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THE R1441C MUTATION ALTERS THE FOLDING PROPERTIES OF THE ROC DOMAIN OF LRRK2

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Keywords: LRRK2, ROCO protein, GTPase, Parkinson's disease, differential scanning fluorimetry, circular dichroism

Abstract

LRRK2 is a 250kDa multidomain protein, mutations in which cause familial Parkinson's disease. Previously, we have demonstrated that the R1441C mutation in the ROC domain decreases GTPase activity. Here we show that the R1441C alters the folding properties of the ROC domain, lowering its thermodynamic stability. Similar to small GTPases, binding of different guanosine nucleotides alters the stability of the ROC domain, suggesting that there is an alteration in conformation dependent on GDP or GTP occupying the active site. GTP/GDP bound state also alters the self-interaction of the ROC domain, accentuating the impact of the R1441C mutation on this property. These data suggest a mechanism whereby the R1441C mutation can reduce the GTPase activity of LRRK2, and highlights the possibility of targeting the stability of the ROC domain as a therapeutic avenue in LRRK2 disease.

Abbreviations: DSF, differential scanning fluorimetry; CD, circular dichroism; GDP, guanosine diphosphate; GTP, guanosine triphosphate; LRRK2, leucine rich repeat kinase 2; ROC, Ras of complex proteins; PD, Parkinson's disease

1. Introduction

Leucine rich repeat kinase 2 (LRRK2) is a member of the ROCO family of proteins, defined by the presence of a ROC (Ras of complex proteins) domain followed by a domain of unknown function termed C-terminal of ROC, or COR [1]. In LRRK2, these domains are flanked towards the C- and N- termini respectively by leucine rich repeat (LRR) and kinase domains that give this protein its name, along with a number of protein/protein interaction domains [2]. The function of LRRK2 is unknown, but several mutations in this protein have been shown to cause autosomal dominant Parkinson's disease (PD) [3, 4]. Overall, mutations in LRRK2 are the most common genetic cause of PD [5, 6]. Due to their recent description and large size, very little is known about the biochemistry and function of the ROCO proteins. Studies focusing on the enzymatic activities of LRRK1 and LRRK2 have suggested that the GTPase activity of these proteins regulates their kinase activity, in a manner analogous to the interaction of small GTPases and associated kinases such as Ras and Raf [7-9]. Studies of mutations linked to Parkinson's disease have shown that the kinase activity of LRRK2 is required for cytotoxicity associated with mutations [10-14]. However, while mutations in the kinase domain have a modest activating effect on kinase activity, mutations in the ROC domain of LRRK2 have not consistently been shown to affect kinase activity [15]. These same ROC domain mutations consistently disrupt the GTPase activity of the protein, which might lead to altered regulation of kinase activity, although the mechanistic basis for this is unclear [16-19]. These studies highlight the role of the enzymatic activities

of LRRK2 in the disease process, but the exact functional and spatial relationship between these two domains is unknown.

We have previously demonstrated that the R1441C mutation decreases the GTPase activity of LRRK2 in the context of the full-length protein [16]. The crystal structure of LRRK2 ROC domain showed that the R1441 residue is distal to the active site of the protein and revealed that the R1441 residue sits at the interface between two constituent monomers in a dimeric structure [19]. This observation suggests that mutations at this residue could destabilize the interaction between the monomers, which might be the mechanism of enzymatic dysfunction. Consistent with this observation, we demonstrated that the ROC domain containing the R1441C mutation has a decreased ability to precipitate the full-length protein from cell lysates. However, the recent report of the crystal structure of a fragment of a ROCO protein from the prokaryote *Chlorobium Tepidium* consisting of the ROC and COR domains (the *C. Tepidium* ROCO protein does not contain a kinase domain) suggested that the role of R1441 is mainly inter-domain association. This alternative model implied that mutations at LRRK2 R1441 should not perturb the structure of the ROC GTPase domain itself, and instead acts to disrupt its interaction with the COR domain [20]. To further investigate the mechanism whereby the R1441C mutation disrupts ROC/ROC interactions and how it decreases the GTPase activity of LRRK2, we have used a variety of biophysical approaches including circular dichroism spectropolarimetry and differential scanning fluorimetry to assess the folding characteristics of the ROC domain and to test whether the R1441C mutation

alters these characteristics.

