* **5**, e9754-.

787 37 Martinello, M., Manzinello, C., Borin, A., Avram, L. E., Dainese, N., Giuliano, I., Gallina,
788 A., Mutinelli, F. 2020 A Survey from 2015 to 2019 to Investigate the Occurrence of Pesticide
789 Residues in Dead Honeybees and Other Matrices Related to Honeybee Mortality Incidents in
790 Italy. *Diversity.* **12**, (<https://doi.org/10.3390/d12010015>)

791 38 Demares, F. J., Schmehl, D., Bloomquist, J. R., Cabrera, A. R., Huang, Z. Y., Lau, P.,
792 Rangel, J., Sullivan, J., Xie, X., Ellis, J. D. 2022 Honey Bee (*Apis mellifera*) Exposure to
793 Pesticide Residues in Nectar and Pollen in Urban and Suburban Environments from Four
794 Regions of the United States. *Environ Toxicol Chem.* **41**, 991-1003.
795 (<https://doi.org/10.1002/etc.5298>)

796 39 Traynor, K. S., Tosi, S., Rennich, K., Steinhauer, N., Forsgren, E., Rose, R., Kunkel, G.,
797 Madella, S., Lopez, D., Eversole, H., *et al.* 2021 Pesticides in honey bee colonies:
798 Establishing a baseline for real world exposure over seven years in the USA. *Environ Pollut.*
799 **279**, 116566. (<https://doi.org/10.1016/j.envpol.2021.116566>)

800 40 Hodge, S., Schweiger, O., Klein, A. M., Potts, S. G., Costa, C., Albrecht, M., de Miranda,
801 J. R., Mand, M., De la Rúa, P., Rundlöf, M., *et al.* 2022 Design and Planning of a
802 Transdisciplinary Investigation into Farmland Pollinators: Rationale, Co-Design, and Lessons
803 Learned. *Sustainability.* **14**, 10549. (<https://doi.org/10.3390/su141710549>)

804 41 Hodge, S., Stout, J. 2019 Protocols for methods of field sampling Deliverable D1.1.

805 42 Serra, G., Costa, C., Cardaio, I., Colombo, R. 2021 Report on exposure of bees to
806 agrochemicals. Deliverable D2.2.

807 43 Bock, R. D., Aitkin, M. 1981 Marginal maximum likelihood estimation of item
808 parameters: Application of an EM algorithm. . *Psychometrika.* **46**, 443-459.

809 44 Van der Linden, W. J., Hambleton, R. K. 1997 *Handbook of modern item response theory*.
810 New-York, NY: Springer.

811 45 Chalmer, R. P. 2012 mirt: A Multidimensional Item Response Theory Package for the R
812 Environment. *Journal of Statistical Software.* **48**, 1-29. (<https://doi:10.18637/jss.v048.i06>)

813 46 Everitt, B. 1974 *Cluster Analysis*. London: Heinemann Educ. Books.

814 47 Husson, F., Lê, S., Pagès, J. 2017 *Exploratory multivariate analysis by example using R*.
815 CRC press.

816 48 Jolliffe, I. T. 1986 *Principal component analysis*. . New-York: Springer Verlag.

817 49 Lê, S., Josse, J., Husson, F. 2008 FactoMineR: An R Package for Multivariate Analysis.
818 *Journal of Statistical Software.* **25**, 1-18. (<https://doi.org/10.18637/jss.v025.i01>)

