

Advances in ophthalmic drug delivery

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Advances in Ophthalmic Drug Delivery

2 Peter W. J. Morrison, Vitaliy V. Khutoryanskiy* 3 School of Pharmacy, University of Reading, Whiteknights, PO Box 224, Reading, RG6 6AD, 4 United Kingdom. E-mail: v.khutoryanskiy@reading.ac.uk; Tel: +44(0)1183786119 5 **Abstract:** 6 Various strategies for ocular drug delivery are considered; from basic formulation techniques for 7 improving availability of drugs; viscosity enhancers and mucoadhesives aid drug retention and 8 penetration enhancers promote drug transport into the eye. The use of drug loaded contact lenses 9 and ocular inserts allows drugs to be better placed where they are needed for more direct 10 delivery. Developments in ocular implants gives a means to overcome the physical barriers that 11 traditionally prevented effective treatment. Implant technologies are under development allowing 12 long term drug delivery from a single procedure, these devices allow posterior chamber diseases 13 to be effectively treated. Future developments could bring artificial corneas to eliminate the need 14 for donor tissue and one-off implantable drug depots lasting the patient's lifetime. 15 **Key Terms** 16 Bandage contact lens: Device designed to fit directly onto the front of the eye to offer 17 protection during the healing process, for example, after corneal surgery. 18 Container molecule: Molecular structures with cavities that can accommodate another molecule

via guest – host complexation.

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- 20 **Hydrotrope:** Water-soluble compound that improves the aqueous solubility of hydrophobic or
- 21 poorly water-soluble compounds.
- 22 In situ gelling system: Liquid formulations that turn in to gel upon dosage form administration.
- 23 These phase transitions can typically be triggered by changes in temperature, pH or electrolyte
- 24 interaction.
- 25 **Mucoadhesive:** Defined as a compound, usually a polymer, with the ability to adhere to mucosal
- tissue.
- Ocular insert: A drug-loaded device designed to reside within the ocular cul-de-sac, attach to
- 28 the conjunctiva or directly onto the cornea.
- 29 **Ocular implant:** Dosage forms implanted directly into the ocular globe; these can be devices
- that bring 'quality of life benefit' such as intraocular lenses used for crystalline lens replacement.
- 31 Implantable devices are also used for sustained and controlled drug delivery to the posterior
- 32 segment.
- 33 'Smart' DDS: Responsive drug delivery systems where a favourable change takes place in
- response to some form of stimulus, for example, change in temperature, pH, ionic interactions or
- 35 stimulation from a light source.

Introduction

- Ocular drug delivery is hampered by the physiological barriers presented by the eyes. These
- 38 include, blinking and wash out by tears, nasolacrimal drainage, non-productive losses and
- impermeability of the cornea. [1,2]

Some of the various structures of the eye are detailed in **Figure 1**, highlighting the intricate complexity of this organ. The conjunctiva (not shown for clarity) is the mucosa lining the inside surface of the eyelids and the external surface of the front of the eye up to the limbus, the edge of the cornea.

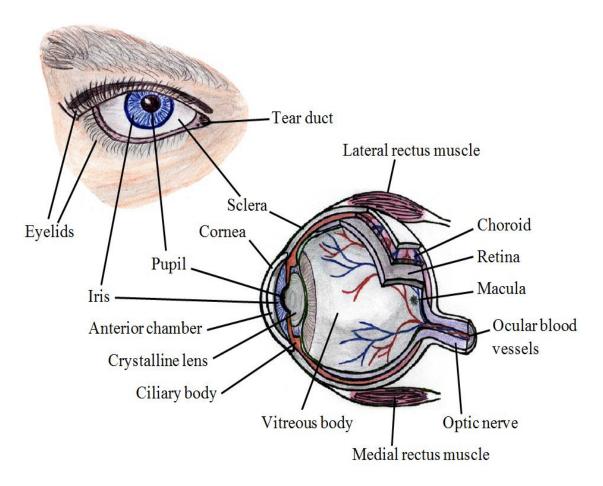


Figure 1. A sketch showing some of the key features of the human eye.

Despite the easy accessibility of the eye for administering medication, in many ways it is an isolated organ with several barriers imposing challenges to drug delivery, tear mechanisms, the physical barriers of its membranes, blood-aqueous and blood-retinal barriers.[3]

Topical, systemic and intraocular are the three main routes for administering ophthalmic medication; each has their own advantages and disadvantages. Topical drug delivery is the most accepted route accounting for ~90% aqueous ophthalmic formulations. Advantages are their relative simplicity to formulate, minimal storage limitations and ease of drug instillation by most patients. Disadvantages include limited drug concentration for lipophilic agents, pre-corneal losses and the barrier function of the cornea.[4,5] For effective systemic delivery a relatively high drug concentration needs to be circulating in the blood plasma in order to achieve a therapeutically effective dose within the eye. Sustained release oral drugs can be suitable for glaucoma patients, allowing for continuous and effective treatment, however this method exposes the whole body to the drug often giving rise to undesired side effects.[6] Intraocular drug delivery by intravitreal injection is an invasive procedure carrying a degree of risk such as retinal hemorrhage or detachment, especially if the technique needs to be repeated when treating chronic disorders. However, it is very effective at getting drugs to the posterior segment.[3] The cornea is the main route for topically applied drugs to gain access into the eye and the conjunctival/scleral route can also be efficient. [7,8] Drops are the most accepted means to apply medication to this organ; [9] they are easy to apply by most patients and they are convenient. However, regardless of the ease of access to the eye for topical application of medication, efficient ocular drug delivery is hampered by a series of clearance mechanisms that protect the ocular structures from foreign matter. Upon administration of traditional eye drops they are immediately diluted in the tear film followed by very quick elimination by action of blinking, wash out by tears, and nasolacrimal drainage. [10,11] After instilling eye drops, there remains a very short time where any residual medication is in contact with the cornea during which time there is opportunity for the drug to penetrate into the eye; however, due to poor corneal

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permeability only a very small portion of active pharmaceutical ingredient will be capable of crossing the cornea. Of the applied dose, only 1% or less will successfully reach the intended target in most cases, the rest will be systemically absorbed via the conjunctiva or nasolacrimal mucosa to be eliminated by metabolic processes.[5] The tear film comprises of several compartments, **Figure 2** shows the 3 layer tear film model comprising of a coating of mucous anchored to the epithelium via microvilli, an aqueous compartment containing soluble mucin and free lipid and a thin lipid layer [11-14].

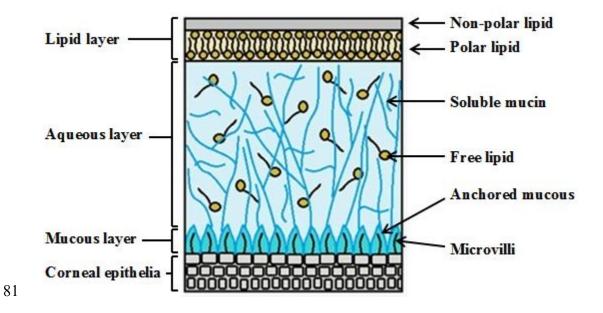


Figure 2. The 3 layer tear film model.

