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1	Earthworm-produced calcite granules: a new terrestrial palaeothermometer?
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14	
15	Abstract
16	In this paper we show for the first time that calcite granules, produced by the earthworm
17	Lumbricus terrestris, and commonly recorded at sites of archaeological interest, accurately
18	reflect temperature and soil water δ^{18} O values. Earthworms were cultivated in an orthogonal
19	combination of two different (granule-free) soils moistened by three types of mineral water
20	and kept at three temperatures (10, 16 and 20 °C) for an acclimatisation period of three weeks
21	followed by transfer to identical treatments and cultivation for a further four weeks.
22	Earthworm-secreted calcite granules were collected from the second set of soils. δ^{18} O values
23	were determined on individual calcite granules ($\delta^{18}O_c$) and the soil solution ($\delta^{18}O_w$). The
24	$\delta^{18}O_c$ values reflect soil solution $\delta^{18}O_w$ values and temperature, but are consistently enriched

by $1.51 (\pm 0.12)$ ‰ in comparison to equilibrium in synthetic carbonates. The data fit the

equation 1000 ln α = [20.21 ± 0.92] (10³ T⁻¹) - [38.58 ± 3.18] (R² = 0.95; n = 96; p < 0.0005). As the granules are abundant in modern soils, buried soils and archaeological contexts, and can be dated using U-Th disequilibria, the developed palaeotemperature relationship has enormous potential for application to Holocene and Pleistocene time intervals.

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1. INTRODUCTION

32 Many organisms form reliable archives of palaeotemperature or water composition, by precipitating calcium carbonate ($CaCO_3$) in oxygen isotopic equilibrium with their 33 environment (Ullmann et al., 2010; Versteegh et al., 2010; Wanamaker et al., 2007). The 34 δ^{18} O values of the CaCO₃ are controlled by temperature and water isotope composition 35 (Epstein et al., 1953; Urey, 1947), with equilibrium being defined via a mineral-specific 36 fractionation factor (Kim and O'Neil, 1997). The δ^{18} O composition of CaCO₃ produced by 37 some biomineralising taxonomic groups shows a systematic offset from equilibrium, and as 38 such is still useful as a proxy (Ford et al., 2010; Rosenheim et al., 2009; Ziveri et al., 2003). 39 The CaCO₃ minerals produced by other taxonomic groups are not in equilibrium with the 40 environment, but exhibit deviations, called vital effects (Correa et al., 2010; Juillet-Leclerc et 41 al., 2009; Owen et al., 2002). Therefore, when a biomineral is investigated as a potential 42 palaeoenvironmental proxy, it is important to develop species-specific and well-constrained 43 palaeotemperature relationships. 44

Although not widely appreciated, many earthworm species are true biomineralisers (Briones
et al., 2008b) and produce calcite granules in specialised glands (Canti, 1998; Lee et al.,
2008b) first noted by Darwin (1881; Fig. 1). The lob worm *Lumbricus terrestris* is common
in Europe and increasingly as an invasive species in the USA and Canada. It is one of the
major producers of earthworm-secreted calcite granules in temperate soils. As such a large
majority of these distinctly shaped granules, that are commonly found in soils, will have been

51 produced by this species (Canti, 2007). The granules are formed in the calciferous glands of 52 the earthworm, occurring in segments 10-12 as three pairs of swellings off the oesophagus (Canti, 1998). The function that granule secretion serves is unknown, with suggestions 53 ranging from excretion of excess calcium as a reaction against calcium toxicity of soils, to 54 neutralisation of gut pH and regulation of CO₂ (Bal, 1977; Becze-Deák et al., 1997; Crang et 55 al., 1968; Darwin, 1881; Piearce, 1972; Robertson, 1936). Data show that granules 56 incorporate both dietary and atmospheric C (Briones et al., 2008b; Canti, 2009). They have 57 been reported from the Pleistocene (Green et al., 2006; Meijer, 1985), but can likely be 58 preserved for longer (Lambkin et al., 2011). Preliminary data suggest they can be dated using 59 U-Th disequilibria: granules recovered from Silbury Hill, a Neolithic monument in Wiltshire, 60 61 UK (Atkinson, 1967), which has an earliest date of 4400 BP produce a U-Th disequilibrium age of 4670 ± 440 years (own data). Further U-Th analyses are ongoing and applications and 62 limitations will be discussed in a future publication. ¹⁴C dating of earthworm granules is 63 possible if enough material is available from the same stratigraphic unit and yields ages that 64 agree with other carbonates (Pustovoytov and Terhorst, 2004). Due to their abundance and 65 good preservation, calcite $\delta^{18}O(\delta^{18}O_c)$ values from these granules form a potentially 66 powerful palaeotemperature proxy. A limited number of oxygen isotope analyses have 67 recently been published on earthworm calcite, showing similar values as directly precipitated 68 secondary carbonates, but no systematic investigation into temperature relationships has yet 69 70 been performed (Koeniger et al., 2012; Pustovoytov and Terhorst, 2004). Therefore, the aim of the experimental work presented here was to investigate this potential by testing the 71 hypothesis that the δ^{18} O values recorded in the earthworm secreted granules vary 72 systematically with soil solution δ^{18} O values and temperature. 73