2. Methods and materials

2.1 Protein production and purification

The ROC domain of LRRK2 was expressed and purified as previously reported [21]. Where only the wild type protein was used, the 6x polyhistidine tag was removed as described [21]. The R1441C mutant protein was expressed and purified as a 6xHis tagged fusion and was used to compare with the 6xHis tagged WT ROC protein. Attempts to remove the 6xhis tag from the mutant protein failed due to the drastically decreased stability of the protein (data not shown). Attempts to purify a GTP binding dead (K1347A) mutant form of the protein in a soluble form were also unsuccessful (data not shown).

2.2 Circular dichroism

Spectra were recorded with a Jasco J715 spectropolarimeter and measured at a protein concentration of 1 mg ml⁻¹ with a 0.01 cm path length. The spectra are an average of 50 scans at 25°C, with background molecular ellipticity due to the buffer subtracted, in the presence of 4mM GDP. Analysis of protein stability was carried out with a protein concentration of 0.1 mg ml⁻¹ and a 5mm path length (measurements were taken over a 10 nm bandwidth centered around 220 nm). Unfolding was carried out by melting over a temperature gradient from 25°C to 85°C, ramping by 1° every minute. The ellipticity signal (h) was converted to

proportion of molecules in the native state α_N , according to the relationship $\alpha_N = (\theta - \theta_U) / (\theta_N - \theta_U)$, where θ_U and θ_N are the ellipticity signals for the unfolded and native states, respectively. Data were fitted to the Van't Hoff function and the mid-point of thermal denaturation (T_m) defined as DH/DS.

2.3 Differential scanning fluorimetry

Thermal denaturation curves were also obtained using a Bio-Rad iCycler Single Wavelength Real-Time PCR Detection System (*Bio-Rad Laboratories, Hercules, CA*) and SYPRO Orange fluorescence dye (*Invitrogen, Carlsbad, CA*) with an excitation and emission wavelength of 490nm and 520nm, respectively. The 6xHis tagged ROC domain proteins (WT and R1441C) were subjected to a thermal gradient of 0.5°C increments from 23°–75°C. The 50µl reaction consisted of 0.25mg/ml protein in 100mM HEPES, 150mM NaCl, 20mM MgCl₂, pH 7.5 and a 2x SYPRO Orange final concentration diluted from concentrated stock. Guanine nucleotide ligands (GDP, GTP and GppNP) were added at a final concentration of 2mM accordingly. All experiments were performed with five replicates. The T_m for each sample was calculated using the first derivative of the normalized fluorescence.

2.4 Pulldown Assay

Purified WT and R1441C 6xhis tagged ROC proteins were mixed with excess amounts of purified WT v5-tagged ROC. The protein mixtures were then incubated with Ni-NTA superflow resin for 1 hour at 4°C, in buffer A containing

20mM Tris, 500mM NaCl, 20mM Imidazole, 20 mM MgCl₂, 10% Glycerol, pH 8.0. Different guanine nucleotide ligands (GDP, GTP and GppNP) were added to a final concentration of 2 mM accordingly. After extensive wash with buffer A, proteins were eluted from Ni-NTA. Co-purified proteins were analyzed by SDS-PAGE and western blotting with biotinylated anti-V5 antibody (1:10000). Streptavidin conjugated HRP (1:2000) were used for developing the membrane. The SDS gels were scanned by CanonPI CS-U 3.8.1x scanner and the western blot was developed and scanned on Alpha Innotech FluoChem Instrument. The intensities of the bands were integrated and recorded using the software Kodak Digital Science 1D 3.0.2 analysis package. Pulldowns were replicated 3 times.

3. Results

To examine the secondary structure and folding properties of the ROC domain, we used two approaches - circular dichroism spectropolarimetry and differential scanning fluorimetry. CD analysis showed that the ROC domain forms ordered secondary structure in solution, with an absorbance spectrum suggestive of a predominantly α -helical fold with double minima at λ 208 and 222 (figure 1A). This is in agreement with data from the crystal structure of the ROC domain, which revealed 46.5% α -helix and 21.6% β -pleated sheet, and with molecular modeling studies based upon the small GTPases [21]. Using graduated thermal melting, and following the percentage of folded protein using absorbance at 220nm with CD, the ROC domain was found to unfold in a co-operative manner (figure 1B). The mid-point of thermal denaturation or T_m determined by the loss of α -helical

structure was 41°C for the ROC domain in water in the absence of nucleotides. Using DSF to report the exposure of the hydrophobic core in the absence of nucleotides, the ROC domain unfolded with a very similar T_m of 39°C (figure 1C). As previously demonstrated with H-Ras, the presence of guanosine nucleotides has a major impact on the stability of the LRRK2 ROC domain (figure 1D), increasing the T_m to 52.5°C and 50.4°C in the presence of GDP and GDP respectively [22]. The R1441C mutation caused a destabilization of the ROC domain, decreasing the T_m by 4° in the presence of GDP and 5.5° in the presence of GTP as compared to the wild type domain (figure 1D and table 1).