The tear film and ocular mucosa are the first external barriers to overcome, after which the multilayered structure of the cornea (**Figure 3**) offers the next challenge; this structure has both lipophilic and hydrophilic properties and there are 5 distinct layers: Epithelium, Bowman's membrane, stroma, Descemet's membrane and endothelium.[6,15] The first corneal layer is the epithelium which is \sim 50 μ m at its center increasing to \sim 100 μ m at the limbus; this layer is

lipophilic, offering ~90% resistance to hydrophilic drugs and ~10% to hydrophobic preparations. Immediately underneath the epithelium is the Bowman's membrane, a transitional acellular structure ~8-14 μ m in thickness. Next we find the hydrophilic stroma; this is a gel-like structure with around 80 % water, consisting of collagen, mucopolysaccharides and proteins and it forms the main bulk of the cornea, some 90 % of its total thickness. Next there is the Descemet's membrane, a tough membrane of around 6 μ m thickness supporting the endothelium, a single layer of loose, epithelia-like cells important in regulating stromal hydration, and this layer is deposited by endothelial cells. The correct level of hydration is important for the cornea to remain clear and transparent.[6,15,16]

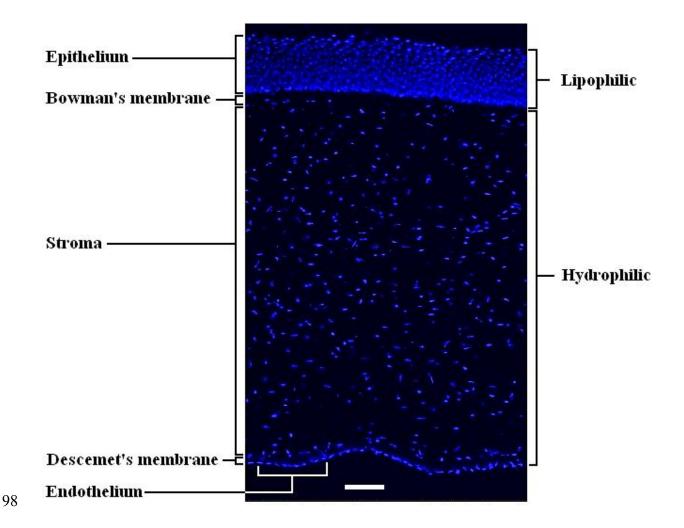


Figure 3. Micrograph of a section of bovine cornea showing the multi-layered structure typical of mammalian corneas. Scale bar = $100 \mu m$.

The corneal epithelial barrier also has different zones; the basement layer consists of newly formed cells firmly attached to the Bowman's layer, here they are columnar in shape. As new cells are formed the preceding basement cells are pushed forwards, becoming polyhedral in shape, eventually as they are moved towards the corneal surface where they become polygonal squamous cells. These superficial epithelial cells have Ca²⁺ dependent membrane adherent regions; zonula occludens, zonula adherens and desmosomes forming tight junctions.[17] Taken together, these tightly bound cell membrane regions and the lipophilic nature of the

epithelium make the structure an extremely efficient barrier that resists intrusion of foreign material including potentially therapeutic compounds; this creates a major challenge for ocular drug delivery.[6,11,18]

Strategies for enhancing ocular drug delivery

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Despite traditional eye drops being convenient and simple to use, they are not very efficient and only a small amount of the dose is effectively delivered to its intended target, most is lost due to clearance mechanisms. There are however certain strategies that can be employed to improve the bioavailability of drugs. First, solubility enhancers can be used, to improve drug concentrations within the formulation; more medication in the dosage form can mean increased bioavailability. This strategy could allow a smaller droplet to be applied, which would be less susceptible to loss by drainage due to induced reflex tearing and blinking.[6] Second, the formulation can be designed in a form that resists clearance; these dosage forms are retained for a longer period, therefore they have more time to interact with ocular tissue. Next, drug penetration enhancers can be incorporated into the formulation to assist their transit across the cornea.[19] Ocular inserts are another area of active research and development. With this method a drug-loaded device resides in the cul-de-sac under the eyelids or fits directly on the cornea like a contact lens; these devices are often designed with controlled release in mind.[20,21] Drug delivery into the cornea and anterior chamber is difficult enough; delivering an effective therapeutic dose to the posterior segment is a major challenge, in many cases it is not possible to deliver sufficient medication to the posterior structures via the topical route.[22] For diseases of the retina, such as age-related macular degeneration (AMD), diabetic retinopathy, and retinitis pigmentosa and related ocular neovascular disease there is often a need to resort to invasive methods for drug delivery. Angiogenesis inhibitor medication via intravitreal injection is an option for getting drugs to the posterior segment but these are often effective for the short term and need repeat injections, which carries risks such as hemorrhage, endophthalmitis, ocular hypertension and retinal detachment.[22-26] Ocular implants are devices that penetrate the sclera or reside within the deeper ocular structures to deliver drugs for an extended period, sometimes many years, minimising the need for repeat injections.[23] Implantable devices that are not designed to deliver drugs are also employed to improve the 'quality of life' for patients with certain conditions, for example, intraocular lenses. However, drugs to counter postoperative bacterial infection are often included in these devices for short term protection.[27,28] These various strategies will be discussed in more detail in the following sections.

Solubility enhancers:

Discovery of potentially therapeutic compounds is accelerating through developments in genomics, combinational chemistry and the ability to use high throughput screening. High proportions of newly screened compounds prove to be hydrophobic and are poorly water-soluble.[29] For efficacious performance in the physiological environment drug candidates need to interact within an aqueous media, the interstitial fluids within tissues.

Drugs used for treatment of ocular disorders often have low aqueous solubility and eye drops are only in contact with ocular tissue for a short time. Formulations that are developed to increase the amount of available drug in solution could improve its bioavailability, therefore solubility enhancement is an important strategy to use when developing ocular medication. Solubility enhancement can be achieved by employing hydrotropic compounds. Evstigneev *et al.*[30] and Coffman and Kildsig [31,32] reported the effectiveness of caffeine, urea and nicotinamide and its derivatives as efficient hydrotropes for enhancing the solubility of riboflavin, a vitamin with poor

aqueous solubility of less than 0.1 mg mL⁻¹ which is used as a photosensitive drug for the treatment of keratoconus. Cyclodextrins are a class of cyclic supramolecular compounds that have been well studied for dissolution enhancement of low solubility drugs; Loftsson and Stefansson discussed the use of cyclodextrins for complexation with steroids, carbonic anhydrase inhibitors, pilocarpine and cyclosporins in eye drop formulations which are well tolerated.[33] Morrison et al.[34] investigated cyclodextrins for their hydrotropic properties and were able to show that β-cyclodextrin achieved solubility enhancement of more than 140% for riboflavin. Whilst the above mentioned studies achieved modest solubility enhancements, research by Kim et al. [29] investigating the performance of two hydrotropes; N,N-diethylnicotinamide (DENA) and N,N-dimethylbenzamide (DMBA) with 13 poorly water-soluble drugs and these compounds were shown to have superior hydrotropic action between 1000- to 10000- fold. Supramolecular structures are sub-micron sized molecules within the realm of nanotechnology and many of these assemblies have solubility enhancement properties. This technology is becoming an important tool within the pharmaceutical industry with substantial investment within the global market. Dendrimers, microemulsions, nanoparticles, nanosuspensions and liposomes belong to this class of compound and are proving to be useful structures to improve bioavailability, all of which are at the forefront of research in ocular drug delivery.[1,2,35-41] Micelles are aggregates of amphiphilic molecules forming self-assembled spheres in aqueous media. They have a monolayer 'shell' of polar groups with their associated fatty acid 'tails' forming the core. These are useful carriers of hydrophobic drugs within the core albeit with limited efficiency due to a high amphiphile / drug ratio.[42] The work of Qu et al.[43] involved chemical modification of chitosan by increasing their hydrophobicity and this allowed them to produce 100 – 300 nm sized micellar clusters which could achieve up to an order of magnitude

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enhancement in hydrophobic drug bioavailability compared to micelles produced using triblock copolymers. In ocular drug formulations they were able to show an initial prednisolone concentration in the aqueous humor equivalent to that found when using a 10-fold dose of prednisolone suspension.

An approach taken by Kulkarni *et al.* [44] was to take the poorly soluble drug, indomethacin, and using simple chemistry, convert this drug into its sodium salt. They found that this improved its aqueous solubility and the drug was stable at physiological pH and compatible with excipients used for ocular drug formulation.

Penetration enhancement:

Materials that modify the corneal epithelia can allow enhancement of drug permeation and this can be achieved using various strategies. Benzalkonium chloride (BAC) is commonly used as a preservative in ocular drug formulations, this together with other compounds; cetylpyridinium chloride (CPC), ethylenediaminetetraacetic acid (EDTA), polyoxyethylene strearyl ether (PSE) and polyethoxylated castor oil (PCO) are compounds with penetration enhancing properties. Their mode of action is due to destabilisation of the tear film and the protection given by its mucus component (for BAC), and ultrastructural alterations [17] and solubilisation of cellular membranes for the other enhancers. Useful as they are for penetration enhancement they can also induce irritation and damage to ocular epithelium even at low concentrations. Chung *et al.* [45] and Burgalassi *et al.* [46] investigated these materials confirming their irritation and cytotoxicity effects. Liu *et al.* [47] state that penetration enhancers should be:

- Non-toxic;
- Non-irritant to the eye;

- Inert and compatible to other excipients within the formulation;
- Fast acting and reversible action;
- Effective at low concentration.