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2. METHODS

We investigated the utility of calcite $\delta^{18}O_c$ values of the earthworm-secreted granules as a palaeothermometer by means of a laboratory experiment with an orthogonal combination of two different types of soil, three types of mineral water (initial $\delta^{18}O$ values -10.0, -7.3 and -6.3 (± 0.2) ‰VSMOW) and three temperatures (10, 16 and 20 °C) with 6 replicates (individual earthworms) per treatment.

81

Soils were collected from agricultural fields in Berkshire, UK: Hamble (SU 61968 70235) a 82 Hamble series Typical Argillic Brown Earth with 1.3 weight% Ca, and Red Hill (SU 56060 83 80033) a Yattendon series Typical Argillic Brown Earth with 0.6 weight% Ca (Avery, 1980; 84 Jarvis, 1968). Following our established methodology for the collection of earthworm 85 86 secreted granules produced over the course of an experiment, the soils were air-dried and 87 sieved to 250 µm prior to use (Lambkin et al., 2011). This ensures soils are granule-free initially, and facilitates granule recovery at the end of the experiments. For each replicate, 88 300 g of soil were mixed with one of 3 different types of mineral water to 65 % water holding 89 capacity (BS ISO, 1998). This was put in a zip-lock bag with 5 g air-dried horse manure 90 rehydrated with 10 ml of the same mineral water. One adult *L. terrestris* was added to each 91 bag, and they were placed in one of three constant temperature rooms at 10, 16, or 20 °C in 92 darkness. There were 6 replicates for each treatment. A scoping study indicated that within 3 93 weeks exposure to new isotopic conditions the oxygen isotopic composition of the granules 94 95 had reached a steady state. Therefore earthworms were acclimatised for three weeks, and then transferred to an identical treatment bag containing the same type and mass of soil, manure 96 and water at the same temperature. After 28 days earthworms were removed and soil solution 97 98 was extracted using rhizon samplers (Van Walt Micro Rhizon; Dickens et al., 2007; Kölling et al., 2005; Seeberg-Elverfeldt et al., 2005). The samplers produce water with a water vapour 99 pressure of 2340 Pa at ambient temperature (20°C) and hence any fractionation of the water 100

isotopes during this process will be minimal. The soil was wet-sieved to 500 µm to retrievegranules, which were air-dried and weighed.

Oxygen isotope analyses on the original mineral waters added and soil solutions extracted at 103 the end of the experiment were performed on a Picarro L2120-i Isotopic Water Analyzer with 104 an A0211 High-Precision Vaporizer and ChemCorrect software. Values were calibrated 105 against reference standards IA-R052, IA-R053 and IA-R054 from Iso-Analytical Limited. 106 Long-term reproducibility was < 0.2 %. Nine soils did not yield enough water for analysis. In 107 order to incorporate granule data from these in our analysis we used the average water δ^{18} O 108 $(\delta^{18}O_w)$ value from the replicates of the same treatment. Calcite granules were individually 109 analysed for $\delta^{18}O_c$ values, using a Thermo Delta V Advantage IRMS with a GasBench II. 110 The raw $\delta^{18}O_c$ values were converted into the VPDB scale after normalising against NBS18 111 and NBS19 carbonate standards. The long-term standard deviation of a routinely analysed in-112 house $CaCO_3$ standard was < 0.1 ‰. Soil solution pH was measured with a combination 113 micro-electrode connected to a Hanna pH21 pH/mV meter and calibrated using pH 4.0 and 114 7.0 buffers. Ca^{2+} concentrations in soil solutions were determined using a Perkin Elmer 115 Optima 3000 ICP-OES. Operational blanks had Ca^{2+} concentrations below detection (16 116 µg/l). Data were calibrated via analysis of 1, 50 and 100 ppm dilutions of the Merck ICP 117 multi-element standard solution IV and accuracy determined through analysis of an in house 118 standard. 119