819 50 Wen, X., Ma, C., Sun, M., Wang, Y., Xue, X., Chen, J., Song, W., Li-Byarlay, H., Luo, S.
820 2021 Pesticide residues in the pollen and nectar of oilseed rape (*Brassica napus* L.) and their
821 potential risks to honey bees. *Sci. Total Environ.* **786**, 147443.
822 (<https://doi.org/10.1016/j.scitotenv.2021.147443>)
823 51 Greenacre, M. J. 1984 *Theory and applications of correspondence analysis*. London:
824 Academic Press.
825 52 Legendre, P., Legendre, L. 1998 *Numerical Ecology*. Second ed ed. Amsterdam: Elsevier.
826 53 Huyen Ton Nu Nguyet, M., Bougeard, S., Babin, A., Dubois, E., Druesne, C., Rivière, M.
827 P., Laurent, M., Chauzat, M. P. 2023 Building composite indices in the age of big data –
828 Application to honey bee exposure to infectious and parasitic agents. *Heliyon*. e15244.
829 (<https://doi.org/10.1016/j.heliyon.2023.e15244>)
830 54 Zurbuchen, A., Landert, L., Klaiber, J., Müller, A., Hein, S., Dorn, S. 2010 Maximum
831 foraging ranges in solitary bees: only few individuals have the capability to cover long
832 foraging distances. *Biol. Conserv.* **143**, 669-676.
833 (<https://doi.org/10.1016/j.biocon.2009.12.003>)
834 55 Beekman, M., Ratnieks, F. L. W. 2000 Long-range foraging by the honey-bee, *Apis*
835 *mellifera* L. *Functional Ecology*. 2000.
836 56 Martel, A. C., Pierotti, N., Bray, E. 2023 Development and validation of two multiresidue
837 methods for the determination of pesticides in nectar collected by honey bees and bumble
838 bees. *Submitted*.
839 57 Storck, V., Karpouzas, D. G., Martin-Laurent, F. 2016 Towards a better pesticide policy
840 for the European Union. *Sci Total Environ.* (<https://doi.org/10.1016/j.scitotenv.2016.09.167>)
841 58 Thomson, H. M. 2010 Risk assessment for honey bees and pesticides - recent
842 developments and 'new issues'. *Pest Manage. Sci.* **66**, 1157-1162.
843 59 Rortais, A., Arnold, G., Dorne, J. L., More, S. J., Sperandio, G., Streissl, F., Szentes, C.,
844 Verdonck, F. 2017 Risk assessment of pesticides and other stressors in bees: Principles, data
845 gaps and perspectives from the European Food Safety Authority. *Sci Total Environ.*
846 (<https://doi.org/10.1016/j.scitotenv.2016.09.127>)
847 60 Rundlöf, M., Stuligross, C., Lindh, A., Malfi, R. L., Burns, K., Mola, J. M., Cibotti, S.,
848 Williams, N. M. 2022 Flower plantings support wild bee reproduction and may also mitigate
849 pesticide exposure effects. *J. Appl. Ecol.* **59**, 2117-2127. ([https://doi.org/10.1111/1365-](https://doi.org/10.1111/1365-2664.14223)
850 [2664.14223](https://doi.org/10.1111/1365-2664.14223))
851 61 EFSA Scientific Committee, More, S. J., Bampidis, V., Benford, D., Bennekou, S. H.,
852 Bragard, C., Halldorsson, T. I., Hernández-Jerez, A. F., Koutsoumanis, K., Naegeli, H., *et al.*
853 2019 Guidance on harmonised methodologies for human health, animal health and ecological
854 risk assessment of combined exposure to multiple chemicals. *EFSA Journal.* **17**, e05634.
855 (<https://doi.org/10.2903/j.efsa.2019.5634>)
856 62 Breda, D., Frizzera, D., Giordano, G., Seffin, E., Zanni, V., Annoscia, D., Topping, C. J.,
857 Blanchini, F., Nazzi, F. 2022 A deeper understanding of system interactions can explain
858 contradictory field results on pesticide impact on honey bees. *Nature communications.* **13**,
859 5720. (<https://doi.org/10.1038/s41467-022-33405-7>)
860 63 Stanley, D. A., Raine, N. E. 2017 Bumblebee colony development following chronic
861 exposure to field-realistic levels of the neonicotinoid pesticide thiamethoxam under laboratory
862 conditions. *Sci. Rep.* **7**, 8005. (<https://doi.org/10.1038/s41598-017-08752-x>)
863