201 In their report they discuss the use of several penetration enhancers for ocular drugs; BAC, 202 EDTA, surfactants, heteroglycosides, bile salts, polycarbophil-cysteine conjugates and boric 203 acid, all of which have been used in ophthalmic formulations despite the fact that even at low 204 concentrations they can cause ocular irritation.[47] Morrison et al. [17] investigated drug 205 penetration enhancement using EDTA and two analogues EGTA and EDDS and they found that this was achieved by sequestering Ca²⁺ and therefore loosen tight junctions which depend on the 206 207 availability of these ions. 208 Gelucires are glycerides composed of mono-, di- and triglycerides with mono- and diesters of 209 polyethylene glycol. They are amphiphilic with surface active properties.[48] Gelucire 44/14 has 210 a melting temperature of 44°C and a hydrophilic – lipophilic balance of 14, hence its name. It is 211 a compound known for its permeation enhancing properties and is 'generally regarded as safe' 212 (GRAS). Liu et al. [47] investigated Gelucire 44/14 for its permeability enhancing performance 213 in vitro and in vivo for various ophthalmic drugs and demonstrated that it enhanced transcorneal 214 permeability of drugs with a range of hydrophilicity / lipophilicity whilst remaining non-215 irritating. Loftsson and Stefansson [33] reviewed cyclodextrins for enhanced topical delivery of 216 steroids for ophthalmic formulation and the cyclodextrin-drug complexes were found to be well 217 tolerated in eye drop formulations. Cyclodextrins and their drug complexes are too large to 218 partition into the cornea and until recently it was generally thought that they kept the drug in 219 solution at the eye surface where the drug was able to diffuse into the tissue, [47,49] or by

modulation of the aqueous diffusion layer on the corneal surface.[50] Morrison *et al.* [34] investigated the use of cyclodextrins as ocular drug delivery excipients for permeability enhancement of riboflavin for the treatment of keratoconus. They have shown that cyclodextrin forms complexes with riboflavin and release their drug payload by preferential take up of cholesterol from corneal epithelial cell membranes. The removal of cholesterol renders the epithelium permeable, allowing enhanced drug penetration. **Figure 4** shows β -cyclodextrin induced histological changes to the epithelium of bovine corneas (b,d,f), compared to those without cyclodextrin exposure (a,c,e). β -Cyclodextrin induced loosening of the epithelium appears to increase with exposure time of 15, 45 and 75 minutes (b,d,f respectively), and this correlates with increased riboflavin penetration without complete destruction of this barrier.

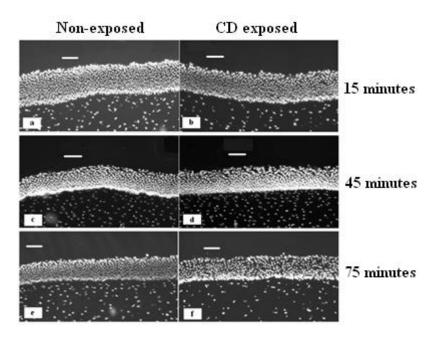


Figure 4. Micrographs of bovine cornea cross-sections showing differences between areas that were exposed to β-cyclodextrin (b,d,f) or not (a,c,e), at 15, 45 and 75 minutes. Scale bar = 100 μ m. Adapted with permission from: Morrison *et al.*[34] Cyclodextrin-mediated enhancement of riboflavin solubility and corneal permeability. *Molecular Pharmaceutics*. 10, 756-762 (2013).

Retention strategies:

Pre-corneal losses have a major impact on ocular drug delivery; it follows that if the drug formulation stays in contact with the intended tissue for longer it is more likely to penetrate the target site to afford its desired action. Adopting an approach for formulation retention is one way to minimize this problem and this can be achieved by several means. Various retention approaches will be discussed in the following section:

Viscosity enhancing polymers;

Natural and synthetic polymers prove useful for their viscosity enhancing properties in ocular drug formulations for improving residence time. These materials absorb water to form viscoelastic gels which prove to be suitable vehicles for drug delivery, and they include derivatives of cellulose, poly(vinyl alcohol), poly(vinyl pyrrolidone), carbomers (weakly crosslinked poly(acrylic acids)), and the natural mucopolysaccharide; hyaluronic acid, a component of the vitreous humour.[51,52] Mechanisms for release of incorporated drugs are determined by their chemical structure, network arrangement and swelling properties.[53] Ocular drug delivery formulations incorporating viscosity enhancing polymers resist lacrimal drainage when residing in the lower conjunctival cul-de-sac. However, disadvantages with this approach are an initial blurring of vision due to changes in refractive index at the corneal surface, and difficulty instilling a precise dose.[24,54,71]

In situ gelling systems;

'In situ' gelling systems undergo phase transition from liquid to gel under physiological conditions and this technique has advantage over the simpler viscosity enhancing systems. Phase transition can be mediated by physiological temperature, pH or electrolyte composition at the cornea surface.

Thermogelling systems include polaxomers, [55,56] pluronics and tetronics, [57]. Ur-Rehman et al. [58] investigated combined formulations of polaxamer 407 with chitosan as thermogelling delivery systems for ocular, vaginal, orthodontal and parenteral drug administration; this process allowed site specific tunable drug delivery with enhanced gel strength and mucoadhesive properties. Gratieri et al. [59,60] also worked with polaxamer/chitosan gel forming systems, their aim was to develop phase transition gels with improved mechanical and mucoadhesive properties. They investigated poly(ethylene oxide) – poly(propylene oxide) - poly(ethylene oxide) triblock polymers (PEO-PPO-PEO) with chitosan of various polymer ratios and found that the polymer/chitosan ratio of 16:1 w/w offered optimum gelation temperature of 32°C, good resistance to shearing forces at 35°C and good retention due to mucoadhesion. Poly(Nisopropylacrylamide) is a well-researched thermogelling polymer with a lower critical solution temperature (LCST) of 32°C, an ideal temperature for thermosensitive applications for ocular drug delivery, although the polymer precipitates above the LCST forming a stiff gel which can be uncomfortable for ocular drug delivery applications.[61] It also shows reduced transparency above LCST,[62] which would be undesirable for eye-drop formulations. Cao et al.[61] investigated thermogelling poly(N-isopropylacrylamide)-chitosan formulation and found it to be a suitable system for ocular delivery of water-soluble drugs, but it is not clear whether they have solved the 'reduced transparency' issue with their development. Mayol et al. [56] investigated thermogelling polaxamers (F127 and F68) and found that on their own their gelling properties were not ideal but could be optimized by addition of the naturally occurring mucoadhesive polysaccharide, hyaluronic acid. They consider that this approach can be exploited for a range of sustained drug delivery scenarios and they are especially suited for ocular drug delivery. PHmediated systems include Carbopol® [63] and cellulose acetate phthalate. [64] Electrolyte

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triggered gelling systems make the transition from liquid to gel by induction of crosslinking in the gelling system mediated by cations present in the tear fluid, and these include gellan gum (Gelrite®), carrageenan,[65-67] and sodium alginate.[68]

Mucoadhesives;

- Mucoadhesion is the interaction between a compound, usually a polymer, natural or synthetic, with mucosa or associated mucus.[53,69] Mucoadhesive drug delivery depends on the interplay between the dosage form and mucus covered mucosal epithelial membranes, residence time increases due to this interaction, allowing more time for the drug to penetrate its intended site of action.[69,70] Mucosal adhesion of dosage forms can be explained using a combination of theories:[71,72]
 - *Electronic theory*, where interaction is due to electron transfer between the dosage form and mucosal surface.
 - Adsorption theory, attraction mechanisms are via electrostatic effects, hydrogen bonds
 and Van der Waals forces. Hydrophobic effects are also implicated, more so when the
 mucoadhesive polymers are amphiphilic. Covalent bonding can also come into effect
 between some specific polymers and mucins.
 - *Wetting theory*, mostly applies to liquid mucoadhesives where there are structural similarities between the polymer and mucin, these effects reduce surface tension and allow the mucoadhesive polymer to spread on the mucosal surface.
 - *Diffusion theory*, considers the interpenetration of polymer into the mucus and diffusion of soluble mucins into the mucoadhesive.