Based upon previous work examining LRRK2 self interaction using LRRK2 and fragments expressed in mammalian cell culture [23], we assessed ROC-ROC domain interactions between WT homodimers and WT/R1441C heterodimers with purified protein in solution, mimicking the situation in humans with heterozygous mutations. Using ROC domains with different epitope tags (6xHistag and V5), a significant decrease was observed in the interaction between mutant and WT domains (figure 2). The interaction between WT and R1441C was lower still in the presence of GTP.

4. Discussion

We have previously shown that the R1441C mutation in the LRRK2 ROC domain results in a decrease in GTPase activity. Based on the model of a dimeric ROC-ROC structure, we predicted that the mutated residue sits at the interface in a dimeric structure for the ROC domain. In the current study, we sought to test the

hypothesis that the R1441C mutation decreases the thermodynamic stability of the ROC domain and to investigate the impact of guanosine nucleotides on the folding, stability and self-interaction of the ROC domain.

We assessed how the R1441C mutation and binding of guanosine nucleotides influence the folding properties of the ROC domain in solution. Using CD to assess secondary structure, we found that the ROC domain of LRRK2 forms a predominantly α -helical secondary structure in solution, agreeing with data from crystallographic studies. Although of low resolution and describing only secondary structure, this is the first report of any structural information for LRRK2 in solution. Using molecular ellipticity as a marker for percent folded protein in solution, we carried out thermal denaturation of the ROC domain, yielding a calculated T_m of 39°C. Using DSF to measure unfolded protein, we examined the impact of ligand binding and the R1441C mutation on the stability of the ROC domain. As previously shown with single domain GTPases, $MgCl_2$ and guanosine nucleotides substantially increase the stability of the ROC domain, with GDP having a more pronounced impact than GTP [22]. This suggests, indirectly, an alteration in the three dimensional structural organization of the domain between the GDP and GTP bound states, and supports the thesis that the ROC domain is acting as a molecular switch [9]. This finding requires further investigation using high-resolution structural approaches. Intriguingly, the R1441C mutation consistently and significantly lowered the stability of the ROC in the presence of either GTP or GDP, suggesting that this mutation acts to destabilize the ROC domain. These data suggest that R1441 plays an important

role in the structural integrity of the GTPase domain itself, which is consistent with the crystal structural data of LRRK2 ROC dimer [21]. Due to the fact that the R1441 residue is not conserved between the ROCO proteins and the small GTPases such as a Ras and Rho, it is not possible to study or compare the impact of alterations at this residue in these proteins. We went on to examine ROC domain self-interaction in the light of data showing the impact of GDP and GTP on the stability of the ROC domain using differentially tagged ROC domain constructs. Results from these experiments demonstrate that the strength of interaction between differentially tagged ROC domain monomers is altered in the presence of GDP or GTP, that these have additive effects on stability of the ROC domain in the presence of the R1441C mutation. These data are consistent with structural information indicating that the R1441C mutation is distal from the GTP binding site and suggests that there are separate effects of guanine nucleotides and the R1441C mutation. These results also support previous data from our group looking at the interaction between full length LRRK2 from mammalian cell lysates and the ROC domain in the presence and absence of the mutation [21]. Both the stability data and the pulldown data suggest that there is an alteration in the three dimensional structure of the ROC domain based upon whether GDP or GTP is bound in the active site, again consistent with studies carried out on the small GTPases [24]. We are currently investigating this further using a variety of techniques. It is important to note that these studies need extending to larger, multi-domain fragments of LRRK2 in order to shed light on the nature and location of the interactions between the components of the LRRK2 dimer. To

date, we have not been able to purify fragments larger than the ROC domain that are stable in solution, and so such further studies wait upon overcoming this technical hurdle.