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3. RESULTS

122 Results are shown in Table 1 and Fig. 2. At the end of the 4 week exposure, soil solution 123 $\delta^{18}O_w$ values ranged from -10.2 to -5.3 % VSMOW. Some influence of evaporation was 124 observed, causing higher $\delta^{18}O_w$ values for higher temperatures. This is most pronounced for 125 the Hamble soil, probably because of its lower water holding capacity (Table 1, Fig. 2). $\delta^{18}O_c$

126	values of individual granules (n = 931) vary between -10.10 and -3.21 ‰VPDB. Within-	
127	treatment $\delta^{18}O_c$ values of granules are normally distributed. The variability in $\delta^{18}O_c$ values	
128	between granules produced by an individual earthworm within a single month is on average	
129	0.61 ‰ (range 1.44 ‰). Higher soil solution $\delta^{18}O_w$ values yield higher $\delta^{18}O_c$ values, and	
130	higher temperatures result in lower $\delta^{18}O_c$ values (Fig. 2).	
131		
132	4. DISCUSSION	
133	For each replicate the expected $\delta^{18}O_c$ value for equilibrium was calculated from the $\delta^{18}O_w$	
134	value and temperature according to the equation of Kim and O'Neil (1997):	
135		
136	$1000 \ln \alpha = 18.03 \ (10^3 \mathrm{T}^{-1}) - 32.42 \tag{1}$	
137		
138	The resulting predicted $\delta^{18}O_c$ ($\delta^{18}O_{pred}$) values are plotted with average measured $\delta^{18}O_c$ per	
139	replicate in Figure 3. A linear regression shows that earthworm granules are systematically	
140	enriched in ¹⁸ O by 1.51 (\pm 0.12 s.d.) ‰ in comparison to equilibrium:	
141		
142	$\delta^{18}O_c = 1.02 \ \delta^{18}O_{pred} + 1.51 \tag{2}$	
143		
144	with $R^2 = 0.98$; n = 96; p < 0.0005.	
145	Subsequently, the fractionation factor α was derived:	
146		
147	$\alpha_{\text{calcite-water}} \left(\left[1000 + \delta^{18} O_c VSMOW \right] / \left[1000 + \delta^{18} O_w VSMOW \right] \right) $ (3)	
148		
149	and plotted against 10^3 T^{-1} (K). Regression analysis yields the palaeotemperature relationship:	4. 4.
150		

151 1000 ln
$$\alpha = [20.21 \pm 0.92] (10^3 \text{ T}^{-1}) - [38.58 \pm 3.18]$$

with $R^2 = 0.95$; n = 96; p < 0.0005. Quoted errors on the slope and intercept are reported at 153 the 95 % confidence interval. Our results are compared to equations by Kim & O'Neil (1997: 154 synthetic carbonates) and Wanamaker et al. (2006; 2007; Blue mussel, Mytilus edulis) in 155 Figure 4. *M. edulis* data are slightly offset from equilibrium (< 0.2 %), but this difference is 156 not statistically significant (Wanamaker et al., 2007). $\delta^{18}O_c$ values of calcite granules 157 produced by L. terrestris reflect soil solution $\delta^{18}O_w$ values and temperature, but are enriched 158 in ¹⁸O in comparison to equilibrium in synthetic carbonates (Kim and O'Neil, 1997) and the 159 bivalve M. edulis (Wanamaker et al., 2007). 160 161

Although the relationship is strong, the range of 1000 ln α for a given temperature is about twice that observed for *M. edulis* (Wanamaker et al., 2007) translating into an average offset in reconstructed temperature of 0.73 °C. A possible explanation for this increased variation is that some evaporation occurred during the experiment, progressively increasing soil solution $\delta^{18}O_w$ values throughout the 4-week experiment. In addition, some of the variation could have been caused by the variability of the controlled temperature chambers in which the experiments were performed at \pm 0.5 °C from the set temperature.