Neither of the above mentioned theories can be used to explain mucoadhesion on their own, more, they each play a part to varying degrees within any given scenario.[71-74] In considering a typical series of events involving a mucoadhesive – mucosa interaction; first of all the *wetting theory* comes into play with wetting and associated swelling of the dosage form; next physical interactions involving *electronic and adsorption theories* take place forming non-covalent bonds between the system components; *diffusion theory* then comes into play when further non-covalent bonds during interpenetration of polymer-protein chains during which physical and covalent (chemical) bonds form again involving *electronic and adsorption theories*.[71,72]

With traditional ocular drug delivery systems residence time is determined by tear turnover, but for mucoadhesive systems this becomes governed by mucus turnover, hence drug retention and bioavailability is substantially increased.[51] Mucoadhesive polymer films could potentially provide a suitable platform to deliver ocular drugs, Khutoryanskaya et al. [75] investigated the use of complexes and blends of poly(acrylic acid) (PAA) and methylcellulose (MC) to produce polymeric films as vehicles for ocular drug delivery. PAA has excellent mucoadhesive properties due to an ability to form hydrogen bonds with mucin, although it has limited application for transmucosal drug delivery due to being very hydrophilic, thus quick dissolving; it also has poor mechanical properties and can cause irritation to delicate mucosa. MC has favourable properties that are applied in transmucosal delivery systems; it has excellent biocompatibility profiles but has poor mucoadhesive properties. The researchers used a polymer blend approach with different combinations of PAA / MC under a range of pH and optimized a formulation bringing together the favourable properties of both polymers. *In vitro* studies of drug-loaded polymer films determined their release profiles and they found that films enriched in MC had significantly slower drug release profiles than films enriched in PAA. This could allow a tunable drug

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delivery system depending on whether rapid or sustained release is required. They further investigated *in vivo* retention of the polymer films using rabbits and found that 100% MC films were retained for up to 50 minutes but successful application was hampered by poor mucoadhesive properties. 100% PAA films were strongly mucoadhesive but retention was poor due to quick dissolution. They concluded that polymer blends had good bioadhesive qualities and showed better retention of 30-60 minutes compared to the films composed of individual polymers. [75]

Nanoparticles;

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Nanoparticle drug delivery systems are more generally described as submicron sized structures; these systems were described by Nagarwal et al.[19] as 10 to 1000 nm particles in which drugs could be loaded by attachment to the matrix or dissolved within, encapsulated or entrapped within the structure giving a versatile drug delivery system. Hans and Lowman [76] discuss biodegradable polymeric nanoparticles for drug delivery, they suggest that surface modified biodegradable solid nanoparticles have an advantage regarding controlled release, principally for targeted drug delivery for the treatment of specific organs, in particular for extended drug delivery to the cornea and conjunctiva.[76] Ibrahim et al.[77] describe a mucoadhesive nanoparticle system as a carrier for gatafloxacin/prednisolone biotherapy for treatment of bacterial keratitis, a serious corneal condition which could lead to blindness without rapid and appropriate intervention. The drug loaded nanoparticle systems they describe were produced from Eudragit® RS 100 and RL 100 and were coated with the bioadhesive polymer hyaluronic acid. Nanoparticles within the suspensions produced using these systems were in the range of 315 nm to 973 nm. For ocular drug delivery, supramolecular structures, complexes and composites belong to nanoparticulate systems and these can include microemulsions, liposomes,

niosomes, dendrimers and cyclodextrins.[1,2,36-41] Kassam et al.[78] investigated the use of nanosuspensions for ophthalmic delivery of three virtually insoluble glucocorticoid drugs in aqueous media; hydrocortisone, prednisolone and dexamethasone. Their findings show an enhancement to the rate and extent of ophthalmic drug absorption together with improved drug performance compared with aqueous solutions and microcrystalline suspensions. De Campos et al.[79] investigated the interaction of poly(ethylene glycol)- or chitosan- coated colloidal nanocapsules with ocular mucosa; they conclude from ex vivo studies that the systems they developed enhanced permeation of dye through the cornea. Evidence from confocal microscopy shows their systems penetrated the epithelium of rabbit cornea via the transcellular pathway and they found that PEG-coated colloids had an enhanced rate of transport across the whole epithelium; whilst chitosan-coated nanocapsules were retained in the superficial epithelial layers. They suggest these systems could be designed as colloidal drug carriers targeting a specific purpose, that is, to attach to the cornea or penetrate into or through it. This implies these systems should prove useful of treating conditions of the cornea and deeper structures within the eye. Diseases of the posterior section of the eye include macular degeneration, diabetic retinopathy, retinitis pigmentosa and related ocular neovascular disease. Topical delivery of drugs to the posterior section of the eye is particularly challenging due not least to ocular barrier function and internal clearance mechanisms within the anterior chamber. Recent developments in the field of nanoparticles involve submicron-sized liposomes (ssLips) and these are proving useful for topical drug delivery systems in the form of eye drops for the treatment of posterior segment diseases. Studies by Hironaka et al. and Inikuchi et al. [80,81] show successful delivery of coumarin-6 to the retina via non-corneal and non-systemic pathways using eye drops. The

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assumption can be made that posterior section delivery is via penetration through the sclera using ssLips [8,41] (emphasis highlights conclusion of the authors of this review).

Ocular inserts:

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Ocular inserts are drug loaded devices placed in the upper or lower cul-de-sac and in some cases, directly on the cornea; their purpose is to act as a controlled release drug reservoir. These systems can be insoluble devices that need to be removed after a given period of time or they can be designed to dissolve, erode or biodegrade at the ocular surface. Early forms of ocular inserts have been used since the middle ages and were given the arabic term *al-kohl*. By the nineteenth century, paper patches soaked with drug solutions were used and in the early twentieth century glycerinated gelatin systems were in use [82] It is not clear how effective these early devices were, however, drug delivery by this means has developed and devices can be of soluble ophthalmic drug inserts (SODI) or insoluble polymers, mucoadhesives or soluble natural materials such as collagen (e.g. from porcine sclera).[4] Ideally these devices could be applied and left in place with no further intervention thereafter. Ocular inserts need to be discreet and comfortable to gain patient acceptance. Sustained release ophthalmic inserts are defined as sterile devices which can be drug impregnated thin, single or multi-layered films, solid or semisolid materials. The objective being to extend ocular contact time thus improving bioavailability. Development of ocular inserts that bring reliable controlled release drug delivery and patient comfort offers a considerable challenge. The main classes of devices are insoluble, soluble and biodegradable inserts.[83] Ocusert® was the first relatively successful product for delivery of pilocarpine for the treatment of ocular hypertension and has been commercialised since 1974. Ocusert® consists of a pilocarpine-alginate reservoir sandwiched between thin ethylene-vinyl acetate films, the devices are designed to deliver pilocarpine at either 20µg per

hour or 40 µg per hour. Some disadvantages of this system were unreliable control of intraocular pressure, leakage, folding, difficulty inserting the devices and ejection or irritation.[82,84] Ocufit SR® are sustained release rod shaped devices made from silicone elastomer, designed to reside in the lower conjunctival fornix; these devices are well tolerated and expulsion is significantly less than with oval or flat inserts. Minidisc ocular therapeutic system (OTS) by Bausch & Lomb are drug-loaded polymer discs with similar shape as contact lenses but are smaller (4-5 mm); they were designed to reside on the sclera in the upper or lower fornix and deliver the antibiotics gentamicin or sulfisoxazole between 3-14 days depending on the system. The company produces non-erodible hydrophobic and hydrophilic systems and erodible devices based on hydroxypropyl cellulose. The inserts are comfortable and easy to use for most patients. Smith & Nephew Pharmaceutical Ltd patented what they term 'new ophthalmic delivery system' (NODS®); these devices offer precision pilocarpine delivery for glaucoma patients from poly(vinyl alcohol) (PVA) film flags. These devices attach to the mucosal surface of the lower conjunctival sac where it takes up fluid from the tears, swells and delivers its drug payload at a pre-determined rate into the lacrimal fluid as it slowly dissolves.[82] Mydriasert® are insoluble devices marketed by IOLTech for the delivery of phenylephrine and tropicamide to induce sustained mydriasis during surgery or for examination of the fundus (interior ocular surface).[3] Human amniotic membrane has been used for corneal transplant to treat corneal disorders and ulcerative ocular conditions. Resch et al. [85,86] investigated its use as drug loaded ocular devices to deliver ofloxacin in vitro and they concluded that single layer human amniotic membrane had a significant reservoir capacity capable of delivering the drug for up to 7 hours in vitro. They propose that drug pretreatment of amniotic membrane could be beneficial when using