In summary, our investigations into the folding, stability and self interaction properties of the ROC domain suggests a potential mechanism for dysfunction and loss of GTPase activity of the ROC domain based upon a decrease in stability associated with the R1441C mutation found in familial Parkinson's disease. They represent the first description of a folding analysis of any of the domains of LRRK2. Future experiments dissecting the relationship between stability and function may shed light on the underlying cause of LRRK2 dysfunction linked to ROC domain mutations, and it will be important to extend these studies using larger fragments of LRRK2 and other mutations. These data also highlight the possibility of targeting the stability of the LRRK2 ROC domain as a potential therapeutic avenue to correcting the dysfunction of this domain, artificially reverting the destabilizing impact of the R1441C mutation.

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competing interest.

Figure Legends

Figure 1 A) CD analysis of the wild type ROC domain, displaying absorption spectra typical of a predominantly α -helical fold B, C) Thermal melt data from the ROC domain in the absence of ligand using CD (B) and DSF (C) to follow loss of structure D) DSF measurement of protein stability in the presence of GDP and GTP for wild type and R1441C ROC domain

Figure 2 Pulldown analysis of the affinity of wild type and R1441C ROC domain for each other showing a significant decrease in the ability of ROC domain containing the R1441C mutation to precipitate the wild type domain. Analysis was carried out by two-way ANOVA with variant and ligand as separate variables, using a Bonferroni *post hoc* test. Indicated P values are **<0.01 and ***<0.001.

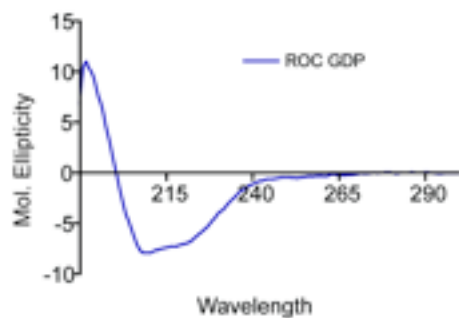
Table 1 T_m values for ROC +/- R1441C and GDP/GTP. Differences in stability due to presence of GDP or GTP for WT ($P < 0.005$) and R1441C ($P < 0.005$) were significant as assessed by Welch's T-Test, with differences due to the presence of mutation (i.e. WT vs R1441C) in the case GDP ($P < 0.0001$) or GTP ($P < 0.000005$) also being significant.

