

Sensory and instrumental analysis of medium and long shelf-life Charentais cantaloupe melons (Cucumis melo L.) harvested at different maturities

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1	Sensory and instrumental analysis of medium and long shelf-life Charentais cantaloupe
2	melons (Cucumis melo L.) harvested at different maturities
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26 ABSTRACT

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27	The flavour profiles of two genotypes of Charentais cantaloupe melons (medium shelf-life
28	and long shelf-life), harvested at two distinct maturities (immature and mature fruit), were
29	investigated. Dynamic headspace extraction (DHE), solid-phase extraction (SPE), gas
30	chromatography-mass spectrometry (GC-MS) and gas chromatography-olfactometry/mass
31	spectrometry (GC-O/MS) were used to determine volatile and semi-volatile compounds.
32	Qualitative descriptive analysis (QDA) was used to assess the organoleptic impact of the
33	different melons and the sensory data were correlated with the chemical analysis. There were
34	significant, consistent and substantial differences between the mature and immature fruit for
35	the medium shelf-life genotype, the less mature giving a green, cucumber character and
36	lacking the sweet, fruity character of the mature fruit. However, maturity at harvest had a
37	much smaller impact on the long shelf-life melons and fewer differences were detected.
38	These long shelf-life melons tasted sweet, but lacked fruity flavours, instead exhibiting a
39	musty, earthy character.
40	

41 Keywords: melon, (*Cucumis melo* L.); flavour; Cantaloupe; Charentais; volatile compounds;
42 semi-volatile compounds; sensory evaluation; GC-MS; GC-O/MS

43 **1. Introduction**

44 Fully ripe orange-fleshed Charentais melons (*Cucumis melo* L. var. *cantalupensis*) are highly 45 considered for their unique aromatic flavour as well as for the sweet taste of the flesh, both characteristics which develop as the fruit reaches full maturity. Volatile compounds, mainly 46 47 esters, increase with increasing fruit maturity, thus contributing to the desirable sweet aroma 48 of the fruit. Moreover, fruit that remains attached to the plant accumulates sucrose, resulting 49 in a fruit with a sweet taste. Therefore, to achieve optimum quality and consumer acceptance, 50 melon fruit should be harvested fully mature. Unfortunately, the shelf-life of Charentais 51 melons tends to be very short. In order to deliver a longer shelf-life, fruits are either harvested 52 partially mature, or varieties with extended shelf-life are used. Hybrids of the latter have been 53 produced by plant breeders in order to extend the shelf-life, although consumers often 54 complain about their poor quality, which is associated with less aroma, compared with wildvarieties (Aubert & Bourger, 2004). 55 56 There have been many studies investigating different types of melons, focusing on the effect 57 of harvest maturity on quality characteristics, including colour, firmness, ethylene, total 58 sugars, organic acids, amino acids, volatile compounds and sensory characteristics (Wyllie, 59 Leach, & Wang, 1996; Wang, Wyllie, & Leach, 1996; Beaulieu & Grimm, 2001; Beaulieu, 60 Ingram, Lea, & Bett-Garber, 2004; Beaulieu, 2006; Beaulieu & Lancaster, 2007; Beaulieu & 61 Lea, 2007; Vallone, Sivertsen, Anthon, Barrett, Mitcham, Ebeler et al., 2013), but very few 62 on Charentais melons (El-Assi & Alsmeirat, 2010; Alsmeirat & El-Assi, 2010). Moreover, 63 there are several studies showing how volatile compounds decrease in Véndrantais melons 64 transformed with an aminocyclopropane-1-carboxylic acid (ACC) oxidase antisense gene 65 (Bauchot, Mottram, Dodson, & John, 1998; Bauchot, Mottram, & John, 2000), however, only 66 a few papers focus on the volatile compounds of medium and long shelf-life varieties

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obtained by conventional breeding methods (Aubert & Bourger, 2004; Lamikanra, Juaraez,

68 Watson, & Richard, 2003).

69 The purpose of this study was to investigate the effect of harvest maturity and the effect of

70 two different genotypes of Charentais melons with extended shelf-life, on the flavour profile

71 (volatile, semi-volatile and non-volatile compounds) of the melons. Moreover, quantitative

72 descriptive analysis was also used in order to confirm the organoleptic impact of the chemical

raise changes and to find correlations between sensory and instrumental data.

74 2. Materials and methods

75 2.1 Melons

76 Charentais melons (C. melo L. var. cantalupensis) of two different genotypes (one medium 77 shelf-life coded as MSL (cv. Match) and one long shelf-life coded as LSL (cv. Vulcano)) 78 harvested at two distinct maturities (immature - harvested prior to commercial harvest point 79 - coded as i, and mature - harvested at commercial harvest point - coded as m) were 80 supplied by Syngenta Seeds Ltd. The harvest point was defined according to the senescence 81 of the leaf next to the fruit, also taking into account changes in the external fruit colour plus 82 the senescence of the peduncle (these are non-slip varieties which means that they do not 83 detach from the plant; however, the peduncle does senesce). Melons were stored at 8 °C 84 before analysis, and all analyses were performed within four days of receipt in June 2009

85 (shipping times were the same for all samples and aligned to commercial practices).

86 2.2 Chemicals

For capillary electrophoresis (CE), the basic anion buffer (Part No.: 5064-8209) used for
sugar and organic acid analysis was purchased from Agilent (Santa Clara, CA). Glucose,
fructose, and citric acid were purchased from Sigma-Aldrich Co. Ltd and sucrose and malic
acid from Fluka (Poole, UK). For solid-phase extraction (SPE), HPLC-grade methanol was
purchased from Merck Ltd (Poole, UK) and methyl acetate, sodium sulphate and HPLC

92	grade water from Fisher Scientific (Loughborough, UK). 3-Chlorophenol and the alkane
93	standard C ₇ –C ₃₀ (1000 μ g/ml) in hexane were purchased from Sigma-Aldrich Co. Ltd
94	(Gillingham, UK). For dynamic headspace extraction (DHE), compounds used as standards
95	were obtained from Sigma-Aldrich Co. Ltd: 1,2-dichlorobenzene in methanol (130.6 µg/ml)
96	and the alkane standards $C_6 - C_{25}$ (100 µg/ml) in diethyl ether. The EZ-Faast amino acid
97	analysis kit (Phenomenex, Torrance, CA) was used for the analysis of amino acids by GC-
98	MS. Norvaline was obtained from Sigma-Aldrich Co. Ltd.
99	2.3 Preparation of sample extracts
100	One melon from each point (maturity, genotype) was rinsed in cold running tap water, the
101	skin (0.8 cm) and the seeds were removed and the remaining fruit was chopped and blended
102	in a food processor. Portions of 200 g were weighed into polypropylene centrifuge bottles
103	(250 ml; Nalge Nunc International, Rochester, NY) and the bottles were centrifuged at
104	21,859 g for 20 min at 4 °C in a RC-6C Plus Sorvall R centrifuge (Thermo Scientific,
105	Waltham, MA). For chemical analysis, the supernatant juice was filtered under vacuum using
106	a Whatman filter No.1 (GE Healthcare UK Ltd, Buckinghamshire, UK), in order to remove
107	any tissue particles, and the filtrate was used for all the analyses. Three replicate fruits were
108	prepared for each point. Portions of the 12 melon extracts were used immediately for sensory
109	and volatile analysis, whilst the remainder was stored at -20 °C prior to semi-volatile and
110	non-volatile analyses.
111	2.4 Volatile compounds
112	2.4.1 Dynamic headspace extraction
113	Melon juice (2 ml) obtained as described above, was transferred to a 250-ml conical flask
114	with a screw-thread neck and 10 ml of water were added. The flask was then placed in the
115	water bath at 37 °C, and a flow of nitrogen swept the volatiles for 1 h at 40 ml/min onto a

116 glass-lined, stainless steel trap (105 mm × 3 mm i.d.) containing 85 mg of Tenax TA

- 117 (Scientific Glass Engineering Ltd, Ringwood, Australia). Internal standard (1 µl of 130.6
- $\mu g/ml$ 1,2-dichlorobenzene in methanol) was added to the trap at the end of the collection,
- and excess solvent and any water retained on the trap were removed by purging the trap with
- nitrogen at 100 ml/min for 10 min.
- 121 2.4.2 GC-MS analysis of DHE extracts
- 122 Traps were thermally desorbed in a CHIS injection port (Scientific Glass Engineering Ltd)
- attached to a HP5890/5972 GC-MS (Agilent) as described by Elmore, Parker, Halford,
- 124 Muttucumaru, and Mottram (2008). Volatiles were identified by comparison of each mass
- spectrum with spectra from authentic compounds analysed in our laboratory, or from the
- 126 NIST mass spectral database (NIST/EPA/NIH Mass Spectral database, 2008), or spectra
- 127 published elsewhere. To confirm the identification, the linear retention index (LRI) was
- 128 calculated for each volatile compound using the retention times of a homologous series of C_6
- 129 $-C_{25}$ *n*-alkanes and by comparing the LRI with those of authentic compounds analysed under
- similar conditions. The approximate quantification of volatiles collected from the headspace
- 131 were calculated from GC peak areas, by comparison with the peak area of the 1,2-
- dichlorobenzene standard, using a response factor of 1.
- 133 2.4.3 GC-O/MS analysis of DHE extracts
- 134 After the extraction onto preconditioned glass traps (4 mm i.d., 6 mm o.d., 89 mm long)
- packed with Tenax TA (Supelco, Bellefonte, PA) as described above (but from 20 ml of
- melon juice), the trap was desorbed onto a HP-5MS column (30 m \times 0.25 mm \times 0.25 μ m film
- 137 thickness) in an Agilent 7890A/5975C GC-MS (Agilent, Santa Clara, CA), equipped with an
- automated thermal desorber (Turbomatrix ATD; Perkin Elmer, Waltham, MA) and fitted
- 139 with an ODO 2 GC-O system (Scientific Glass Engineering Ltd). After desorption, the oven
- 140 was maintained at 40 °C for a further 2 min and then the temperature was raised at 4 °C/ min
- to 300 °C. The mass spectrometer was operated in the electron impact mode with a source

- temperature of 230 °C, an ionising voltage of 70 eV, and a scan range from m/z 20 to 400.
- 143 Two assessors were used for the detection and verbal description of the odour-active
- 144 components of extracts and only those odours which were detected by both assessors were
- recorded in the results. The assessors scored each odour on a seven-point line-scale (2-8)
- where 3 = weak, 5 = medium and 7 = strong. *n*-Alkanes C₆-C₂₅ were analysed under the
- same conditions to obtain linear retention index (LRI) values for the components.
- 148 2.5 Semi-volatile compounds
- 149 2.5.1 Solid-phase extraction
- 150 3-Chlorophenol (100 µl of a solution containing 1 mg/ml in 10% methanol/water) was added
- to the filtrate $(20 \pm 0.1 \text{ ml})$ as internal standard and the extraction was performed as described
- 152 by Lignou, Parker, Oruna-Concha and Mottram (2013).
- 153 2.5.2 GC-MS analysis of SPE extracts
- 154 Extracts were analysed by an Agilent 6890/5975 GC-MS as described by Lignou et al.
- 155 (2013). Semi-volatile compounds were identified as described above for the volatile
- 156 compounds. The semi-quantification of semi-volatile compounds was calculated from the GC
- 157 peak areas, by comparing with the peak area of the 3-chlorophenol standard, using a response

158 factor of 1.