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this tissue for ocular transplant when treating infectious keratitis.[85,86] **Table 1** lists some advantages and disadvantages for using ocular inserts. [20,82,87]

Table 1. Advantages and disadvantages using ocular inserts. **Disadvantages Advantages** Increased residence time / bioavailability Physical and psychological obstacles of placing solid objects on the eye, foreign Precision dosing with controlled release, body sensation avoids pulsate drug delivery Movement around the eye could interfere Minimal systemic absorption with vision Administration frequency reduced Potential accidental loss Conjunctival / scleral route to internal Some devices difficult to insert or target remove Better shelf life and no preservatives Potential burst release upon insertion Combinational therapeutic approaches prior to controlled delivery

Recent developments in ocular insert drug delivery systems:

Colo *et al.* [88] investigated the effect of adding chitosan hydrochloride (CH-HCl) to mucoadhesive erodible ocular inserts produced from poly(ethylene oxide) (PEO) of various molecular weight for delivery of ofloxacin. They added 10, 20 and 30 % medicated CH-HCl microparticles to PEO formulations made from 900 kDa or 2000 kDa. Erosion of the devices was accelerated proportional to CH-HCl content. The lower molecular weight PEO proved more suitable for prolonged drug release. They conclude that inclusion of CH-HCl in the devices aids erosion and enhances corneal permeability of ofloxacin when compared to devices not containing CH-HCl. Hornof *et al.* [89] developed mucoadhesive devices based on thiolated poly(acrylic acid) (PAA) and these were evaluated in human in vivo studies. Their aim was to

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develop mucoadhesive ocular inserts for controlled delivery of ophthalmic drugs using fluorescein as a fluorescent tracer to determine release rates from the devices in humans. They compared mean fluorescein concentrations in the tear film and cornea as a function of time after instillation of eye drops and inserts composed of thiolated and unmodified PAA. The thiolated polymer inserts formed a soft, insoluble hydrogel and were well tolerated by volunteers. Their findings show this material offers a promising platform for ocular drug delivery for a prolonged duration. Mishra and Gilhotra [63] designed and characterized a bioadhesive in-situ gelling ocular insert for the delivery of gatifloxacin using a mixture of sodium alginate with chitosan, which was plasticized with glycerin. They combined sodium alginate for its gelling properties, with chitosan for its bioadhesive qualities, formulations of various proportions were prepared and films were produced using the solvent casting technique as described by Pandit et al. [90] Using this system they found an accumulative drug release of 95-99% during 8-12 hours and the formulation consisting of 2% alginate with 1% chitosan had the most sustained release of 12 hours. They conclude that this system allowed production of uniform in situ gelling polymer films suitable for controlled release of gatifloxacin for the treatment of bacterial keratitis and conjunctivitis.[63] Natamycin is a polyene antibiotic used for the treatment of fungal blepharitis, bacterial keratitis and conjunctivitis and it has the ability to reduce intraocular pressure. Rajasekaran et al. [91] compared the controlled release performance of natamycin from ocular inserts they designed from a variety of polymeric materials; Eudragit® L-100, S-100, RL-100, hydroxypropyl methyl cellulose phthalate (HMCP) and cellulose acetate phthalate (CAP) in different proportions with poly(ethylene glycol-400) (PEG-400) as a plasticizer. Their aim was to develop devices for in situ sustained drug delivery and their approach was to prepare polymeric films using the solvent casting method. 1 cm discs were cut from the films to be used

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as inserts; these were evaluated for their physicochemical properties such as drug concentration, weight, folding durability, thickness, moisture absorption and vapour transmission rate. FTIR studies established that there was no chemical interaction between the drug and polymers used. *In vitro* studies were conducted to determine their drug release kinetics; devices made from CAP, HPMCP and Eudragit® S-100 released all of their drug payload within 10-15 hours, whilst inserts made from increased concentrations of Eudragit® RL-100 continued release for 18-23 hours; best performance was shown for formulations consisting of 3% Eudragit® RL-100 and 1% Eudragit® L-100. They conclude that nataycin loaded ocular inserts produced from 3% Eudragit® RL-100 and 1% Eudragit® L-100 plasticised with 33% PEG-400 are capable of controlled drug delivery up to 23 hours.

Contact lenses for drug delivery

Contact lenses are hard or soft polymeric devices designed to fit directly onto the cornea to correct refractive abnormalities; they can be produced from hydrophilic or hydrophobic polymers. Hydrogel contact lenses are realistic products to act as ocular drug delivery systems; they are able to imbibe a large volume of aqueous solution relative to their anhydrous form. If the aqueous solution that hydrates the contact lens contains sufficient pharmaceutically active material this will be able to diffuse from the polymer matrix into the tear film bathing the eye and subsequently interact with the ocular tissue. However, there still remains a need to retain the drug within the devices sufficiently to provide sustained release.

The idea of using hydrogel contact lenses as drug delivery devices was first suggested by Wichterle *et al.* [29,92] in their 1965 patent, in which they suggest the inclusion of medication upon lens hydration to offer extended drug availability during wear. Contact lens design determines how they are to be used; daily, weekly and monthly disposable options are

available.[92] Early approaches to contact lens aided drug delivery relied on absorbance of drug loaded solution during pre-wear soaking. Conventional contact lenses have limited drug loading potential and drug delivery using this method proves unreliable, giving an initial 'burst release' followed by rapid decline over a relatively short period. [20,93] Other methodologies include molecular imprinting technology, drug loaded coating or addition of a sandwhich layer of drugloaded polymer, inclusion of drug-loaded nanoparticles and cyclodextrin grafting.[28] Molecular imprinting technology is a technique whereby the polymer formulation is modified to give it a higher affinity towards drug molecules, thus increasing their drug loading potential and prolonging delivery [94-96]. Hiratani et al. [93] took this approach in developing a system employing methacrylic acid, N,N-diethylacrylamide and the drug timolol; from this system they were able to achieve sustained timolol release for almost 48 hours in vitro. Alvarez-Lorenzo et al. [97] applied the same strategy to produce norfloxacin-loaded poly(hydroxyethyl methacrylate) contact lenses and they report that reservoir capacity was enhanced by up to 300 fold compared with pHEMA lenses without molecular imprinting technology. Hyatt et al. [98] investigated the release profiles of gentamicin and vancomycin from fibrin coated and fibrin sandwiched contact lenses in vitro; their aim was to develop a system that could offer controlled and sustained drug delivery for a minimum period of 8 hours. They conclude that the fibrin gel/lens systems performed better for extended delivery of gentamicin compared to normal lenses soaked with the antibiotic solution, however, their performance for delivering vancomycin was poor compared to soaked lenses. Lenses incorporating fibrin showed potential for treating microbial keratitis. Ciolino et al. [99,100] investigated poly(lactic-co-glycolic acid) (PLGA) coatings and sandwiched films with contact lenses as potential drug delivery devices. They found that contact lenses incorporating PLGA film retained antifungal properties up to 3 weeks in vitro,