References

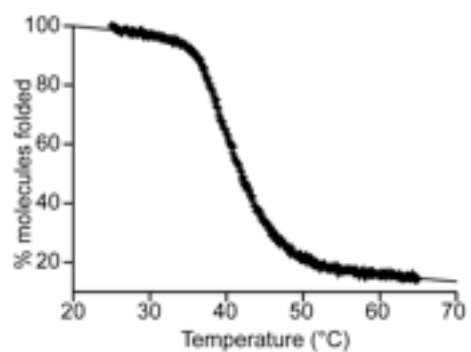
- [1] L. Bosgraaf, P.J. Van Haastert, Roc, a Ras/GTPase domain in complex proteins, *Biochim Biophys Acta* 1643 (2003) 5-10.
- [2] I. Marin, The Parkinson disease gene LRRK2: evolutionary and structural insights, *Molecular biology and evolution* 23 (2006) 2423-2433.
- [3] C. Paisan-Ruiz, S. Jain, E.W. Evans, W.P. Gilks, J. Simon, M. van der Brug, A. Lopez de Munain, S. Aparicio, A.M. Gil, N. Khan, J. Johnson, J.R. Martinez, D. Nicholl, I.M. Carrera, A.S. Pena, R. de Silva, A. Lees, J.F. Marti-Masso, J. Perez-Tur, N.W. Wood, A.B. Singleton, Cloning of the gene containing mutations that cause PARK8-linked Parkinson's disease, *Neuron* 44 (2004) 595-600.
- [4] A. Zimprich, S. Biskup, P. Leitner, P. Lichtner, M. Farrer, S. Lincoln, J. Kachergus, M. Hulihan, R.J. Uitti, D.B. Calne, A.J. Stoessl, R.F. Pfeiffer, N. Patenge, I.C. Carbajal, P. Vieregge, F. Asmus, B. Muller-Myhsok, D.W. Dickson, T. Meitinger, T.M. Strom, Z.K. Wszolek, T. Gasser, Mutations in LRRK2 cause autosomal-dominant parkinsonism with pleomorphic pathology, *Neuron* 44 (2004) 601-607.
- [5] L.N. Clark, Y. Wang, E. Karlins, L. Saito, H. Mejia-Santana, J. Harris, E.D. Louis, L.J. Cote, H. Andrews, S. Fahn, C. Waters, B. Ford, S. Frucht, R. Ottman, K. Marder, Frequency of LRRK2 mutations in early- and late-onset Parkinson disease, *Neurology* 67 (2006) 1786-1791.
- [6] C. Paisan-Ruiz, LRRK2 gene variation and its contribution to Parkinson disease, *Hum Mutat* 30 (2009) 1153-1160.
- [7] D. Korr, L. Toschi, P. Donner, H.D. Pohlentz, B. Kreft, B. Weiss, LRRK1 protein kinase activity is stimulated upon binding of GTP to its Roc domain, *Cell Signal* 18 (2006) 910-920.
- [8] G. Ito, T. Okai, G. Fujino, K. Takeda, H. Ichijo, T. Katada, T. Iwatsubo, GTP Binding Is Essential to the Protein Kinase Activity of LRRK2, a Causative Gene Product for Familial Parkinson's Disease, *Biochemistry* 46 (2007) 1380-1388.
- [9] B. Weiss, ROCO kinase activity is controlled by internal GTPase function, *Science signaling* 1 (2008) pe27.
- [10] A.B. West, D.J. Moore, S. Biskup, A. Bugayenko, W.W. Smith, C.A. Ross, V.L. Dawson, T.M. Dawson, Parkinson's disease-associated mutations in leucine-rich repeat kinase 2 augment kinase activity, *Proc Natl Acad Sci U S A* 102 (2005) 16842-16847.
- [11] C.J. Gloeckner, N. Kinkl, A. Schumacher, R.J. Braun, E. O'Neill, T. Meitinger, W. Kolch, H. Prokisch, M. Ueffing, The Parkinson disease causing LRRK2 mutation I2020T is associated with increased kinase activity, *Hum Mol Genet* 15 (2006) 223-232.
- [12] E. Greggio, S. Jain, A. Kingsbury, R. Bandopadhyay, P. Lewis, A. Kaganovich, M.P. van der Brug, A. Beilina, J. Blackinton, K.J. Thomas, R. Ahmad, D.W. Miller, S. Kesavapany, A. Singleton, A. Lees, R.J. Harvey, K. Harvey, M.R. Cookson, Kinase activity is required for the toxic effects of mutant LRRK2/dardarin, *Neurobiol Dis* 23 (2006) 329-341.

- [13] W.W. Smith, Z. Pei, H. Jiang, V.L. Dawson, T.M. Dawson, C.A. Ross, Kinase activity of mutant LRRK2 mediates neuronal toxicity, *Nat Neurosci* 9 (2006) 1231-1233.
- [14] M. Jaleel, R.J. Nichols, M. Deak, D.G. Campbell, F. Gillardon, A. Knebel, D.R. Alessi, LRRK2 phosphorylates moesin at Thr558; characterisation of how Parkinson's disease mutants affect kinase activity, *Biochem J* (2007).
- [15] E. Greggio, M.R. Cookson, Leucine Rich Repeat Kinase 2 mutations and Parkinson's disease: Three Questions, *ASN Neuro* doi:10.1042/AN20090007 (2009).
- [16] P.A. Lewis, E. Greggio, A. Beilina, S. Jain, A. Baker, M.R. Cookson, The R1441C mutation of LRRK2 disrupts GTP hydrolysis, *Biochem Biophys Res Commun* 357 (2007) 668-671.
- [17] L. Guo, P.N. Gandhi, W. Wang, R.B. Petersen, A.L. Wilson-Delfosse, S.G. Chen, The Parkinson's disease-associated protein, leucine-rich repeat kinase 2 (LRRK2), is an authentic GTPase that stimulates kinase activity, *Exp Cell Res* 313 (2007) 3658-3670.
- [18] A.B. West, D.J. Moore, C. Choi, S.A. Andrabi, X. Li, D. Dikeman, S. Biskup, Z. Zhang, K.L. Lim, V.L. Dawson, T.M. Dawson, Parkinson's disease-associated mutations in LRRK2 link enhanced GTP-binding and kinase activities to neuronal toxicity, *Hum Mol Genet* 16 (2007) 223-232.
- [19] X. Li, Y.C. Tan, S. Poulouse, C.W. Olanow, X.Y. Huang, Z. Yue, Leucine-rich repeat kinase 2 (LRRK2)/PARK8 possesses GTPase activity that is altered in familial Parkinson's disease R1441C/G mutants, *J Neurochem* (2007).
- [20] K. Gotthardt, M. Weyand, A. Kortholt, P.J. Van Haastert, A. Wittinghofer, Structure of the Roc-COR domain tandem of *C. tepidum*, a prokaryotic homologue of the human LRRK2 Parkinson kinase, *Embo J* 27 (2008) 2352.
- [21] J. Deng, P.A. Lewis, E. Greggio, E. Sluch, A. Beilina, M.R. Cookson, Structure of the ROC domain from the Parkinson's disease-associated leucine-rich repeat kinase 2 reveals a dimeric GTPase, *Proc Natl Acad Sci U S A* 105 (2008) 1499-1504.
- [22] J. Zhang, C.R. Matthews, Ligand binding is the principal determinant of stability for the p21(H)-ras protein, *Biochemistry* 37 (1998) 14881-14890.
- [23] E. Greggio, I. Zambrano, A. Kaganovich, A. Beilina, J.M. Taymans, V. Daniels, P. Lewis, S. Jain, J. Ding, A. Syed, K.J. Thomas, V. Baekelandt, M.R. Cookson, The Parkinson disease-associated leucine-rich repeat kinase 2 (LRRK2) is a dimer that undergoes intramolecular autophosphorylation, *J Biol Chem* 283 (2008) 16906-16914.
- [24] I.R. Vetter, A. Wittinghofer, The guanine nucleotide-binding switch in three dimensions, *Science* 294 (2001) 1299-1304.