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The enrichment in ¹⁸O by 1.51 ‰ compared to equilibrium is statistically indistinguishable from the equation for synthetic calcite precipitated from a solution with initial [Ca²⁺] and [HCO₃⁻] of 15 mM (Fig. 4; Kim and O'Neil, 1997). The soil solutions collected after the experiment had [Ca²⁺] values varying between 8.30 (\pm 1.02 s.d.) mM and 16.79 (\pm 1.42 s.d.) mM with higher values for higher temperatures, for mineral waters with higher initial [Ca²⁺], and for the Red Hill soil. Average pH values were 6.5 \pm 0.8 s.d. (Fiji), 6.8 \pm 0.5 s.d.

176 (England) and 7.0 \pm 0.3 s.d. (Norway; Table 1). pH did not significantly differ between temperatures or soils. At the measured pH values, most DIC will be present in the form of 177 HCO₃⁻ (Zeebe and Wolf-Gladrow, 2001). HCO₃⁻ was not determined directly, but 178 concentrations of dissolved inorganic carbon (DIC) are likely to have been high (up to 32 179 mM based on equivalence of charge of DIC with Ca^{2+}), due to earthworm and microorganism 180 respiration (601-1329 µmol/mol CO₂ in soil air, own data). As such, it appears that elevated 181 initial $[Ca^{2+}]$ and $[HCO_3^{-}]$ caused the ¹⁸O enrichment in earthworm granules, through the 182 same unknown mechanism as in synthetic calcite. Earthworms do not produce calcite 183 granules at low $[Ca^{2+}]$ (own data) or low pH (Lambkin et al., 2011), and thus the range of 184 these parameters in soils containing granules will be constrained at the lower end. Therefore, 185 different soils are unlikely to yield radically different palaeotemperature equations. 186 187 A possible alternative or additional explanation for the 1.51 ‰ offset of the granules lies in 188 precipitation kinetics. Calcite precipitation in the calciferous gland likely takes place along a 189 pathway of dissolved CO₂ transforming to HCO_3^{-1} and CO_3^{-2} , then to amorphous CaCO₃. 190 which finally stabilises into calcite through a dissolution-reprecipitation mechanism (Briones 191 et al., 2008a; Lee et al., 2008a). If calcite precipitation were very fast, there would be 192 incomplete fractionation between HCO_3^- and $CaCO_3$, resulting in ¹⁸O enrichment of 193 earthworm granules in comparison to equilibrium (kinetic effect) (Lachniet, 2009; Mickler et 194 al., 2004). The precipitation rate of calcite varied between 1.24 (\pm 0.47 s.d.) and 2.89 (\pm 0.31 195 s.d.) mg/day and was higher for higher temperatures and the Red Hill soil, and lower for 196 Norway water (Table 1). However, no relationship between precipitation rate and $\delta^{18}O_c$ 197 values was found. 198

Several recent publications on speleothems and inorganically precipitated calcite suggest that $\alpha_{calcite-water}$ is greater than the commonly accepted value by up to 1.5 % (Coplen, 2007; Day and Henderson, 2011; Dietzel et al., 2009; Feng et al., 2012; Tremaine et al., 2011). Although these systems are very different from the earthworm calciferous gland, it is possible, that they have a common cause for disequilibrium, such as fractionation reactions at the crystal surface.

206

For application in palaeotemperature reconstructions, a good estimate of soil water $\delta^{18}O_w$ 207 values is needed. Due to the combined influence of seasonal variations in precipitation $\delta^{18}O_w$ 208 values and evapotranspiration, these vary over time and with depth (Hsieh et al., 1998). 209 However, in a high-rainfall temperate climate, like that of the British Isles, the influence of 210 evaporation is likely to be small. This is supported by data showing that the $\delta^{18}O_w$ 211 composition of groundwater reflects that of local precipitation within 0.5 ‰ (Darling et al., 212 2003) and that $\delta^{18}O_c$ values of modern soil carbonate are correlated with $\delta^{18}O_w$ values of 213 local meteoric water (Cerling, 1984). For the past 4000 years, modern values for the isotopic 214 composition of precipitation can be used (McDermott et al., 2011). For earlier Holocene and 215 Pleistocene time intervals, $\delta^{18}O_w$ values of precipitation are often not well-constrained and 216 the earthworm-calcite palaeothermometer can only be used if an independent reconstruction 217 of soil water $\delta^{18}O_w$ values is available. Alternatively, the equation can serve as an 218 independent proxy for soil water $\delta^{18}O_w$ values (and hence $\delta^{18}O_w$ values of precipitation) in 219 the presence of existing palaeotemperature reconstructions. 220

221

222

5. CONCLUSIONS

In summary, our study shows that the relationship between temperature and earthworm calcite $\delta^{18}O_c$ values is significant and predictable. In combination with the wide distribution,