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and their prototype ciprofloxacin eluting contact lens demonstrated controlled release at therapeutically active concentrations for up to 4 weeks in vitro. Although fibrin or PLGA film sandwiched and coated lenses bring sustained drug delivery benefits, the lenses are opaque; therefore they require clear 'window' in the centre of the lens allowing the patient to see during treatment.[97-100] Inclusion of drug loaded nanoparticles within the polymer matrix of contact lens is an effective strategy for prolonged drug delivery. This approach can allow sustained release which can be tuned towards the patient's needs, anything between a few hours to several weeks. Gulsen and Chauhan [101] conducted a pilot study to determine the effectiveness of nanoparticle laden pHEMA. The nanoparticles were based on oil-in-water microemulsion loaded with lidocaine, a hydrophobic drug; the droplets were then encapsulated in a silica shell which stabilized the nanoparticles and these were incorporated in the hydrogel matrix during polymerization. Hydrophobic lidocaine has a slight and finite solubility in water; therefore it is able to slowly diffuse from the nanoparticles into the aqueous phase of the gel matrix where it would then be able to further diffuse into the tear film. The nanoparticle-laden hydrogels remained clear and drug release studies in vitro showed an initial burst release followed by slow and steady release thereafter; by day 10 virtually all the drug had been released. They conclude that the nanoparticle-loaded hydrogels could be suitable for controlled drug delivery for several days at therapeutically effective concentrations. Gulsen and Chauhan [102] followed up their previous investigation of nanoparticle-laden pHEMA by developing four more microemulsion based formulations, type 1 and 2 were based on canola oil with Tween® 80 and Panadon SDK, with or without a stabilizing silica shell, and type 3 and 4 were based on hexadecane with Brij® 97 with or without a stabilizing silica shell; they incorporated lidocaine as a model drug. Type 1 formulation was opaque due to the poor solubility of Tween® 80 in HEMA, type 2 formulation

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lost some transparency but was not opaque indicating that the silica shell reduced interaction between the surfactant and HEMA. Type 3 showed minimal transparency reduction but was not as transparent as pHEMA, type 4 showed no observable loss of transparency due to stabilization afforded by the silica shell. Release studies in vitro determined that formulations based on hexadecane with Brij® 97 were suitable for sustained drug delivery at therapeutic rates for up to 8 days, Tween®80 based formulation was deemed unsuitable due to poor stability and particle aggregation. Gulsen and Chauhan speculate that furthering this work to develop 'smart' particulate based systems which could respond to pH or temperature change could minimise burst release and decaying release rates.[101,102] The approach followed by Jung and Chauhan [103] was to develop a timolol loaded nanoparticle / HEMA based contact lens system. Their aim was to produce nanoparticles without using surfactant due to opacity issues when these are used with HEMA. Using thermal polymerization techniques they formed drug loaded nanoparticles based on crosslinking monomers; propoxylated glycerol triacrylate (PGT) and ethylene glycol dimethacrylate (EGDMA) and incorporated these in pHEMA hydrogels. Their product was a transparent drug loaded hydrogel with temperature dependent release rates between 2-4 weeks. They conclude their system maintains drug stability under refrigerated conditions and the temperature change promotes drug release upon insertion of the lenses into the eyes. Figure 5 shows how nanoparticles could release entrapped drug molecules into the preand post-tear films.

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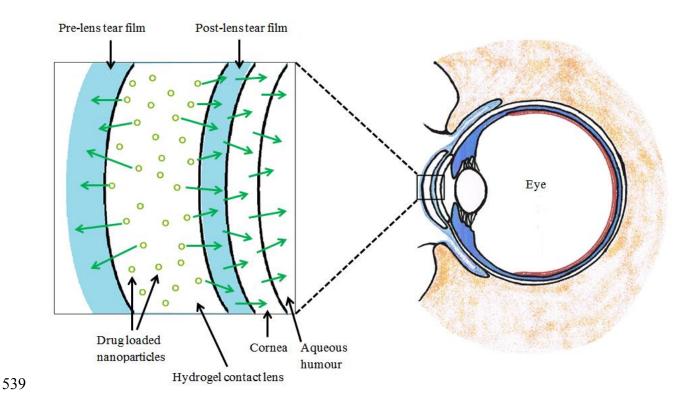


Figure 5. Drug diffusion from nanoparticles encapsulated within hydrogel contact lens. The scale used in this image has been exaggerated for clarity.

Drug loading capacity of hydrogel contact lenses can be enhanced by the inclusion of 'container molecules'. Cyclodextrins, with their 'guest-host' properties have been investigated for this purpose. Complexation between cyclodextrins and drug molecules is a dynamic process due to the weak non-covalent interactions in play. The strategy followed by dos Santos *et al.*[104] was to synthesise methacrylated β -cyclodextrin and use it to form co-polymer with HEMA and EGDMA, the polymers formed had clear gel properties. Drug loading was achieved by soaking the anhydrous polymers in solutions of acetazolamide or hydrocortisone for 4 days. The performance of these methacrylated β -cyclodextrin hydrogels was studied *in vitro* and they were found to offer tunable drug loading/release rates with capacity for sustained drug delivery over several days. They followed up this study with development of another hydrogel formulation

using β -cyclodextrin grafted onto pHEMA-co-GMA (glycidyl methacrylate). This system was able to enhance diclofenac loading by 1300% and could sustain drug release for 2 weeks in lacrimal fluid. They conclude that these systems could have potential for pharmaceutical applications in soft contact lenses and other medicated devices.[105] Xu *et al.*[106] produced hydrogel films and contact lenses from HEMA, mono-methacrylated β -cyclodextrin and trimethylolpropane trimethacrylate. Puerarin was incorporated as a model drug by soaking in drug solution to hydrate the gel. *In vitro* studies determined loading and release rates were dependent on β -cyclodextrin content. *In vivo* studies using rabbits showed the gels offered sustained drug release with superior performance compared to commercial puerarin eyedrops. The devices had excellent mechanical properties and the researchers propose the material is suitable for drug delivery from re-usable daily wear contact lenses.

Ocular implants:

Treating the posterior segment

Historically, the posterior segment has been exceptionally difficult to treat due to the many barriers that obstruct ingress of foreign matter into the eye. The development of ocular implants have allowed these external barriers to be overcome. Modern devices allow long term treatments for otherwise impossible to treat conditions, many devices provide medication for years from a single procedure. [107,112]

Drug eluting intraocular lenses

Intraocular lens (IOL) surgery is a well-established and safe procedure routinely performed worldwide; however as with any surgical technique there is always risk from infection or other

complications, for example, postoperative inflammation, posterior capsule opacification (PCO) and secondary cataracts caused by epithelial cell adhesion and proliferation in the posterior lens capsule. Introduction of preventative medication during surgery is subject to decay or elimination before it can be effective. Much research is currently carried out for development of drug eluting IOL's to minimise postoperative problems, and also to address concurrent pathologies. IOL / drug combinations can be achieved by pre-insertion soaking in concentrated drug solution (only useful for drugs with a high affinity for the polymer), coating with layers of drug/polymer, chemical grafting of drugs, drug impregnation using super critical fluids and attaching inserts onto the haptics (the 'arms' of the IOL).[28] A study by Kleinmann et al.[113] determined that commercial hydrophilic acrylic lenses (C-flex, Rayner intraocular lenses) [114] have affinity for fourth generation fluoroquinolones and were able to release this drug above the minimum inhibitory concentration in rabbits for at least 12 hours. They conclude C-flex/drug combination is safe and effective for delivery of these antibiotics. Davis et al.[115] investigated concentrations of 4 antibiotics (moxifloxacin, gatifloxacin, linezolid and ceruroxime) in aqueous and vitreous humour samples from rabbit eyes. Drug released from implanted hydrophilic IOL's was analysed using HPLC to determine drug concentration in the ocular fluid samples. The IOL's used were STAAR Nanoflextm Colamer®, 40% water content material comprised of a collagen, pHEMA blend,[116] pre-soaked in antibiotic solution. Ocular fluid samples were taken for analysis at intervals up to 24 hours. It was established that the antibiotics studied were above the minimum inhibitory concentration in the aqueous humour for at least 6 hours, notably, gatifloxacin concentrations remained above this level at 24 hours after implantation.[116] Layer-by-layer deposition is a technique used for coating opposing charge polymers to rigid hydrophobic IOL's, a drug can be incorporated during this process. Coating pHEMA based