- 159 2.5.3 GC-O/MS analysis of SPE extracts
- 160 The extract (1 µL) was injected into the injection port of an Agilent 7890A/5975C Series GC-
- 161 MS system equipped with an ODO 2 GC-O system. The column used was a DB-Wax column
- $162 (30 \text{ m} \times 0.25 \text{ mm} \times 0.25 \text{ } \mu\text{m} \text{ film thickness})$. The temperature programme employed was 1
- 163 min at 40 °C, a ramp of 4 °C/min to 240 °C, and hold for 10 min. The extract was injected in
- splitless mode. The helium carrier gas flow rate was 1 ml/min. The mass spectrometer was
- operated in electron impact mode with a source temperature of 230 °C, an ionising voltage of
- 166 70 eV, and a scan range from m/z 29 to 400. One assessor was used for the detection and

- 167 verbal description of the odour-active components of extracts. Each odour was scored on a
- seven-point line-scale (2–8) where 3 = weak, 5 = medium and 7 = strong. *n*-Alkanes $C_7 C_{30}$
- 169 were analysed under the same conditions to obtain linear retention index (LRI) values for the
- 170 components.
- 171 2.6 Non-volatile compounds
- 172 2.6.1 Sample preparation
- 173 An aliquot (1.5 ml) of melon juice was centrifuged at 7200 g for 15 min and then the
- 174 centrifuged supernatant (400 μl) was transferred to an Amicon Ultra 3,000 MWCO filter
- unit (Millipore, Carrigtwohill, Co. Cork, Ireland) and centrifuged at 7200 g for 30 min.
- 176 2.6.2 Determination of free amino acids by GC-MS
- 177 An aliquot of the centrifuged supernatant (100 µl) was derivatised using the EZ-Faast amino
- acid derivatisation technique (Phenomenex). GC-MS analysis of the derivatised samples was
- 179 carried out using an Agilent 6890/5975 GC-MS instrument, as described by Elmore,
- 180 Koutsidis, Dodson, Mottram, and Wedzicha (2005).
- 181 2.6.3 Determination of organic acids and carbohydrates by capillary electrophoresis (CE)
- 182 An aliquot of the centrifuged supernatant (100 μ l) was analysed as described by Lignou et al.

183 (2013).

184 2.7 Sensory Analysis

The permanent in-house panel of 13 experienced assessors was used to develop a sensory profile to describe the sensory characteristics of the melon juice and the characteristics were estimated quantitatively. Aliquots (20 ml) of melon juice (prepared as described above and filtered through a tea strainer to remove particulate matter) were presented to each assessor at room temperature in clear polypropylene tasting cups. During the development of the sensory profile, the assessors were asked to sniff and then taste (and swallow) the samples to produce as many descriptive terms as seemed appropriate. Reference materials (including a number of

192 fruit and vegetables, such as strawberries, pineapple, aged apple and banana, citrus, plum, 193 kiwi, butternut squash, different types of melon (honeydew and Galia), stored cantaloupe 194 melon, pips and centre from cantaloupe melon, cucumber and other materials like sugar 195 syrup) were used in order to help the assessors to standardise the language development 196 process. These terms were discussed by the assessors, as a group, with the help of the panel 197 leader, and this led to an agreed profile comprising 13 odour terms, 19 taste/flavour terms, 6 198 mouthfeel terms, and 10 after-effects terms. The quantitative sensory assessment took place 199 in the sensory booths, each equipped with computer screen and a mouse. Compusense 200 version 5 software (Compusense Inc., Guelph, Ontario, Canada) was used to acquire the 201 sensory data. A warm-up sample (a mixture of the examined samples) was presented first to 202 eliminate first position bias and then the samples were presented to the assessors in a 203 balanced randomised order. The assessors were instructed to sniff the samples to score the 204 aroma attributes, and then taste (and swallow) the samples to score the overall taste/flavour 205 attributes and the mouthfeel attributes. There was a 45-s pause after the end of the mouthfeel 206 attributes and the assessors then scored the after-effects which included both taste and 207 mouthfeel effects. The intensity of each attribute for each sample was recorded by the 208 assessors on a 100-point unstructured line scale. The same three replicates used for chemical 209 analysis were also used for sensory analysis. Between samples, panellists cleansed their 210 palate with yoghurt, cracker and water.

211 2.8 Statistical analysis

The quantitative data for each compound identified in the GC-MS analyses (volatile, semivolatile and non-volatile compounds) were analysed by both one- and two-way analysis of variance (ANOVA) and principal component analysis (PCA) using XLSTAT Version 2012.1.01 (Addinsoft, Paris, France). For those compounds exhibiting significant difference in the one-way ANOVA, Fisher's least significant difference (LSD) test was applied to

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determine which sample means differed significantly (p < 0.05). These data are shown in

Table 1. SENPAQ version 3.2 (Qi Statistics, Reading, UK) was used to carry out ANOVA

and PCA of sensory panel data. The means for the sensory data were taken over assessors and

- correlated with the means from instrumental data *via* multiple factor analysis (MFA) using
- 221 XLSTAT.

222 **3. Results**

223 3.1 Volatile compounds

More than 70 compounds were identified in the headspace of the two genotypes. The most 224 225 abundant compounds are listed in Table 1. These included 31 esters (acetates and non-acetate 226 esters), 8 sulfur-containing compounds, 10 alcohols, 8 aldehydes, 2 terpene derivatives and 2 227 other compounds. Quantitative differences were observed between the two maturity stages 228 (immature (i) and mature (m) fruit) and the two genotypes (medium shelf-life (MSL) and 229 long shelf-life (LSL)). Esters (acetates and non-acetate esters) comprised more than 87% of 230 the total volatiles collected from the iMSL fruit, a percentage which increased to more than 231 93 % in the mMSL fruit. Similarly, the percentage of esters increased from 69% in the iLSL 232 fruit to more than 77% in the mature fruit of the same genotype. The most abundant esters 233 identified were ethyl acetate, 2-methylpropyl acetate, butyl acetate, 2-methylbutyl acetate and 234 ethyl butanoate. Wyllie et al. (1996) and Bauchot et al. (2000) reported that these compounds 235 were predominant in Makdimon (C. melo var. reticulatus) and Vedrantais (C. melo var. 236 *cantalupensis*) cultivars respectively. These compounds were also the most abundant in a 237 number of Charentais cantaloupe cultivars (Aubert & Bourger, 2004) and in Jiashi 238 muskmelon (var. reticulatus, Hami melon) (Pang, Guo, Qin, Yao, Hu, & Wu, 2012). 239 Both immature fruits contained very few esters compared to their respective mature fruit. Ten 240 out of 13 acetates and 12 out of 18 non-acetate esters were found significantly higher in the 241 mMSL fruit compared to the iMSL fruit. The same trend was observed for the LSL fruits, but

242	the levels were much lower and the differences were not significant. However, the levels of
243	ethyl esters and particularly ethyl acetate, ethyl propanoate, ethyl 2-methylpropanoate, ethyl
244	butanoate and ethyl 2-methylbutanoate increased 4-fold for LSL and 26-fold for MSL with
245	increasing maturity.
246	Generally, the levels of esters were remarkably lower in the LSL genotype, even in mLSL.
247	Similar results were reported by Lamikanra et al. (2003), where hybrids with long shelf-life
248	and hybrids with extended shelf-life presented significantly lower contents of total volatile
249	aromas than traditional shelf-life C. melo var. reticulatus cv. Mission melons. Aubert and
250	Bourger (2004), who studied the volatile compounds of 15 Charentais melon cultivars,
251	reported the same trends: a reduction in a range of 43-77% of total esters in LSL melons
252	compared to MSL or wild melons. They reported that these differences were more obvious
253	for compounds with low odour threshold values, such as ethyl 2-methylbutanoate (0.006
254	μ g/kg), ethyl butanoate (1 μ g/kg), ethyl hexanoate (1 μ g/kg), butyl acetate (2 μ g/kg) and
255	hexyl acetate (2 μ g/kg). Bauchot et al. (1998) also noted that in transformed Charentais
256	melons with an ACC oxidase antisense gene, the total volatiles were 60-85% lower than that
257	of the nontransformed hybrids. They observed that the reduction in volatiles in these melons
258	was greater for ethyl esters than for acetates, and since ethyl esters have lower odour
259	threshold values than acetates, the reduction of ethylene production in these melons, had the
260	greatest effect on the most potent odorants (Bauchot et al., 2000).
261	Eight sulfur-containing compounds were identified in the headspace of the samples including
262	six thioether esters. Wyllie and Leach (1992) reported that 2-(methylthio)ethyl acetate and 3-
263	(methylthio)propyl acetate were the dominant sulfur compounds in all melon cultivars
264	studied, as was the case in the Charentais melon under study, but only in mMSL fruit. Ethyl
265	2-(methylthio)acetate was another important compound and again present only in mMSL
266	fruit. Generally, the sulfur-containing esters were not detected in the LSL fruit and only two