225	good preservation, and direct U-Th dating of the granules, these will make a novel terrestrial
226	temperature proxy. Important questions to be addressed are if this relationship holds under
227	field conditions when evaporation of soil moisture may occur, and for other earthworm
228	species. Future work will investigate these issues and also include application of the
229	developed palaeothermometer to granules retrieved from various Holocene and Pleistocene
230	locations and archaeological sites in Great Britain, the Netherlands and Germany.
231	
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239	
240	REFERENCES
241	Atkinson R.J.C. (1967) Silbury Hill. Antiquity XLI, 259-262.
242	Avery B.W. (1980) Soil classification for England and Wales [higher categories].
243	Rothamsted Experimental Station, Harpenden.
244	Bal L. (1977) The formation of carbonate nodules and intercalary crystals in the soil by the
245	earthworm Lumbricus rubellus. Pedobiologia 17, 229-240.
246	Becze-Deák J., Langohr R., Verrecchia E.P. (1997) Small scale secondary CaCO ₃
247	accumulations in selected sections of the European loess belt. Morphological forms
248	and potential for paleoenvironmental reconstruction. Geoderma 76, 221-252.

240	Briones MII	I ánaz E	Mándaz I	Podríguez I B	Gago-Duport L.	(2008a)	Riological
249	DITORIES IVI.J.I.,	, Lopez E.	, Menuez J.,	KOULIQUEZ J.D.,	Gago-Dupon L.	(2000a)	Diological

250 control over the formation and storage of amorphous calcium carbonate by

251 earthworms. *Mineral. Mag.* **72**, 227-231.

252 Briones M.J.I., Ostle N.J., Piearce T.G. (2008b) Stable isotopes reveal that the calciferous

253 gland of earthworms is a CO_2 -fixing organ. *Soil Biol. Biochem.* **40**, 554-557.

BS ISO (1998) Soil quality - Determination of the water-retention characteristic. BS ISO,
Geneva.

Canti M.G. (1998) Origin of calcium carbonate granules found in buried soils and Quaternary
 deposits. *Boreas* 27, 275-288.

258 Canti M.G. (2007) Deposition and taphonomy of earthworm granules in relation to their

interpretative potential in Quaternary stratigraphy. J. Quatern. Sci. 22, 111-118.

260 Canti M.G. (2009) Experiments on the origin of ¹³C in the calcium carbonate granules

261 produced by the earthworm *Lumbricus terrestris*. *Soil Biol. Biochem.* **41**, 2588-2592.

- Cerling T.E. (1984) The stable isotopic composition of modern soil carbonate and its
 relationship to climate. *Earth Planet. Sci. Lett.* **71**, 229-240.
- Coplen T.B. (2007) Calibration of the calcite–water oxygen-isotope geothermometer at
 Devils Hole, Nevada, a natural laboratory. *Geochim. Cosmochim. Acta* 71, 3948 3957.
- 267 Correa M.L., Montagna P., Vendrell-Simón B., McCulloch M., Taviani M. (2010) Stable 268 isotopes (δ^{18} O and δ^{13} C), trace and minor element compositions of recent
- scleractinians and Last Glacial bivalves at the Santa Maria di Leuca deep-water coral
 province, Ionian Sea. *DSR* 57, 471-486.
- Crang R.E., Holsen R.C., Hitt J.B. (1968) Calcite Production in Mitochondria of Earthworm
 Calciferous Glands. *Bioscience* 18, 299-301.

772	Douling W/C	Doth A II	Talkat IC	(<u>2002)</u> Th	$\sim 0 \sim 1$ II	atable instance	e composition of
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4 13	During W.O.	, Duun Inin,	1 11001 0.0.	(2000) III			