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hydrophilic IOL's by immersion in octadecyl isocyanate can be an effective method to give controlled release from norfloxacin containing IOL's. Grafting drug molecules onto the IOL surface can provide a permanently active surface to prevent cell adhesion, or allow release of drugs by some external trigger, for example light irradiation. High drug concentrations within a polymeric matrix can be achieved using supercritical CO₂ as a means to force drugs into the polymer without the need for organic solvent.[28] Duarte et al.[117] employed supercritical CO₂ technology to impregnate p(MMA-EHA-EGDMA), a suitable polymer for IOL manufacture, with flurbiprofen, an anti-inflammatory drug used for intraocular delivery. Their experiments found the process allowed higher drug impregnation and release studies showed the system to be effective for up to 3 months. The approach employed by Garty et al. [27] was to produce norfloxacin loaded pHEMA cylinders in 1.0 mm diameter microglass tubes with 0.09 mm stainless steel wire through the centre during room temperature polymerization. When fully polymerized the hydrogel was ejected from the tube and the wire removed leaving a tubular hydrogel structure, this was washed with sterilized water to remove unreacted components. The gel was cut into 1.0 mm lengths and lyophilized. Next they added a hydrophobic coating using octadecyl isocyanate to control drug release. The devices were used as sleeves placed over IOL haptics and this assembly was used in lens replacement procedures in the rabbit model. Results from in vivo studies showed the devices offered sustained drug delivery above the minimum inhibitory concentration for over 4 weeks. They conclude that these controlled release devices are effective at sustained delivery of therapeutic levels of drugs within the anterior chamber post operatively. Incorporation of drugs with IOL's has predominantly aimed at postoperative delivery of antibiotics and anti-inflammatory medication.

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Drug delivery by intravitreal injection

There are many debilitating and sight threatening conditions resulting from posterior segment diseases and in most cases the only way these can be treated is by invasive procedures, for example 'intravitreal injection'. In the main this still remains so, however, developments have brought a diverse range of effective implantable drug delivery systems targeting posterior segment disease and the various options will now be considered. [22] The most common means to place drugs in the posterior chamber employs injection into the vitreous humour; this provides a high concentration of drug where it is needed and minimises systemic complications. Xu *et al.* investigated the diffusion of polystyrene nanoparticles of various size and surface chemistries in fresh bovine vitreous and they were able to achieve tuneable drug transport within the posterior chamber depending the designed properties of the nanoparticle [118]. However, many conditions require repeated treatment and this can cause intraocular problems, for example, cataract, retinal detachment, haemorrhage, endophthalmitis and ocular hypertension.

Intraocular implants

In an attempt to overcome the problem of frequent injections biodegradable and non-biodegradable drug depot devices which can offer long term drug release into the posterior chamber have been developed and further research in this area is ongoing. Solutions, liposomes, micelles, nanoparticles and vectosomes are suitable for intravitreal injection although these dosage forms only give short term drug availability, generally days to several weeks.[23,119] Biodegradable and non-biodegradable drug depot devices have been developed and further research in this area is ongoing. Implantable devices for long term drug delivery are on the market or currently undergoing clinical trial. Vitrasert® is a drug depot device for sustained delivery of ganciclovir via a rate limiting poly(vinyl acetate)/ethylene vinyl acetate (PVW/EVA)

membrane for up to 8 months.[22,119,120] Retisert® intraocular inserts were approved by the FDA in 2005. They are inserts for delivery of the corticosteroid, fluocinolone acetonide for treatment of posterior uveitis, a serious sight threatening condition. The devices are designed for long term drug release up to 30 months.[121] Vitrisert® and Retisert® inserts are nondegradable and require surgical implantation and removal.[22] Medidur® are implantable devices for delivering fluocinolone acetonide for up to 36 months. This device consists of a narrow cylindrical polyimide tube loaded with the drug and PVA-based end caps provide rate limiting drug delivery. The 3.5 mm long device is inserted through a 25-g needle carried out under local anaesthesia and creates a self-healing wound eliminating the need for surgery.[122] Implants employing biodegradable polymers are promising systems for intraocular drug delivery. Sivaprasad et al. [123] report the use of the Posurdex® biodegradable polymer device for treatment of macula oedema using dexamethasone. This drug has a half-life of less than 24 hours therefore it provides only limiting management of this condition by injecting the drug. However, dexamethasone containing Posurdex® devices were shown to deliver the drug at a constant rate for up to 4 months, these devices have been re-named Ozurdex® and are marketed by Allergan Inc. [124] In vivo studies using monkeys showed the system was effective at reducing retinal vasculopathy and neuropathy.[125] Surodex® is a poly(lactic-glycolic acid) device to be inserted in the anterior or posterior chamber at the time of cataract surgery to deliver dexamethasone for up to 10 days. Tan et al. [126] conducted a randomized clinical trial to evaluate the effectiveness of the Surodex® insert as a safe and effective treatment of intraocular inflammation in post-cataract surgery. Their study employed flare meter readings to determine inflammation and this showed that measured values were lower in all readings from the Surodex® group compared to those treated post operatively with dexamethasone eye drops, they

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conclude that implantation of a single Surodex® device at the time of cataract surgery reduces post-surgery inflammation [126,127].

Future perspectives:

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In this review the various strategies for enhancing bioavailability of ophthalmic drugs have been considered; how drug bioavailability can be improved using solubility, retention and permeability enhancers has been explored. Drug loaded contact lenses allow localised delivery directly to the cornea, where the lenses offer controlled release whilst isolating the post corneal tear film from lachrymal clearance. Nanoparticle technology is allowing drug delivery to the posterior chamber via topically applied formulations. Future research is likely to bring discoveries of materials with superior performance compared with those in current use. The use of ocular inserts for extended and intimate contact between the dose form and ocular tissue proves to be a beneficial strategy and the use of ocular implants allows all external barriers to be overcome, giving direct access to internal tissues whilst minimising side effects. Many of these approaches have been developed in recent decades and continue to be improved upon with new innovations. Looking to the future innovative advances to delay or prevent blindness could be made; developments in two main areas could be speculated; the cornea and vitreous humour. First, corneal disease has a major influence on visual health; corneal tissue engineered constructs are being developed to test new ocular drugs. Future development of artificial corneas could become a possibility to replace diseased ones without the need for donor tissue, which is a scarce commodity.[127,128] Another area for advanced drug delivery is the posterior segment; vitrectomy is an invasive but well-established procedure for many posterior segment disorders. A synthetic material is used to replace natural vitreous humour. The possibility of developing synthetic materials for whole or partial vitrectomy as a drug depot could allow long term

controlled release for decades. A one off procedure would be more favourable than many less effective ones over the course of a lifetime.[129,130]

Executive summary:

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Strategies to enhance the bioavailability of drugs are;

Drug solubility and penetration enhancement

- Many ocular drugs have low aqueous solubility; this can be improved using hydrotropic compounds. Formulating for higher drug concentration means increased availability.
- Inclusion of penetration enhancers within a formulation improves drug partitioning into tissue.

Drug retention strategies

- Viscosity enhancing polymers, in situ gels and bioadhesives allow eye drop formulation
 to resist pre-corneal losses and they retain intimate contact with ocular tissue longer
 giving the dose form more time to penetrate ocular membranes.
- Drug delivery from ocular inserts are a means to place the dose form in immediate contact with ocular mucosa, this strategy allows controlled and sustained drug release for an extended period.

Ocular implants

• Implantable devices are designed to penetrate the ocular membranes or reside entirely within the eye. This strategy overcomes all external barriers and can offer short term medication or deliver medication for several years when treating chronic conditions.

707 Future perspectives

• A speculative outlook considered the possibility of innovative technologies developing

synthetic tissues to enable testing new drugs and possibly even produce artificial corneas

for transplant. The idea of developing novel materials for vitreous humour replacement as

lifetime drug delivery depots could potentially become realised.