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267	were detected in the iMSL fruit. These compounds are very important in the overall aroma
268	profile of melons, because many are potent odorants with low odour thresholds. A few
269	authors have reported that trace amounts of these compounds have a major impact on the
270	musky note of some melon aromas (Wyllie et al., 1992; Wyllie & Leach, 1990; Wyllie,
271	Leach, Wang, & Shewfelt, 1994; Jordan, Shaw, & Goodner, 2001; Hayata, Sakamoto,
272	Kozuka, Sakamoto, & Osajima, 2002; Hayata, Sakamoto, Maneerat, Li, Kozuka, &
273	Sakamoto, 2003). Aubert and Bourger (2004) also reported a considerable reduction in the
274	levels of these compounds in LSL cultivars, whereas the total levels of them in wild or MSL
275	cultivars were up to 17 times higher than in LSL cultivars.
276	Besides esters and sulfur-containing compounds, some alcohols and aldehydes were
277	identified in the samples. The levels of most alcohols increased with increasing maturity for
278	both genotypes, and this increase was significantly higher, particularly for mMSL fruit.
279	Regarding the aldehydes found, no significantly changes were observed between the different
280	samples except for 2-methyl-2-butenal and 6-nonenal. 2-Methyl-2-butenal was significantly
281	higher in mMSL fruit and 6-nonenal was significantly higher in iMSL fruit. Terpenes like
282	limonene, eucalyptol and geranylacetone were also found, however, only eucalyptol was
283	found significantly higher in mMSL fruit. Finally, 2-methylbutanenitrile and 3-
284	methylbutanentrile were reported for the first time in melons. These compounds were found
285	to be significantly higher in mMSL fruit.
286	To sum up, among all the volatiles identified, 30 compounds were significantly affected by
287	the maturity and 34 by the genotype, supporting the hypothesis that both factors were very
288	important. The two-way ANOVA showed a clear trend, with many of the compounds (mainly
289	esters, sulphur-containing compounds and several alcohols) showing a significant interaction

290 between the two variables. The combination of an MSL variety, and a fruit harvested at

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291	maturity, produced a far greater increase in these compounds than would have been predicted
292	from a simple additive model. This synergy is reflected in the GC-O data.
293	GC-olfactometry analysis of the samples yielded a total of 18 odorants in the chromatogram,
294	which are presented in Table 2. All but one of these compounds were identified in the GC-
295	MS analysis, the exception being 4-heptenal which was recognised by its characteristic aroma
296	and confirmed by comparison of its LRI with that of the authentic sample. Quantitative
297	differences were observed between the two maturity stages and the two genotypes. It is
298	clearly illustrated in Table 2 that esters were the most important contributors to the desirable
299	sweet and fruity aroma of the fruit. In particular, seven esters, including ethyl propanoate,
300	propyl acetate, ethyl 2-methylpropanoate, methyl 2-methylbutanoate, ethyl butanoate, ethyl
301	2-methylbutanoate and butyl propanoate, contributed to the fruity, pineapple-like and sweet
302	aroma, particularly of mMSL. Four of these esters were only detected in mMSL, and the
303	other three branched esters were also detected in the less mature and the LSL fruits, but
304	tended to have higher scores for mMSL.
305	Schieberle, Ofner, and Grosch (1990) studied the potent odorants in muskmelons by aroma
306	extraction dilution analysis (AEDA), and they reported that indeed the volatile esters were
307	responsible for the fruity notes in the aroma of muskmelon and that methyl 2-
308	methylbutanoate and ethyl 2-methylbutanoate were the most intense odorants in the ester
309	fraction. Jordan et al. (2001) also found that these two esters contributed to a fruity, sweet
310	and cantaloupe-like aroma. Pang et al. (2012) studied the odour-active compounds of Jiashi
311	muskmelon using both detection frequency analysis (DFA) and odour activity values (OAV).
312	They reported that ethyl 2-methylpropanoate, ethyl butanoate and ethyl 2-methylbutanoate
313	were the esters with the greatest relative importance and were characterised as having fruity,
314	sweet and cantaloupe-like odours. Hexanal, which imparts a fresh green note (Schieberle et
315	al., 1990), and (Z)-3-hexen-1-ol, which imparts a herbal green note (Jordan et al., 2001), were

316 detected in these samples and described as having green and grass notes, respectively. 317 Eucalyptol, reported by Schieberle et al. (1990), was another important odorant detected only 318 in mMSL samples. Kemp, Knavel, and Stoltz (1972), and Kemp, Knavel, Stoltz, and Lundin 319 (1974) concluded that (Z)-6-nonenal and 3,6-nonadien-1-ol were two potent odorants 320 contributing to muskmelon flavour. These two compounds were also identified in these 321 samples, having a cucumber and green note, respectively. 6-Nonenal was scored consistently 322 higher in the immature fruits, consistent with the greener notes of under-ripe fruit. These 323 compounds were also reported by Pang et al. (2012) in Jiashi muskmelons and along with 324 2,6-nonadienal and 2-nonenal were the important contributors for green and cucumber-like 325 aromas. Pang et al. (2012) also stated that although esters were superior in concentration 326 (86%), their contribution rate (OAV percentages) to the aroma profile of Jiashi muskmelons 327 was only 10%, whereas alcohols and aldehydes were just the opposite. The contents of 328 aldehydes and alcohols were only 11 and 4 % that of esters, respectively, but their 329 contribution rates were 56 % and 34 % respectively. 330 Finally, of the eight sulfur compounds which were identified in the headspace of the melons, 331 four were detected by the assessors. S-Methyl 2-methylbutanethioate had a sulfury odour, 332 whereas dimethyl trisulfide imparted a pickled onions and cabbage odour. Ethyl 2-333 (methylthio)acetate and ethyl 3-(methylthio)propanoate were only identified in mMSL and 334 had an earthy but slightly cucumber note and a cardboard but slightly green odour, 335 respectively. Overall, comparing the odours between the two maturity stages and the two 336 genotypes, it can be observed that mMSL fruit presented the highest intensities, which 337 resulted in a more aromatic fruit compared to the others. 338 3.2 Semi-volatile compounds

More than 40 compounds were identified in melon SPE extracts and 29 of them were

quantified and listed in Table 1. Semi-volatile compounds included 9 esters (acetates and

diacetates), 5 sulfur-containing compounds and a few other compounds (alcohols, aldehydes,

342 furans, acids).

343 2,3-Butanediol diacetate and its precursor 2,3-butanediol monoacetate were identified and found to be significantly higher in mMSL genotype. These compounds were also identified in 344 345 Japanese melon (cv. Golden Crispy) (Wyllie et al., 1990). 2,3-Butanediol diacetate possesses 346 two asymmetric carbons (erythro and threo forms and a meso-form diastereoisomer), thus 347 producing two peaks on GC (Aubert & Pitrat, 2006). According to Wyllie et al. (1990), the 348 most abundant peak would be the D and/or L isomer, whereas the other would be the meso 349 isomer. 1,2-Propanediol and 1,2-ethanediol diacetate were also identified and found to be 350 significantly higher in mMSL genotype. 351 Five sulfur-containing compounds were identified with this method, three of which had been 352 previously found in the headspace of these melons. The additional compounds were 2-353 (methylthio)-1-ethanol and 3-(methylthio)-1-propanol and these were, again, significantly 354 higher in mMSL genotype. The relative quantities of these compounds showed good 355 agreement between the two analytical methods. 356 Other compounds identified were alcohols, including 1-hexanol, 3-hexen-1-ol, benzyl alcohol 357 and phenylethanol, compounds that increased with increasing maturity. 5,6,7,7a-Tetrahydro-358 4,4,7a-trimethyl-2[4H]-benzofuranone (dihydroactinidiolide) is potentially an important 359 compound since it imparts a fruity musky note and was found in higher concentrations in the 360 mature fruits. 2-Ethyl-4-hydroxy-5-methyl-3[2H]-furanone (homofuraneol) and 4-hydroxy-361 5-methyl-3[2H]-furanone (norfuraneol) were also identified in larger amounts in mature fruits 362 of both genotypes. Finally hexadecanoic acid and 9-hexadecenoic acid were present in the 363 extracts and increased as well with increasing maturity. 364 To sum up, among all the semi-volatiles identified, 17 compounds were significantly affected 365 by maturity and only 11 by genotype, suggesting that the maturity factor was more important

15

366 for this set of results. There was, again, a clear trend defined by two-way ANOVA where the 367 majority of esters and sulfur-containing compounds showed a strong interaction between the 368 variables, and the synergy between the maturity at harvest and genotype was evident. GC-olfactometry analysis of the SPE extracts yielded a total of 20 aromatic regions in the 369 370 chromatogram, which were described with a range of terms, including cabbage, cheesy, 371 vinegar, Brie, mushroom, soil, bread, onions, balsamic, cucumber, green, vegetable, cooked 372 potato, floral, synthetic, rubbery, woody, smoky, strawberry, caramel, candyfloss, and rose 373 petals. A number of these odours were detected in our previous study (Lignou et al., 2013); 374 however, the identities of many of these compounds remain unknown. A number of 375 compounds were positively identified including 3-hexen-1-ol with a very strong cut grass 376 odour in mMSL genotype. 2,3-Butanediol diacetate had an earthy, soily odour, and was also 377 described by Wyllie, Leach, Wang and Shewfelt (1995) as having an earthy note. Among the 378 sulfur compounds, ethyl 2-(methylthio)acetate had a slight green odour, 3-(methylthio)propyl 379 acetate had a mushroom-like odour and 3-(methylthio)-1-propanol an onion-like odour, 380 respectively. Homofuraneol and norfuraneol were responsible for the strawberry sweet, 381 caramel-like note in the aroma. 382 Principal component analysis was used to visualise graphically the differences in volatile and 383 semi-volatile concentrations in the two maturity stages and the two genotypes. Twelve 384 samples were used (2 maturity stages \times 2 genotypes \times 3 replicates) and 87 variables (61 385 volatile compounds and 26 semi-volatile compounds). The first two principal components 386 accounted for 76% of the variation in the data (Figure 1). The first axis mainly discriminated 387 the mMSL fruit from the iMSL and the LSL genotype, whereas the second axis mainly 388 discriminated the iMSL from the LSL genotype. For the LSL genotype, the immature and the 389 mature fruits were not well separated on PC1 or PC2, and the effect of maturity at harvest for 390 the LSL fruits was shown to be small compared to that for the MSL fruits. The distribution of