- freshwaters in the British Isles. 2. Surface waters and groundwater. *HESS* **7**, 183-195.
- Darwin C. (1881) *The formation of vegetable mould, through the action of worms, with observations on their habits.* The Echo Library, Teddington.
- Day C.C., Henderson G.M. (2011) Oxygen isotopes in calcite grown under cave-analogue
 conditions. *Geochim. Cosmochim. Acta* **75**, 3956-3972.
- Dickens G.R., Koelling M., Smith D.C., Schnieders L. (2007) Rhizon sampling of pore
 waters on scientific drilling expeditions: an example from the IODP Expedition 302,
- 281 Arctic Coring Expedition (ACEX). *Scientific Drilling* **4**, 22-25.
- 282 Dietzel M., Tang J., Leis A., Köhler S.J. (2009) Oxygen isotopic fractionation during
- inorganic calcite precipitation Effects of temperature, precipitation rate and pH. *Chem. Geol.* 268, 107-115.
- Epstein S., Buchsbaum R., Lowenstam H.A., Urey H.C. (1953) Revised carbonate-water
 isotopic temperature scale. *Bulletin of the Geological Society of America* 64, 13151326.
- Feng W., Banner J.L., Guilfoyle A.L., Musgrove M., James E.W. (2012) Oxygen isotopic
 fractionation between drip water and speleothem calcite: A 10-year monitoring study,
 central Texas, USA. *Chem. Geol.* 304–305, 53-67.
- Ford H.L., Schellenberg S.A., Becker B.J., Deutschman D.L., Dyck K.A., Koch P.L. (2010)
- 292 Evaluating the skeletal chemistry of *Mytilus californianus* as a temperature proxy:
- 293 Effects of microenvironment and ontogeny. *Paleoceanography* **25**.
- 294 Green C.P., Branch N.P., Russell Coope G., Field M.H., Keen D.H., Wells J.M.,
- 295 Schwenninger J.-L., Preece R.C., Schreve D.C., Canti M.G., Gleed-Owen C.P. (2006)
- 296 Marine Isotope Stage 9 environments of fluvial deposits at Hackney, north London,
- 297 UK. Quatern. Sci. Rev. 25, 89-113.

298	Hsieh J.C.C., Chadwick O.A., Kelly E.F., Savin S.M. (1998) Oxygen isotopic composition of
299	soil water: Quantifying evaporation and transpiration. Geoderma 82, 269-293.
300	Jarvis R.A. (1968) Soils of the Reading district. Agricultural Research Council, Harpenden.
301	Juillet-Leclerc A., Reynaud S., Rollion-Bard C., Cuif J.P., Dauphin Y., Blamart D., Ferrier-
302	Pagès C., Allemand D. (2009) Oxygen isotopic signature of the skeletal
303	microstructures in cultured corals: Identification of vital effects. Geochim.
304	Cosmochim. Acta 73 , 5320-5332.
305	Kim ST., O'Neil J.R. (1997) Equilibrium and nonequilibrium oxygen isotope effects in
306	synthetic carbonates. Geochim. Cosmochim. Acta 61, 3461-3475.
307	Koeniger P., Barta G., Thiel C., Bajnóczi B., Novothny Á., Horváth E., Techmer A., Frechen
308	M. (2012) Stable isotope composition of bulk and secondary carbonates from the
309	Quaternary loess-paleosol sequence in Süttő, Hungary. Quatern. Int.
310	Kölling M., Seeberg-Elverfeldt J., Schlüter M. (2005) Rhizon-an excellent pore water
311	sampler for low maintenance collection and filtration of small volume samples. In (
312	Lachniet M.S. (2009) Climatic and environmental controls on speleothem oxygen-isotope
313	values. Quatern. Sci. Rev. 28, 412-432.
314	Lambkin D.C., Gwilliam K.H., Layton C., Canti M.G., Piearce T.G., Hodson M.E. (2011)
315	Production and dissolution rates of earthworm-secreted calcium carbonate.
316	Pedobiologia 54, S119-S129.
317	Lee M.R., Hodson M.E., Langworthy G.N. (2008a) Crystallization of calcite from amorphous
318	calcium carbonate: earthworms show the way. <i>Mineral. Mag.</i> 72 , 257-261.
319	Lee M.R., Hodson M.E., Langworthy G.N. (2008b) Earthworms produce granules of
320	intricately zoned calcite. Geology 36, 943-946.