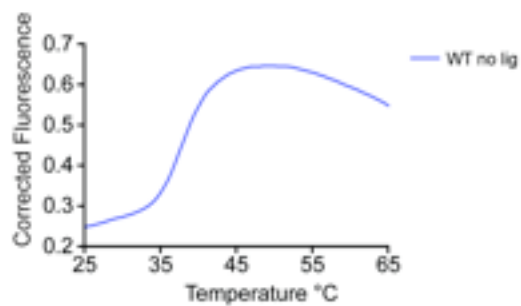
A



B



C



D

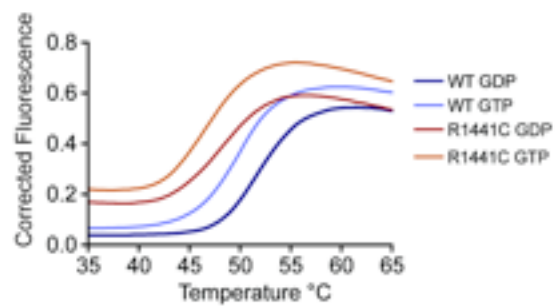
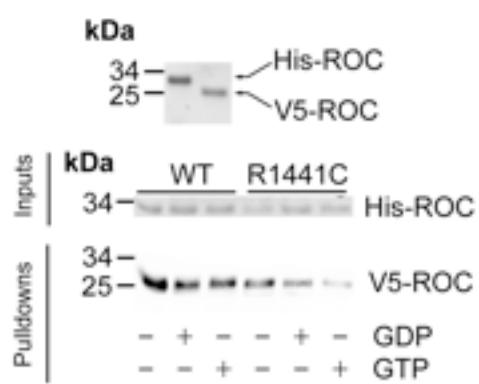


Figure 1

A



B

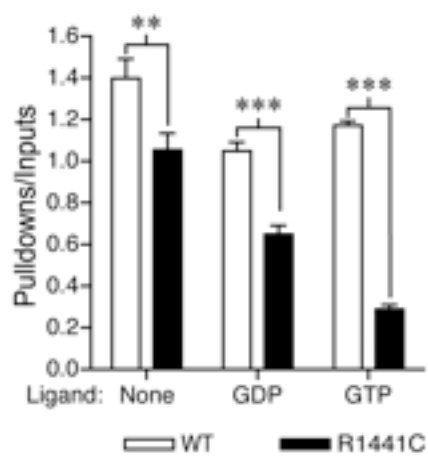


Figure 2

	Wild type	R1441C
GDP	52.5±0.8	47.8±0.3
GTP	50.4±0.2	46.6±0.4
GppNp	40.1±0.3	N/A
No ligand	37.7±0.1	N/A

Table 1