391	the variables is shown in Figure 1B. The majority of acetates (a02, a04-a13), non-acetate
392	esters (b03, b05, b07, b08, b11-b14, b16, b18), diacetates (g02-g05, g08, g09), sulfur-
393	containing compounds (c02, c05-c08 and h01-h05), several alcohols (d02-d05, d07, i01, i02,
394	i07) and a few other compounds were positively correlated with the first axis. Methyl esters,
395	including methyl acetate (a01), methyl propanoate (b01), methyl 2-methylpropanoate (b02),
396	methyl butanoate (b04), methyl 2-methylbutanoate (b06), methyl pentanoate (b09) and
397	methyl hexanote (b17), as well as S-methyl 2-methylbutanethioate (c03), 6-nonenal (e06) and
398	2,6-nonadienal (i03), were positively correlated with the second axis.
399	Mature MSL fruit, positively correlated with the first axis, was characterised by greater
400	numbers of esters (including acetates, diacetates and non-acetate esters), sulfur-containing
401	compounds, several alcohols and furans. Immature MSL, positively correlated with the
402	second axis, was characterised by greater levels of methyl esters, 6-nonenal and 2,6-
403	nonadienal. Immature LSL and mLSL fruit were negatively correlated with both first and
404	second axis because the concentrations of esters (acetates, diacetates and non-acetate esters)
405	were low and, moreover, sulfur-containing esters were not detected.
406	3.3 Non-volatile compounds
407	Two organic acids were identified: citric and malic acid (Table 1). Citric was the dominant
408	acid in both maturity stages and genotypes. The levels of malic acid were approximately
409	eight times lower than citric acid. The same acids were the dominant acids in cantaloupe
410	melon (cv. Mission) (Lamikanra, Chen, Banks & Hunter, 2000). Wang et al. (1996) found
411	that citric acid increased slightly with increasing maturity in the melon of cv. Makdimon.

412 This was also observed in our results; however, the increase of citric acid was not significant

413 for either genotype (Table 1).

414 The sugars identified in the samples were glucose, fructose and sucrose. The results agree

415 with those stated by Wang et al. (1996), Lester and Dunlap (1985), and Beaulieu, Lea,

Eggleston and Peralta-Inga (2003). As shown in Table 1, glucose and fructose decreased with 416 417 increasing maturity, whereas sucrose increased significantly for both genotypes. Comparing 418 the two genotypes, it can be seen that sucrose was significantly higher in LSL genotype. This 419 probably happened because LSL fruit do not develop an abscission zone, and as a result the 420 fruit may be harvested later, thus allowing for a longer period of sugar accumulation and 421 higher sugar content (the major component of soluble solids in melon). 422 The dominant amino acids in both varieties (Table 1) were glutamine and aspartic acid; 423 however, quantitative differences existed for a number of other amino acids between the 424 maturity stages and genotypes. Almost all amino acids markedly increased with increasing 425 maturity, except glutamine which decreased in the mMSL fruit, and leucine and isoleucine, 426 which did not change significantly. Also alanine was found significantly higher in the mMSL 427 fruit, whereas γ -ABA was one of the dominant amino acids in the LSL genotype. 428 It is well-known that there is a biogenetic relationship between the formation of certain aroma 429 volatiles and levels of free amino acids (Wang et al., 1996). In particular, the amino acids 430 alanine, valine, leucine, isoleucine and methionine are precursors of the majority of the esters 431 found in melons (Wyllie et al., 1995; Wang et al., 1996; Bauchot et al., 1998). The trends 432 observed in this study (increasing free amino acids during development and ripening, leucine 433 and isoleucine remaining constant and glutamine decreasing) were also observed by Wang et al. (1996), who suggested that the type and extent of ester formation may be determined by 434 435 substrate availability in the fruit. In mature melons, the total volatiles content is high, so 436 considerable quantities of precursors are required for their formation. Although the 437 concentrations of leucine and isoleucine remained constant during maturation, esters having 438 carbon skeletons derived from isoleucine did increase with maturity. Wang et al. (1996) 439 suggested that there is a series of steps in ester formation where a considerable degree of 440 selectivity (enzymes involved) must happen as the substrates are drawn from the amino acid

18

441 pool. Thus, the differences between cultivars in esters derived from amino acids are likely to 442 be due to the efficiencies of the different enzyme pathways within each melon. 443 Consequently, it can be concluded that the extent of ester formation will depend on the amount of available substrates. Harvest time will influence the total volatile production, since 444 445 fruit that was harvested prematurely would not accumulate sufficient concentrations of 446 required volatiles substrates and this will lead to a poor flavour profile of that fruit. However, 447 in addition to the availability of different substrates, subcellular localisation should be taken 448 into account as well as the expression of synthesising enzymes, which play an important role 449 in the reactions. Finally, the response to the climacteric genotypes (climacteric or non-450 climacteric) is also an important factor, since it was observed that the expression levels of 451 genes responsible for biosynthesis of melon aroma volatiles are generally higher in 452 climacteric genotypes as compared with non-climacteric genotypes (Gonda et al., 2010). 453 3.4 Sensory analysis The sensory profile of the samples was generated by a trained panel of experts who, at the 454 end of the profile development, agreed to use 49 terms for the quantitative assessment of the 455 456 samples. Table 3 gives the mean panel scores for these attributes and significant differences 457 for the samples, the assessors and their interactions as determined by ANOVA. This table 458 shows 30 out of 49 attributes were found to be significantly different (3 nearly significantly 459 different) between the four samples. A highly significant effect of assessor for all attributes 460 was also found. This suggested that the assessors were using the scales differently; however, 461 only a few attributes (mainly after-effects attributes) had a significant assessor \times sample 462 interaction, thus indicating that the assessors were ranking the samples in a similar way. 463 As shown in Table 3, sweet aroma, floral aroma and honey aroma were found to be 464 significantly higher in mMSL, hence confirming the GC-MS results, where the levels of 465 esters (acetates and non-acetate esters) were higher in these samples. These attributes were

466 highly positively correlated with the sum of acetate and non-acetate esters, having correlation 467 coefficients of more than 0.8 (data not shown). Brown orchard fruit aroma was also 468 significantly higher in mMSL fruit. On the contrary, green and cucumber odour and 469 taste/flavour attributes were scored significantly higher in iMSL fruit followed by iLSL fruit 470 This is also confirmed by both the GC-O and the GC-MS results which showed 6-nonenal 471 (cucumber) was significantly higher in the immature fruit of both genotypes. Sweet and 472 syrupy taste/flavour, as well as sweet aftertaste, were significantly higher in both maturity 473 stages of LSL genotype and in mMSL fruit. This also agrees with the results for sucrose 474 (Table 1). 475 Principal component analysis was carried out on the correlation matrix of all samples and all 476 attributes (Figure 2). The difference in maturity stage was the predominant distinguishing 477 factor in the sensory analysis, with principal component 1 separating the immature from 478 mature MSL fruit and principal component 2 separating the immature from the mature LSL 479 and MSL fruits. Desirable sweet (001), floral (002), honey (003), strawberries (004) and ripe 480 tropical fruit (o12) odour attributes, as well as floral (tf06), honey (tf07), strawberries (tf09) and ripe tropical fruit (tf19) taste/flavour attributes were associated with the mMSL fruit. On 481 482 the other hand, cucumber odour (007), cucumber taste/flavour (tf12), green odour (008), 483 green taste/flavour (tf13), acidic taste (tf04) and aftertaste (ae04), and savoury taste/flavour (tf02) were highly correlated with the iMSL fruit. Regarding the LSL genotype, earthy (009-484 485 tf16) and musty (010-tf17) odour and taste/flavour, and salty (tf03) taste/flavour attributes 486 were associated with the iLSL fruit, whereas taste/flavour attributes like sweet (tf01), syrupy 487 (tf08), brown orchard fruit (tf18), as well as sweet (ae01) aftertaste, were associated with the 488 mLSL fruit. Similar results were reported by Beaulieu et al. (2004) who studied the effect of 489 harvest maturity on the sensory characteristics of fresh-cut cantaloupe. They found that the 490 maturity level at harvest coincided with significant differences in flavour attributes. Sweet

- 491 aromatic flavour and taste significantly increased with increasing maturity, whereas cucurbit
- 492 flavour decreased.
- 493 3.5 Multiple factor analysis (MFA)
- 494 MFA was used in order to simultaneously analyse several tables of variables (three tables for
- 495 instrumental data: volatiles, semi-volatiles and non-volatiles and one table for sensory data),
- thus facilitating a study of the relationship between the observations (different samples), the
- 497 variables and the tables. This was achieved by successively examining the PCA for each
- table, and then the value of the first eigenvalue of each analysis was used to weight the
- 499 various tables in a further PCA. Finally, a weighted PCA on the columns of all the tables was
- 500 performed (Pages, 2004). The coordinates of the tables were displayed and used to create the
- 501 map of the tables (Figure 3A). As it can be seen on the map, the first factor was related with
- the tables of volatiles, semi-volatiles and sensory attributes, whereas the second factor was
- 503 mostly related with the non-volatiles but also with sensory tables.
- 504 The correlation maps of observations and variables are shown in Figure 3B and C
- respectively. Although the plots do not implicitly detail coefficients of correlation, one can
- ascribe relative relationships between parameters closely related, and inversely related
- 507 (separation close to 180°). Observing the variables map it can be concluded that the sensory
- analysis linked well with the instrumental data.
- 509 Mature MSL fruit was positively correlated with the first factor, in other words with sweet
- 510 (o01), honey (o02), floral (o03) and strawberry (o04) odours and floral (tf06), honey (tf07),
- 511 strawberries (tf09) and ripe tropical fruit (tf19) taste/flavour terms. These variables were then
- 512 highly positively correlated with the majority of the esters, which are associated with
- desirable flavour. On the opposite side (negatively correlated with factor one and factor two),
- iMSL fruit was correlated with all the cucumber and green notes (007, 008, tf12, tf13), as
- 515 well as with acidic after-taste (ae04).