321	McDermott F., Atkinson T.C., Fairchild I.J., Baldini L.M., Mattey D.P. (2011) A first
322	evaluation of the spatial gradients in δ^{18} O recorded by European Holocene
323	speleothems. Global Planet. Change 79, 275-287.
324	Meijer T. (1985) The pre-Weichselian non-marine molluscan fauna from Maastricht-
325	Belvédère (Southern Limburg, The Netherlands). In Maastricht-Belvédère:
326	Stratigraphy, palaeoenvironment and archaeology of the Middle and Late Pleistocene
327	depositis (Geological Survey of The Netherlands, Haarlem, pp. 75-103.
328	Mickler P.J., Banner J.L., Stern L., Asmerom Y., Edwards R.L., Ito E. (2004) Stable isotope
329	variations in modern tropical speleothems: Evaluating equilibrium vs. kinetic isotope
330	effects. Geochim. Cosmochim. Acta 68, 4381-4393.
331	Owen R., Kennedy H., Richardson C. (2002) Isotopic partitioning between scallop shell
332	calcite and seawater: effect of shell growth rate. Geochim. Cosmochim. Acta 66,
333	1727-1737.
334	Piearce T.G. (1972) The Calcium Relations of Selected Lumbricidae. J. Anim. Ecol. 41, 167-
335	188.
336	Pustovoytov K., Terhorst B. (2004) An isotopic study of a late Quaternary loess-paleosol
337	sequence in SW Germany. Revista Mexicana de Ciencias Geológicas 21, 88-93.
338	Robertson J.D. (1936) The function of the calciferous glands of earthworms. J. Exp. Biol.
339	XIII , 279-297.
340	Rosenheim B.E., Swart P.K., Willenz P. (2009) Calibration of sclerosponge oxygen isotope
341	records to temperature using high-resolution δ^{18} O data. <i>Geochim. Cosmochim. Acta</i>
342	73 , 5308-5319.
343	Seeberg-Elverfeldt J., Schlüter M., Feseker T., Kölling M. (2005) Rhizon sampling of pore
344	waters near the sediment/water interface of aquatic systems. Limnol. Oceanogr.
345	<i>Methods</i> 3 , 361-371.

346	Tremaine D.M., Froelich P.N., Wang Y. (2011) Speleothem calcite farmed in situ: Modern
347	calibration of δ^{18} O and δ^{13} C paleoclimate proxies in a continuously-monitored natural
348	cave system. Geochim. Cosmochim. Acta 75, 4929-4950.
349	Ullmann C.V., Wiechert U., Korte C. (2010) Oxygen isotope fluctuations in a modern North
350	Sea oyster (Crassostrea gigas) compared with annual variations in seawater
351	temperature: Implications for palaeoclimate studies. Chem. Geol. 277, 160-166.
352	Urey H.C. (1947) The thermodynamic properties of isotopic substances. Journal of the
353	Chemical Society (Resumed), 562-581.
354	Versteegh E.A.A., Vonhof H.B., Troelstra S.R., Kaandorp R.J.G., Kroon D. (2010)
355	Seasonally resolved growth of freshwater bivalves determined by oxygen and carbon
356	isotope shell chemistry. GGG 11, 16.
357	Wanamaker A.D., Jr., Kreutz K.J., Borns H.W., Jr., Introne D.S., Feindel S., Barber B.J.
358	(2006) An aquaculture-based method for calibrated bivalve isotope
359	paleothermometry. GGG 7, 13.
360	Wanamaker A.D., Jr., Kreutz K.J., Borns Jr. H.W., Introne D.S., Feindel S., Funder S.,
361	Rawson P.D., Barber B.J. (2007) Experimental determination of salinity, temperature,
362	growth, and metabolic effects on shell isotope chemistry of Mytilus edulis collected
363	from Maine and Greenland. Paleoceanography 22, 12.
364	Zeebe R.E., Wolf-Gladrow D. (2001) CO ₂ in seawater: equilibrium, kinetics, isotopes.
365	Elsevier, Amsterdam.
366	Ziveri P., Stoll H., Probert I., Klaas C., Geisen M., Ganssen G., Young J. (2003) Stable
367	isotope 'vital effects' in coccolith calcite. Earth Planet. Sci. Lett. 210, 137-149.
368	

370 Figure and Table legends

371

Table 1. Experimental conditions. Average soil solution pH, [Ca²⁺], CaCO₃ production and

373 $\delta^{18}O_w$ values are all means \pm SD.

374

Figure 1. SEM pictures of a *L. terrestris*-produced calcite granule. (a) entire granule, (b)

376 surface calcite crystal structure, (c) individual calcite crystal. Images are back-scattered

377 electron images taken using a FEI Quanta FEG 600 Environmental Scanning Electron

378 Microscope. (a) and (b) are taken from Lambkin et al. (2011).

379

380 Figure 2. $\delta^{18}O_c$ values of individual granules and soil solution $\delta^{18}O_w$ values. Granules

381 produced at 10 °C (blue), 16 °C (purple) and 20 °C (red) in Hamble soil (triangles) and Red

Hill soil (circles). Higher $\delta^{18}O_w$ values result in higher $\delta^{18}O_c$ values and higher temperatures

383 give lower $\delta^{18}O_c$ values.