21

516 Compounds like 6-nonenal (e06) and two methyl esters (a01 and b01) were positively 517 correlated with iMSL. It is interesting that 2,6-nonadienal (i03) was positively correlated with 518 citrus taste/flavour (tf11). Additionally, the fact that this fruit was negatively correlated with 519 sweet taste/flavour and after-effects terms, gave a fruit with an undesirable odour and taste. 520 This can be drawn from the variables map, where all the esters are negatively correlated with 521 iMSL fruit. Regarding the iLSL fruit (positively correlated with factor two), although it 522 exhibited very low levels of esters compared to iMSL, the high concentration of sucrose and 523 several amino acids contributing to taste (glutamic acid (116) and aspartic acid (112)), gave a 524 fruit with an acceptable taste but lacking in desirable aroma. This was emphasised by high 525 scores for earthy and musty odour, taste/flavour and after-effects (009, 010, tf16, tf17, ae08). 526 Finally, mLSL was correlated with sweet (tf01) and syrupy (tf08) taste/flavour and sweet 527 (ae01) after-effects terms. These terms were associated with sucrose (k03) and, indeed, this 528 mLSL fruit contained the greatest quantity of sucrose. The slightly increased levels of esters 529 (compared to iLSL and iMSL) gave a fruit a quite nice odour with a very sweet taste. 530 4. Conclusions

531 Both sensory and instrumental analysis of volatile, semi-volatile and non-volatile compounds 532 have identified significant differences between four melon samples that can be attributed to 533 either the maturity stage or the genotype. The mature fruit of MSL exhibited the highest 534 amount of esters (acetates, diacetates and non-acetate esters), and those melons were 535 generally described by the assessors as having desirable fruity and sweet odours. Moreover, 536 the combination of quite high sucrose levels, along with other compounds, like homofuraneol 537 and norfuraneol, resulted in a fruit with a very sweet taste, while exhibiting the highest levels 538 of strawberry taste/flavour and the lowest levels of bitter and acidic taste. The immature fruit 539 of the MSL exhibited green, cucumber notes typical of an under-ripe melon and lacked the 540 fruity flavour of the mature MSL. Both LSL melons, harvested immature and mature, were

22

- relatively sweet, with a sweet syrupy flavour but lacking in the fruity character of the mature
- 542 MSL, exhibiting instead an earthy, musty quality. Overall, the mature MSL fruit was full of
- flavour confirming the hypothesis that fruit from MSL genotypes harvested mature will
- develop a strong aromatic flavour, whereas fruit either harvested too early or from LSL
- 545 genotypes will develop a less aromatic flavour.

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Figure Captions

Figure 1. Principal component analysis of four different samples showing correlation with volatile and semi-volatile compounds. (A) Projection of the samples (MSL = medium shelf-life, LSL = long shelf-life, m = mature, i = immature); (B) Distribution of variables (codes on plot refer to compound codes in Table 1).

Figure 2. Principal component analysis of four different samples (\blacktriangle) (MSL = medium shelf-life, LSL = long shelf-life, m = mature, i = immature) showing correlations with sensory attributes (O) (codes on plot refer to sensory attribute codes in Table 3).

Figure 3. MFA: (A) Representation of groups (tables) of variables; (B) Representation of the samples (MSL = medium shelf-life, LSL = long shelf-life, m = mature, i = immature); (C) Distribution of variables (O = volatiles, \bullet = semi-volatiles, \blacksquare = non-volatiles and Δ = sensory variables - codes on plot refer to codes in Tables 1 and 3).

Figure Captions

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Code	Compound	LRI ^a	ID^b	A	oproxima	LSD^d	\mathbf{P}^{e}		
				iLSL	mLSL	iMSL	mMSL		
Volati	le analysis								
Acetat	es								
a01	methyl acetate	<600	А	68 ^a	53 ^a	193 ^b	37 ^a	65	**
a02	ethyl acetate	616	А	118 ^a	458 ^a	196 ^a	3314 ^b	512	***
a03	1-methylethyl acetate	656	А	29	36	44	32	29	ns
a04	propyl acetate	715	А	16 ^a	99 ^a	49 ^a	497 ^b	154	***
a05	2-methylpropyl acetate	773	А	134 ^a	412 ^a	214 ^a	1469 ^b	736	*
a06	butyl acetate	817	А	18 ^a	186 ^a	92 ^a	1538 ^b	690	**
a07	3-methylbutyl acetate	878	А	0.6 ^a	2.7 ^a	1.7 ^a	24 ^b	5.4	***
a08	2-methylbutyl acetate	880	А	16 ^a	61 ^a	102 ^a	1227 ^b	685	**
a09	pentyl acetate	915	А	nd	3.6 ^a	3.4 ^a	105 ^b	59	**
a10	3-hexen-1-yl acetate	1005	А	34 ^a	13 ^a	46 ^a	577 ^b	380	*
a11	hexyl acetate	1013	А	6.4 ^a	36 ^a	26 ^a	598 ^b	262	**
a12	heptyl acetate	1111	А	nd	nd	nd	7.0		
a13	benzyl acetate	1168	А	1.3 ^b	2.9 ^b	nd	35 ^a	28	ns ^(0.060)
	total acetates			441	1361	967	9460		
Non-a	rotato ostors								
h01	methyl propanoate	632	Δ	19 ^a	16 ^a	122 ^b	38 ^a	39	***
b02	methyl 2-methylpropanoate	685	A	9.6ª	12 ^a	44 ^b	29 ^{ab}	25	*
b02	ethyl propanoate	710	Δ	4.2^{a}	24^{a}	11 ^a	559 ^b	211	***
b04	methyl butanoate	722	A	9.0^{a}	2. 8.0 ^a	141 ^b	159 ^b	83	**
b05	ethyl 2-methylpropanoate	758	A	nd	3.9 ^a	1.5 ^a	155 ^b	60	***
b06	methyl 2-methylbutanoate	782	A	21 ^a	17 ^a	98 ^b	131 ^b	54	**
b07	ethyl butanoate	803	A	1.5^{a}	15 ^a	9 9 ^a	1348 ^b	590	**
b08	propyl propanoate	814	A	nd	3.0^{a}	nd	13 10 18 ^b	11	*
b09	methyl pentanoate	830	A	nd	nd	13	0.9	0.8	#
b10	isopropyl butanoate	844	A	0.4^{a}	1.8 ^b	0.8^{a}	1.9 ^b	0.8	**
b10	ethyl 2-methylbutanoate	851	A	1.5 ^a	7.6^{a}	8.7 ^a	422 ^b	189	**
b12	propyl butanoate	901	Δ	nd	,.o	nd	30	105	
b13	ethyl pentanoate	903	A	nd	nd	nd	16		
b14	butyl pronanoate	910	Δ	nd	0 Q ^a	0.7^{a}	4 0 ^b	22	*
b15	methyl hexanoate	926	A	nd	nd	43	7.0 7.0	5.0	#
b16	propyl 2-methylbutanoate	947	Δ	nd	0.1^{a}	4.5 0.1 ^a	2 3 ^b	1.7	*
h17	2-methylpropyl butanoate	956	А А	nd	3 0 ^{ab}	0.1^{a}	2.5 4 5 ^b	35	ns ^(0.055)
b18	ethyl hexanoate	999	Δ	nd	o.u nd	nd	т .5 110	5.5	115
010		177	Л	nu	110		2027		
	total non-acetate esters			66	112	444	3037		
Sulfur-	containing compounds								
c01	S-methyl thioacetate	703	А	nd	nd	2.2	3.1	2.8	#
c02	dimethyl disulfide	748	А	3.4 ^a	7.8 ^a	2.0^{a}	14 ^b	6.0	**
c03	S-methyl 2-methylbutanethioate	944	А	nd	nd	9.8	7.9	5.6	#
c04	dimethyl trisulfide	981	А	0.3	0.7	nd	0.5	0.5	#
c05	ethyl (methylthio)acetate	989	А	nd	nd	nd	52		
c06	2-(methylthio)ethyl acetate	1010	A	nd	nd	nd	69		

Table 1. Approximate quantities of volatile, semi-volatile and non-volatile compounds identified in the headspace, SPE extracts or melon juice respectively of two genotypes of Charentais melon harvested at two different maturity stages.

Code	Compound	LRI ^a	ID^b	Ap	proxima	te quan	tity ^c	LSD^{d}	P ^e
			•	iLSL	mLSL	iMSL	mMSL		
c07	ethyl 3-(methylthio)propanoate	1104	А	nd	nd	nd	8.0		
c08	3-(methylthio)propyl acetate	1127	А	nd	nd	nd	38		
	total sulfur-containing compounds			3.4	8.5	14	193		
Alcoho				011	0.0		1,0		
d01	2 methylpropanol	633	٨	18 ^a	63 ^b	7 0 ^a	34 ^{ab}	35	*
d02	1-butanol	668	Δ	21^{a}	11 ^a	7.0 4 1 ^a	33 ^b	96	***
d03	2-methyl-1-butanol	749	A	2.1 36 ^a	125 ^b	-7.1 28 ^a	295°	71	***
d04	3-hexen-1-ol	866	A	5 5 ^a	2.3^{a}	3.0^{a}	52 ^b	13	***
d05	1-hexanol	874	A	4.1 ^{ab}	20 ^b	2.0^{a}	93°	17	***
d06	eucalyptol	1041	A	1.1 ^a	0.6ª	4.9 ^a	14 ^b	8.2	*
d07	1-octanol	1072	A	3.5 ^a	5.1 ^a	3.3ª	35 ^b	22	*
d08	3-nonen-1-ol	1157	\mathbf{B}^{f}	34 ^{ab}	53 ^a	15 ^b	3.8 ^b	44	ns ^(0.073)
d09	3,6-nonadien-1-ol	1165	\mathbf{B}^{f}	14	10	3.6	1.7	18	ns
d10	1-nonanol	1173	А	21	27	8.2	10	28	ns
	total alcohols			139	317	79	572		
Aldeby	des						0.2		
e01	2-methylbutanal	666	А	48	6.0	8.0	34	8.0	ns
e02	2-methyl 2-butenal	745	A	0.5ª	1.5^{a}	0.0^{a}	9.8 ^b	4 5	**
e03	hexanal	811	A	9.4	1.5	17	11	13	ns
e04	heptanal	907	A	8.0	7.6	9.0	9.0	6.7	ns
e05	benzaldehvde	974	A	9.9 ^{ab}	31 ^b	6.6 ^a	6.5 ^a	23	ns
e06	6-nonenal	1104	A	2.0^{a}	nd	13 ^b	nd	5.4	*
e07	nonanal	1108	A	30	27	36	35	33	ns
e08	decanal	1210	А	18 ^a	16 ^a	16 ^a	36 ^b	17	ns ^(0.062)
	total aldehydes			83	109	106	111		
Other 1	compounds								
f01	2-methylbutanenitrile	728	А	nd	0.4^{a}	1 1 ^a	56 ^b	18	***
f02	3-methylbutanenitrile	735	A	nd	nd	0.6^{a}	18 ^b	59	***
f03	limonene	1036	A	1.3	1.7	1.9	2.4	1.4	ns
f04	geranylacetone	1451	A	nd	0.2	1.3	4.4	5.0	Ns
101	Bermilia	1.01		110	0.2	110		0.0	110
Semi-v	volatile analysis								
Esters									
g01	2-acetoxy-3-butanone	1358	А	nd	nd	nd	4.6		
g02	2,3-butanediol diacetate ^f	1462	А	0.1^{a}	0.8^{a}	0.6^{a}	8.5 ^b	4.6	**
g03	1,2-propanediol diacetate	1486	А	nd	0.2^{a}	0.1^{a}	0.6 ^b	0.3	*
g04	2,3-butanediol diacetate ^f	1497	А	0.1^{a}	0.8^{a}	0.2^{a}	6.1 ^b	1.5	***
g05	1,2-ethanediol diacetate	1518	А	0.1^{a}	0.6^{a}	0.9 ^a	2.5 ^b	1.0	**
g06	2,3-butanediol monoacetate ^g	1536	А	0.2^{a}	0.6^{a}	0.2^{a}	10^{b}	6.5	*
g07	2,3-butanediol monoacetate ^g	1549	А	0.2^{a}	1.1^{a}	0.8^{a}	30 ^b	4.3	***
g08	1,3-butanediol diacetate	1593	В	nd	nd	nd	1.0		
g09	1,4-butanediol diacetate	1748	В	nd	nd	nd	1.2		
Sulfur-	containing compounds								
h01	ethyl (methylthio)acetate	1423	А	nd	nd	nd	6.3		
h02	2-(methylthio)ethyl acetate	1468	A	nd	nd	nd	21		

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Code	Compound	d LRI^a ID^b Approximate quantity ^c					tity ^c	LSD^{d}	\mathbf{P}^{e}	
			-	iLSL	mLSL	iMSL	mMSL			
h03	2-(methylthio)ethanol	1503	А	nd	nd	nd	4.6			
h04	3-(methylthio)propyl acetate	1601	А	nd	nd	nd	14			
h05	3-(methylthio)-1-propanol	1689	А	nd	nd	nd	5.2			
Other		1336	А	0.3 ^a	1.4 ^a	0.2^{a}	13 ^b	2.8	***	
i01	1-hexanol	1363	В	1.1^{a}	0.3 ^a	0.6^{a}	14 ^b	1.9	***	
i02	3-hexen-1-ol	1557	В	0.2^{ab}	0.1^{a}	0.6°	0.4^{bc}	0.2	**	
i03	2,6-nonadienal	1844	А	8.7 ^b	17 ^c	1.5 ^a	23 ^c	5.7	***	
i04	benzyl alcohol	1879	А	1.2 ^b	2.6 ^c	0.2^{a}	3.7 ^d	0.8	***	
i05	phenylethanol	1995	В	0.3 ^a	1.0^{b}	0.1^{a}	1.6 ^c	0.4	***	
i06	dihydro-3-hydroxy-4,4-dimethyl- 2(3H)-furanone	2014	В	0.2 ^a	0.6 ^a	nd	3.2 ^b	1.0	***	
i07	benzenepropanol	2064	В	nd	0.6^{a}	nd	2.5 ^b	1.5	*	
i08	2-ethyl-4-hydroxy-5-methyl- 3(2 <i>H</i>)-furanone	2081	В	2.0 ^a	15 ^b	0.6 ^a	13 ^b	6.2	**	
i09	4-hydroxy-5-methyl-3(2H)- furanone	2315	В	0.5 ^a	2.3 ^b	0.8 ^a	2.1 ^b	0.5	***	
i10	5,6,7,7a-tetrahydro-4,4,7a- trimethyl-2(4H)-benzofuranone	1336	А	0.3ª	1.4 ^a	0.2ª	13 ^b	2.8	***	
i11	hexadecanoic acid	2886	В	14 ^a	34 ^a	33 ^a	56 ^b	22	*	
i12	9-hexadecenoic acid	2928	В	5.9 ^{ab}	17 ^b	4.3 ^a	31 ^c	13	**	
Non-vol	atile analysis									
Organic	c acids			2.1	2.4	1.0	4.5	1.5		
j01 :02				3.1	3.4 0.5	4.0	4.5	1.5	ns	
j02	calle acid			0.4	0.5	0.5	0.4	0.2	ns	
Sugars	Emotoos			14	12	20	14	0.4		
KU1	iluciose			14	13	20	14	9.4	ns	
KU2	glucose			13 57 ^b	10 0 4 c	19 15 ^a	11 67 ^b	9.1	11S ***	
коз Бала ана	sucrose			57	04	15	07	10		
<i>F ree am</i>				200ª	714 ^b	771 ^a	1204 ^c	261	***	
101				299 103 ^b	714 220°	271 27ª	1304 02 ^b	301	***	
102				105 6 0 ^a	$\Delta \Delta 0$	$0 0^{ab}$	92 10 ^b	30	*	
103	Val			0.0 216 ^b	9.0 348°	5.0 50 ^a	160 ^b	5.0 60	***	
104	I en			210	21	59 75	20	17	ne	
105	Пе			25 20	31	23	39 10	17	ne	
100	Thr			121 ^b	174°	63 ^a	+∠ 100 ^{ab}	15 /6	**	
107	γ-ABA			1485 ^b	2216°	371 ^a	515 ^a	-+0 388	***	
109	Ser			402 ^b	673°	162^{a}	336 ^{ab}	103	**	
110	Pro			-102 65°	9023 90d	76 ^a	Δ1 ^b	175	***	
111	Asn			171 ^b	22 252°	20 111 ^a	136 ^{ab}	13	***	
117	Asn			35/1/b	252 5627°	170/a	12/12 ^a	43 1015	***	
112	Met			63°	106 ^d	1∠24 01ª	1243 27 ^b	1015	***	
11.5	Chu			305 ^{ab}	569 ^b	∠1 15ª	580b	362	*	
114	Dha			505 62 ^b	120°	13 27 ^a	209 40 ^{ab}	303 40	**	
115	Gln			6449 ^b	8659 ^b	3176 ^a	49 2460 ^a	2515	**	
117	Lys			19	21	20	28	13	ns	

Code	Compound	\mathbf{LRI}^{a}	ID^b	Ap	oproxima	LSD^{d}	P ^e		
				iLSL	mLSL	iMSL	mMSL		
118	Tyr			21 ^a	31 ^b	14 ^a	22^{ab}	9.0	*
119	Trp			21 ^b	33 ^c	7.0^{a}	10 ^a	6.0	***

^a For compounds a to f: linear retention index on DB-5 column, for compounds g to i: linear retention index on a DB-WAX. ^b Å, mass spectrum and LRI agree with those of authentic compound; B, mass spectrum agrees with reference spectrum in the NIST/EPA/NIH mass spectra database and LRI agree with those in the literature (reference given). ^c For compounds a to f: estimated quantities (ng) collected from the headspace of 2 ml of melon juice diluted in 10 ml of HPLC water, calculated by comparison with 130.6 ng of 1,2-dichlorobenzene used as internal standard; for compounds g to i: estimated quantities (mg) from 20 ml melon juice, calculated by comparison with 100 mg of 3-chlorophenol used as internal standard; for compounds j and k: estimated quantities (g/l) of melon juice and for compounds l: estimated quantities (mg/l) of melon juice; means not labelled with the same letters are significantly different (p < 0.05); means of three replicate samples; nd, not detected.^d Least significant difference at p = 0.05.^e Probability, obtained by ANOVA, that there is a difference between means; ns, no significant difference between means (p > 0.05); * significant at the 5% level; ** significant at the 1% level; *** significant at 0.1% level, # difference between samples (absent vs. present) but no significant difference between those samples where the compound was present. fg Pair of diastereoisomers. A COLORINA AND A COLO

Code	Compound	LRIexpt ^a	Odour description	Intensity ^b			
				iLSL	mLSL	iMSL	mMSL
1	ethyl propanoate	713	fruity, over-ripe	-	-	-	9
2	propyl acetate	715	pungent, sweet fruit	-	-	-	12
3	ethyl 2-methylpropanoate	759	fruity, pineapple	-	10	6	12
4	methyl 2-methylbutanoate	778	fruity, pineapple	9	11	9	11
5	hexanal	805	green, grass	4	9	7	6
6	ethyl butanoate	806	sweet fruity, fake	-	-	-	10
7	ethyl 2-methylbutanoate	849	fruity sweet, pineapple	8	11	8	13
8	3-hexen-1-ol	856	fresh-cut grass	-	-	-	5
9	1-hexanol	870	herbaceous	-	-	-	5
10	4-heptenal	902	lamb fat, cheesy	-	-	11	-
11	butyl propanoate	911	ripe banana	-	-	-	4
12	S-methyl 2-methylbutanethioate	940	sulfury	7 -	-	5	3
13	dimethyl trisulfide	972	pickled onions, cabbage	10	13	9	13
14	ethyl (methylthio)acetate	985	earthy, slightly	-	-	-	5
15	eucalyptol	1032	pine	-	-	-	3
16	ethyl 3-(methylthio)propanoate	1102	cardboard, slightly	-	-	-	4
17	6-nonenal	1110	cucumber	10	-	12	-
18	3,6-nonadien-1-ol	1164	rags, dry	8	5	4	3

Table 2. Odorants identified by GC-O/MS in the headspace of two genotypes of Charentais melon harvested at two different maturity stages.