384

Figure 3. Comparison of $\delta^{18}O_{pred}$ values according to Kim and O'Neil (1997) and measured $\delta^{18}O_c$ values for earthworm-produced calcite. *L. terrestris* CaCO₃ granules are enriched in ¹⁸O by 1.51 ‰.

388

389 Figure 4. Relationship between $10^3 \ln \alpha$ and temperature for several types of calcite. L.

390 *terrestris* granules (red diamonds; this study), equilibrium (grey solid line), synthetic at initial

391 $[Ca^{2+}]$ and $[HCO_3^{-}]$ of 15 mM (black), synthetic at initial $[Ca^{2+}]$ and $[HCO_3^{-}]$ of 25 mM (grey

dashed line) (Kim and O'Neil (1997)), and *M. edulis* shells (blue triangles) (Wanamaker et al.

(2007)). Earthworm granules are enriched in ¹⁸O compared to equilibrium, in a similar way as

394 synthetic carbonates precipitated at elevated initial concentrations of Ca^{2+} and HCO_3^{-} .

T (°C)	Soil	Mineral	Water	Soil	Soil solution	CaCO ₃ production	$\delta^{18}O_w$	N*
		water added	holding	solution	[Ca ²⁺] (mM)	per earthworm	(‰ VSMOW)	
			capacity (%)	рН		(mg/day)		
		Fiji		7.8	0.46		-6.3 ±0.2	1
Pure min	eral waters	England		7.4	1.20		-7.3 ±0.2	1
		Norway		7.7	0.18		-10.0 ±0.2	1
10	Hamble	Fiji	33	7.2 ±0.4	10.93 ±0.89	1.45 ±0.28	-5.93 ±0.05	5
10	Hamble	England	33	6.7 ±0.9	10.44 ± 0.62	1.66 ±0.43	-7.11 ±0.04	6
10	Hamble	Norway	33	5.5 ± 1.0	8.30 ± 1.02	1.24 ±0.47	-9.60 ±0.05	6
10	Red Hill	Fiji	56	6.9 ± 0.1	15.86 ± 2.09	1.82 ±0.21	-6.04 ±0.11	6
10	Red Hill	England	56	6.7 ±0.1	13.30 ±0.82	1.77 ±0.26	-7.24 ± 0.03	6
10	Red Hill	Norway	56	6.9 ±0.1	15.21 ±0.92	1.76 ±0.46	-9.85 ±0.06	6
16	Hamble	Fiji	33	7.3 ±0.2	12.13 ±1.52	2.30 ±0.24	-5.72 ±0.14	5
16	Hamble	England	33	7.1 ±0.2	11.22 ±1.31	2.11 ±0.25	-7.10 ± 0.08	6
16	Hamble	Norway	33	6.7 ±0.8	9.09 ±0.87	1.79 ±0.37	-9.44 ±0.10	6
16	Red Hill	Fiji	56	6.9 ±0.1	15.82 ±0.84	2.56 ±0.27	-6.20 ± 0.12	5
16	Red Hill	England	56	6.7 ±0.2	13.44 ± 1.45	2.75 ±0.27	-7.24 ± 0.09	6
16	Red Hill	Norway	56	6.7 ±0.2	15.25 ± 0.49	2.19 ±0.49	-9.94 ±0.10	6
20	Hamble	Fiji	33	7.0 ± 0.4	12.34 ± 1.35	2.39 ±0.47	-5.43 ±0.10	6
20	Hamble	England	33	6.9 ±0.7	11.39 ± 0.58	2.06 ±0.60	-6.74 ±0.04	6
20	Hamble	Norway	33	5.7 ±0.5	11.38 ±0.43	1.92 ±0.47	-9.02 ±0.11	5
20	Red Hill	Fiji	56	6.8 ±0.2	16.79 ± 1.42	2.46 ±0.27	-5.85 ±0.05	6
20	Red Hill	England	56	6.8 ±0.3	13.81 ±0.53	2.89 ±0.31	-7.02 ± 0.05	5
20	Red Hill	Norway	56	6.9 ±0.2	15.10 ± 0.89	2.70 ±0.53	-9.60 ±0.10	4

Table 1. Experimental conditions.

* N is number of replicates per treatment. Average soil solution pH, [Ca²⁺], CaCO₃

production and $\delta^{18}O_w$ values are all means \pm s.d.