^{*a*} Linear retention index on DB-5 column, calculated from a linear equation between each pair of straight chain *n*-alkanes C₆-C₂₅. ^{*b*} The sum of intensities recorded by two assessors for each sample (scoring scale: weak = 3, medium = 5, strong = 7), - = not detected.

ibits ibits <t< th=""><th>Code</th><th>Attribute</th><th></th><th>Sc</th><th>ore^a</th><th></th><th>LSD^b</th><th></th><th>\mathbf{P}^{c}</th><th></th></t<>	Code	Attribute		Sc	ore ^a		LSD ^b		\mathbf{P}^{c}	
Odduar Odduar o01 sweet 41 ^b 41 ^b 40 ^b 50 ⁱ 5.4 #* *** ms o02 floral 11 ^b 10 ^b 14 ^b 21 ^a 6.4 **** ms o03 honey 11 ^b 10 ^b 14 ^b 21 ^a 4.4 **** ms o05 orange squash 13 ^a 18 ^a 14 ^b 18 ^a 4.4 ms *** ms o06 citrus 10 10 11 10 2.8 ms *** ms o07 cucumber 17 ^b 12 ^c 22 ^a 12 ^c 4.1 *** ms o08 green ⁴ 18 ^d 14 ^b 8.1 ^c 11 ^b 4.0 *** ms o11 brown orchard fruit' 11 11 11 14 4.8 ms *** ms o12 ript tropical fruit' 11 ^b 10 ^b 9.9 ^b 9.2 ^b			iLSL	mLSL	iMSL	mMSL		S	Α	Ι
old sweet 41^p 41^p 40^p 50^a 54^a $***$ $***$ ns ol2 floral 17^b 19^b 21^a 26^b 6.4 **** ns ol3 honey 11^b 10^b 14^a 21^a 4.4 $***$ ns od3 honey 11^b 10^b 14^a 4.4^a $***$ ns od4 strawberries 6.5^b 10^{abb} 8.8^b 14^a 4.4 $***$ ns od5 citrus 10 10 11 10^b 2.8 ns $***$ ns of0 cutumer 17^b 12^c 22^a 12^c 4.1^a $***$ ns		Odour								
c02 floral 17^b 19^b 21^a $2e^b$ 6.4 $****$ ns c03 honey 11^b 10^b 14^b 21^a 4.4 $****$ ns c04 stawberries 6.5^b 10^{ab} 8.8^b 14^a 4.4 $****$ ns c05 orange squash 13^a 18^a 14^{ab} 12^c 4.1 $****$ ns c06 citrus 10 11 10 22^a 4.1 $****$ ns c07 cucumber 17^b 12^c 22^a 4.1 $****$ ns $****$ ns c08 green ⁴ 14^b 11^b 11^b 10^b 9.9^b $9.b^b$ 6.3 $***$ ns ns	o01	sweet	41 ^b	41 ^b	40 ^b	50 ^a	5.4	**	***	ns
c03 honey 11^b 10^{bb} 14^b 21^a 4.4 *** *** ns c04 strawberries 6.5^b 10^{ab} 8.8^b 14^a 4.4 *** *** ns c05 orange squash 13^a 18^a 14^{ab} 8.8^b 14^a 4.4 *** *** ns c06 citrus 10 10 11 10 2.8 ns *** ns c07 cucumber 17^b 12^c 22^1 12^c 4.1 *** *** ns c00 carthy 18^a 14^{bb} 21^a 11^b 4.0 *** ns ol1 brown orchard fmit' 13^a 10^b 9.2^b 13^a 2.9 *** ns ns **** ns ol1 sweet 60^a 66^a 31^b 65^a 8.8 **** ns t*** ns t03 slty 18^a 12^{bc} 17^a 14^c	002	floral	17 ^b	19 ^b	21^{ab}	26 ^b	6.4	*	***	ns
oO4 strawberries 6.5^{b} 10^{ab} 8.8^{b} 14^{a} 4.4 $***$ ns oO5 orange squash 13^{a} 18^{a} 14^{b} 18^{a} 4.4 ns $***$ ns oO6 citrus 10 10 11 10 2.8 ns $***$ ns oO7 cucumber 17^{b} 12^{c} 22^{a} 12^{c} 4.1 $***$ ns oO8 green ^d 14^{b} 14^{b} 21^{a} 11^{b} 4.0 $***$ ns oO10 musty 16^{a} 8.9^{b} 5.1^{b} 90^{b} 6.3 $***$ ns o11 brown orchard fruit' 11 01^{b} 9.9^{b} 17^{a} 4.4 $***$ ns o12 ripe tropical fruit' 11 11 11 11^{c} 3.1 $***$ ns o13 fermenting 13^{a} 9.9^{b} 12^{a} 13^{a} 4.8 $***$ ns	o03	honey	11 ^b	10^{b}	14 ^b	21 ^a	4.4	***	***	ns
o05 orange squash 13^a 18^a 14^{ab} 18^a 4.4 ns **** ns o06 citrus 10 10 11 10 2.8 ns **** ns o07 cucumber 17^b 12^c 22^a 12^c 4.1 **** ns o08 green ² 14^b 44^b 8.1^c 11^{bc} 5.5 *** *** ns o09 earthy 16^a 8.9^b 5.1^b 9.0^b 6.3 *** ns *** ns o10 musty 16^a 8.9^b 5.1^b 9.0^b 6.3 *** ns *** ns o11 brown orchard fruit' 13^a 9.9^b 9.2^b 13^a 2.9 *** ns ms ns *** ns **** ns **** n	o04	strawberries	6.5 ^b	10^{ab}	8.8^{b}	14 ^a	4.4	**	***	ns
o06 citrus 10 10 11 10 2.8 ns **** ns o07 cucumber 17 ⁵ 12 ^c 22 ^a 12 ^c 4.1 **** ns o08 green ^d 14 ^b 14 ^b 21 ^a 11 ^b 4.0 **** ns o09 carthy 18 ^a 14 ^b 8.1 ^c 11 ^{bc} 5.5 *** ns o10 musty 16 ^a 8.9 ^b 5.1 ^b 9.0 ^b 6.3 *** ns o11 brown orchard fruit ^e 11 11 11 14 3.8 ns *** ns o12 ripe tropical fruit ^e 11 11 11 14 3.8 ns *** ns o13 fermenting 13 ^a 9.9 ^b 9.2 ^b 13 ^a 2.9 *** ns t013 savet 60 ^a 66 ^a 31 ^b 65 ^a 8.8 **** ns t03 salty 18 ^a 15 ^a 13 ^b 14 ^a 3.0 s**	005	orange squash	13 ^a	18 ^a	14^{ab}	18 ^a	4.4	ns	***	ns
o07 cucumber 17^b 12^c 22^a 12^c 4.1 **** **** ms o08 green ^d 14^b 14^b 21^a 11^b 4.0 **** ms ms o09 earthy 16^a 8.9^b 5.1^b 9.0^b 6.3 *** ms	006	citrus	10	10	11	10	2.8	ns	***	ns
o08 grcen ^d 14 ^b 14 ^b 21 ^a 11 ^b 4.0 *** *** ns o09 earthy 18 ^a 14 ^b 8.1 ^c 11 ^{bc} 5.5 ** *** ns o10 musty 16 ^a 8.9 ^b 5.1 ^b 9.0 ^b 6.3 ** ** ns o11 brown orchard fruit ^c 13 ^a 0.9 ^b 9.2 ^b 13 ^a 2.9 ** *** ns o13 fermenting 13 ^a 9.9 ^b 9.2 ^b 13 ^a 2.9 ** *** ns t01 sweet 60 ^a 66 ^a 31 ^b 65 ^s 8.8 *** ns *** ns t103 satuy 18 ^a 15 ^b 13 ^b 4.4 ^a and ns *** ns titt t04 acidic 15 17 20 15 4.7 ns *** ns titt t05 bitter 17 14 15 13 4.8 ns *** *** ts <td>007</td> <td>cucumber</td> <td>17^b</td> <td>12^c</td> <td>22^a</td> <td>12^c</td> <td>4.1</td> <td>***</td> <td>***</td> <td>ns</td>	007	cucumber	17 ^b	12 ^c	22 ^a	12 ^c	4.1	***	***	ns
or 009earthy earthy18° 16°14°b 8.1°11°c 11°5.5 5.5 8.8 9.0°*** 8.8 8.1°*** 9.0°ns 6.3 8***** 8*** 8*** 8*** 8*** 8*** 8***010musty16° 8.9°8.9° 9.9°17° 9.9°4.4 4.4 4***** *** **** 8*** 8***011brown orchard fruit' 1111111114 143.8 15***** 8*** **** **** 8***013fermenting raste/Flavour13° 13°9.9° 9.2°9.2° 13°2.9 ************** **** **** ****t001sweet60° 66° 66° 31°65° 4.3 13°8.8 4.3 4.3 4.3 13°**** **** **** ****t003salty18° 12° 15°17° 12° 1513 4.8 4.3 4.4**** **** ****t004acidic 1517 17 1415 13 4.8 8.4 4.3 4.4**** **** ****t005bitter 17° 17° 14° 14° 10° 10° 13° 3.5° 13° 4.8 4.8 4.4 <b< td=""><td>008</td><td>green^d</td><td>14^b</td><td>14^b</td><td>21^a</td><td>11^b</td><td>4.0</td><td>***</td><td>***</td><td>ns</td></b<>	008	green ^d	14 ^b	14 ^b	21 ^a	11 ^b	4.0	***	***	ns
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$\begin{array}{cccccccccccccccccccccccccccccccccccc$	011	brown orchard fruit ^e	13 ^{ab}	10 ^b	9 0 ^b	17 ^a	<u>о.</u> 1 Л	**	***	ne
111111141414141414141415132.9***#**nsTaste/Flavourfermenting 13^a 9.9^b 9.2^b 13^a 2.9 ******nstf01sweet 60^a 66^a 31^b 65^a 8.8 *********tf02savoury 15^{ab} 12^{bc} 17^a 11^c 3.1 ******nstf03salty 18^a 15^{ab} 13^b 43^b 4.3 ns***nstf04acidic 15 17 20 15 4.7 ns***nstf05bitter 17^a 14^a 15^{2a} 4.8 ns***nstf06floral 21^a 19^{ab} 44^b 26^a 6.3 ******nstf05bitter 17^a 14^a 9.2^b 18^a 5.1 ******nstf06floral 21^a 19^{ab} 42^b 26^a 6.3 ********tf09strawberries 7.5^b 7.9^b 3.5^b 13^a 4.8 *********tf10orange squash 11 9.1 11 11 47 ns******tf12cucumber 16^b 10^{bc} 23^a 9.4^c 6.4 ******nstf13green 11^b 8.5^b <	012	ripe tropical fruit	15	11).) 11	1/	т.т 2 8	no	***	ne
ord Taste/Flavour 1.3 9.9 9.2 1.3 2.9 $***$ Ins t01 sweet 60^a 66^a 31^b 65^a 8.8 *** *** Ins t02 savoury 15^{ab} 12^{bc} 17^a 11^c 3.1 *** *** ns t03 salty 18^a 15^{ab} 13^b 13^b 4.3 ns *** ns t04 acidic 15 17 20 15 4.7 ns *** ns t05 bitter 17^a 14^a 9.2^b 18^a 5.1 ** *** ns t06 floral 21^a 19^{ab} 14^a 9.2^b 18^a 5.1 *** *** ns t07 honey 17^a 14^a 9.2^b 18^a 5.1 *** *** s t08 strawberries 7.5^b 7.9^b 3.5^b 13^a 4.8 *** *** *** <td< td=""><td>012</td><td>formonting</td><td>11 12^a</td><td></td><td>0.2^b</td><td>14 12⁸</td><td>2.0</td><td>**</td><td>***</td><td>115</td></td<>	012	formonting	11 12 ^a		0.2 ^b	14 12 ⁸	2.0	**	***	115
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tf04acidic151720154.7ns***nstf05bitter171415134.8ns***nstf06floral 21^a 19^{ab} 14^b 26^a 6.3*******tf07honey 17^a 14^a 9.2^b 18^a 5.1*****nstf08syrupy 37^a 41^a 10^b 37^a 9.4 ********tf09strawberries 7.5^b 7.9^b 3.5^b 13^a 4.8 *******tf10orange squash 11 9.1 11 11 4.7 ns***nstf11citrus 6.4^b 6.5^b 11^a 8.4^b 2.7 ***nstf12cucumber 16^b 10^{bc} 23^a 9.4^c 6.4 ***nstf13green 11^b 8.5^b 17^a 9.8^b 4.1 ***mstf14metallic 22^a 17^b 17^a 20^b 3.9 $ns^{(0.050)}$ ***nstf15pithy 17 16 13 12 7.6 ns***mstf15pithy 17^a 17^a 6.9^b 18^a 6.3 ***nstf16earthy 22^a 17^b 11^b 1^5^a 5.7 ****nstf17musty 18^a 15^a <t< td=""><td>tf03</td><td>salty</td><td>18^a</td><td>15^{ab}</td><td>13^b</td><td>13^b</td><td>4.3</td><td>ns</td><td>***</td><td>ns</td></t<>	tf03	salty	18 ^a	15^{ab}	13 ^b	13 ^b	4.3	ns	***	ns
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111chus 0.7 0.5 11 0.7 2.7 1.4 1.8 tf12cucumber 16^{b} 10^{bc} 23^{a} 9.4^{c} 6.4 $***$ $***$ $***$ tf13green 11^{b} 8.5^{b} 17^{a} 9.8^{b} 4.1 $***$ $***$ $***$ tf14metallic 22^{a} 17^{b} 17^{a} 20^{ab} 3.9 $ns^{(0.050)}$ $***$ ns tf15pithy 17 16 13 12 7.6 ns $***$ ns tf16earthy 22^{a} 17^{b} 11^{b} 11^{b} 5.7 $***$ ns tf16earthy 22^{a} 17^{b} 11^{b} 11^{b} 5.7 $****$ ns tf17musty 18^{a} 15^{a} 5.4^{b} 13^{a} 6.1 $****$ ns tf18brown orchard fruit ^e 17^{a} 17^{a} 6.9^{b} 18^{a} 6.3 $***$ ns tf19ripe tropical fruit ^e 9.8^{b} 13^{ab} 8.1^{b} 16^{a} 5.7 $*$ $****$ m01mouth drying 41 41 37 40 6.2 ns $****$ ns m01mouth coating 41^{ab} 43^{a} 32^{c} 37^{bc} 5.5 $****$ ns m03tongue tingling 8.0 6.9 7.9 7.9 4.0 ns $****$ ns m04body 46^{a	tf11	citrus	6 /b	6.5 ^b	11 ^a	8 /b	т., Э.7	**	***	ne
112Current of the term of term o	tf12	cucumber	16 ^b	10 ^{bc}	73 ^a	0.4 0 /c	2.1 6.1	***	***	***
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m04body 46^{a} 46^{a} 24^{b} 42^{a} 7.5 *******m05salivating333232325.7ns***nsm06smoothness 44^{ab} 44^{ab} 37^{b} 47^{a} 6.5 $ns^{(0.052)}$ ***nsae01sweet 50^{a} 55^{a} 26^{b} 52^{a} 10 *********ae02sayoury 14 11 16 14 4.8 ns	m03	tongue tingling	8.0	6.9	7.9	7.9	4.0	ns	***	ns
m05 salivating 33 32 32 32 5.7 ns *** ns m06 smoothness 44^{ab} 44^{ab} 37^{b} 47^{a} 6.5 $ns^{(0.052)}$ *** ns ae01 sweet 50^{a} 55^{a} 26^{b} 52^{a} 10 *** *** *** ae02 sayoury 14 11 16 14 4.8 ns	m04	body	46 ^a	46 ^a	24 ^b	42 ^a	7.5	***	***	*
m06 smoothness 44^{ab} 44^{ab} 37^{b} 47^{a} 6.5 $ns^{(0.052)}$ *** ns $After-effects$ $ae01$ sweet 50^{a} 55^{a} 26^{b} 52^{a} 10 *** *** *** $ae02$ sayoury 14 11 16 14 4.8 ns	m05	salivating	33	32	32	32	5.7	ns	***	ns
After-effects ae01 sweet 50^a 55^a 26^b 52^a 10 *** *** *** ae02 sayoury 14 11 16 14 4.8 ns	m06	smoothness	44 ^{ab}	44 ^{ab}	37 ^b	47 ^a	6.5	ns ^(0.052)	***	ns
$ae01$ sweet 50^a 55^a 26^b 52^a 10 *** *** $ae02$ sayoury 14 11 16 14 4.8 ns		After-effects								
-ae02 sayoury 14 11 16 14 4.8 ns	ae01	sweet	50 ^a	55 ^a	26 ^b	52 ^a	10	***	**	***
		savourv	14	11	16	14	4.8	ns	***	*

Table 3. Mean panel scores for sensory attributes of two genotypes of Charentais melon harvested at two different maturity stages.

Code	Attribute	Score ^{<i>a</i>}				LSD ^b	\mathbf{P}^{c}		
		iLSL	mLSL	iMSL	mMSL		S	Α	Ι
ae03	salty	15	15	13	13	4.4	ns	***	*
ae04	acidic	15 ^{ab}	13 ^b	21 ^a	13 ^b	5.7	*	***	*
ae05	bitter	16 ^{ab}	14 ^b	19 ^a	14 ^b	4.2	ns ^(0.050)	***	ns
ae06	mouthcoating	42 ^a	43 ^a	33 ^b	41 ^a	4.6	**	***	*
ae07	drying	42	43	39	42	7.8	ns	***	***
ae08	musty	21 ^a	17 ^a	8.4 ^b	15 ^{ab}	7.3	*	***	*
ae09	soapy	4.5	5.2	8.9	6.6	5.0	ns	***	ns
ae10	metallic	22	22	19	18	6.5	ns	***	**

^a Means not labelled with the same letters are significantly different (p < 0.05); means are from three replicate samples. ^b Least significance difference at p = 0.05. ^c Probability, obtained from ANOVA, that there is a difference between means; ns, no significant difference between means (p > 0.05); * significant at the 5% level; ** significant at the 1% level; *** significant at the 0.1% level; F-ratios for sample and assessor were calculated by comparing is as dotted to the second tot the mean square of the effect with the mean square of the samplexassessor interaction; S: significance of samples, A: significance of assessors, I: significance of the interaction (S×A).^d Odour associated with freshly cut grass and green beans. ^e Odour or taste-flavour associated with overripe apples and pears. ^f Odour or taste-flavour associated

Highlights

- Flavour of medium and long shelf-life Charentais cantaloupe melons was compared.
- Volatile and semivolatile profiles were correlated with sensory data using multifactorial analysis.
- Maturity at harvest has a significant impact on the flavour of medium-shelf life fruit.
- Maturity at harvest had much less impact on a long shelf-life genotype.

• Esters and sulfur-compounds were more abundant in mature medium shelf-life fruit.