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- 713 References
- Papers of special note have been highlighted as;
- 715 * of interest
- 716 **of considerable interest
- 1. Tseng CL, Chen KH, Su WY, Lee YH, Wu CC, Lin FH. Cationic gelatin nanoparticles for
- 718 drug delivery to the ocular surface: in vitro and in vivo evaluation. *Journal of Nanomaterials*.
- 719 http://dx.doi.org/10.1155/2013/238351 (2013).
- 720 2. Gupta H, Aqul M, Khar RK, Ali A, Bhatnagar A, Mittal G. Sparfloxacin-loaded PLGA
- nanoparticles for sustained ocular drug delivery. *Nanomedicine*. 6(2), 324-333 (2010).
- 3. Yasukawa T, Tabata Y, Kimura H, Ogura Y. Recent advances in intraocular drug delivery
- 723 systems. Recent Patents on Drug Delivery & Formulation, 5(1), 1-10 (2011).
- 4. Bourlais CL, Acar L, Zia H, Sado PA, Needham T, Leverge R. Ophthalmic drug delivery
- systems recent advances. *Progress in Retinal and Eye Research*, 17(1), 33-58 (1998)
- 5. Abdulrazik M, Beher-Cohen F, Benita S. Drug delivery systems for enhanced ocular
- absorption. In: Enhancement in drug delivery. Touitou E, Barry BW (Eds), CRC Press, Florida,
- 728 USA, 489-525 (2007).

- 729 6. Washington N, Washington C, Wilson CG. Ocular drug delivery. In: Physiological
- 730 Pharmaceutics: Barriers to Drug Absorption, 2nd ed. CRC Press: Florida, USA, 249-270 (2001).
- 731 7. Das S, Suresh PK. Drug delivery to the eye: special reference to nanoparticles. *International*
- 732 *Journal of Drug Delivery*. 2, 12-21 (2010).
- 8. Schoenwald RD, Deshpande GS, Rethwisch DG, Barfknecht CF. Penteration into the anterior
- chamber via the conjunctival/scleral pathway. Journal of Ocular Pharmacology and
- 735 *Therapeutics*. 13(1), 41-59 (1997).
- 9. Chandran S, Roy A, Saha RN. Effect of pH and formulation variables on in vitro
- transcorneal permeability of flurbiprofen: a technical note. AAPS PharmSciTech. 9(3), 1031-
- 738 1037 (2008).
- 739 10. Kumaran KSGA, Karthika K, Padmapreetha J. Comparative review on conventional and
- 740 advanced ocular drug delivery formulations. International Journal of Pharmacy and
- 741 *Pharmaceutical Sciences*. 2(4), 1-5 (2010).
- 742 11. Kaur IP, Batra A. Ocular Penetration Enhancers. In: Enhancement in Drug Delivery.
- Touitou E, and Barry BW. (Eds.) CRC Press, Florida, USA, 527-547 (2007).
- 744 12. Wilson CG, Semenova EM, Hughes PM, Olenik O. Eye structure and physiological
- functions. In: Enhancement in drug delivery. Touitou E, Barry BW (Eds), CRC Press, Florida,
- 746 USA, 473-487 (2007).
- 13. Bron AJ, Tiffany JM, Gouveia SM, Yokoi N, Voon LW. Functional aspects of the tear film
- 748 lipid layer. *Experimental Eye research*. 78(3), 347-360 (2004).
- 749 14. McCulley JP, Shine W. A compositional based model for tear the film lipid layer.
- 750 Transactions of the American Ophthalmological Society. 95, 79-88 (1997).

- 751 15. Wilson CG, Zhu YP, Kumala P, Rao LS, Dhillon B. Ophthalmic drug delivery. In: Drug
- 752 Delivery and Targeting for Pharmacists and Pharmaceutical Scientists. Hillery AM, Lloyd AW,
- 753 Swarbrick J (Eds.) CRS Press, Florida, USA, 2001 329-353 (2001).
- 754 16. Jarvinen T, Jarvinen K. Prodrugs for improved ocular drug delivery. Adv. Drug. Deliver.
- 755 Rev. 19(2), 203-224 (1996).
- 756 17. Morrison PWJ, Khutoryanskiy VV. Enhancement in corneal permeability of riboflavin using
- 757 calcium sequestering compounds. *International Journal of Pharmaceutics*.
- 758 DOI:10.1016/j.ijpharm.2014.06.007 (2014).
- 759 18. Jain-Vakkalagadda B, Pal D, Gunda S, Nashed Y, Ganapathy V, Mitra AK. Identification of
- a Na⁺-Dependent Cationic and Neutral Amino Acid Transporter, B^{0,+}, in Human and Rabbit
- 761 Cornea. *Molecular Pharmaceutics*. 1(5), 338-346 (2004).
- 762 19. Nagarwal RC, Kant S, Singh PN, Maiti P, Pandit JK. Polymeric nanoparticulate system: A
- potential approach for ocular drug delivery. Elsevier, Journal of Controlled Release. 136(1), 2-
- 764 13 (2009).
- 765 20. Kumari A, Sharma PK, Garg VK, Garg G. Ocular inserts Advancement in therapy of eye
- diseases. Journal of Advanced Pharmaceutical Technology and Research. 1(3), 291-296 (2010).
- 767 21. Hu X, Hao L, Wang H, Yang X, Zhang G, Wang G, Zhang X. Hydrogel contact lens for
- 768 extended delivery of ophthalmic drugs. *International Journal of Polymer Science*. 2011, 1-9
- 769 (2011).
- 22. del Amo EM, Urtti A. Current and future ophthalmic drug delivery systems: A shift to the
- 771 posterior segment. *Drug Discovery Today*. 13(3-4), 135-143 (2008).
- 772 23. Thrimawithana TR, Young S, Bunt CR, Green C, Alany RG. Drug delivery to the posterior
- segment of the eye. *Drug Discovery Today*. 16(5-6), 270-277 (2011).

- 774 24. Ding S. Recent developments in ophthalmic drug delivery. Pharmaceutical Science and
- 775 Technology Today, 1(8), 328-335 (1998).
- 776 25. Dong A. Current and potential therapies for ocular neovascular diseases. Clinical and
- 777 Experimental Pharmacology. 3(4), (2013).
- 778 26. Hironaka K, Inokuchi Y, Tozuka Y, Shimazawa M, Hara H, Takeuchi H. Design and
- evaluation of a liposomal delivery system targeting the posterior segment of the eye. Journal of
- 780 *Controlled Release*, 136, 247-253 (2009).
- ** Designed a topical drug delivery system using sub-micron sized liposomes (ss-Lips) as
- 782 carriers targeting the retina from a non-invasive system.
- 783 27. Garty S, Shirakawa R, Warsen A, Anderson EM, Noble ML, Bryers JD, Ratner BD, Shen
- TT. Sustained antibiotic release from an intraocular lens-hydrogel assemble for cataract surgery.
- 785 Investigative Ophthalmology & Visual Science, 52(9) 6109-6116 (2011).
- 786 ** Developed a drug delivery device designed to attach to the haptics of IOL's, potentially
- vseful for inclusion of other drugs for post-surgery
- 788 28. Gonzalez-Chomon C, Concheiro A, Alvarez-Lorenzo C. Drug-eluting intraocular lenses.
- 789 *Materials*, 4, 1927-1940 (2011).
- 790 29. Kim JY, Kim S, Papp M, Park K, Pinal R. Hydrotropic solubilisation of poorly water-soluble
- 791 drugs. Journal of Pharmaceutical Sciences, 99(9), 3953-3965 (2010).
- 792 30. Evstigneev MP, Evstigneev VP, Santiago AA, Davies DB. Effect of a mixture of caffeine
- and nicotinamide on the solubility of vitamin (B₂) in aqueous solution. European Journal of
- 794 *Pharmaceutical Science*. 28(2), 59-66 (2006).
- 795 31. Coffman RE, Kildsig DO. Effect of Nicotinamide and Urea on the Solubility of Riboflavin
- 796 in Various Solvents. Journal of Pharmaceutical Sciences. 85(9), 951-954 (1996).

- 797 32. Coffman RE, Kildsig DO. Hydrotropic solubilisation mechanistic studies. Journal of
- 798 Pharmaceutical Research. 13(10), 1460-1463 (1996).
- 799 33. Loftsson T, Stafansson E. Cyclodextrins in eye drop formulations: enhanced topical delivery
- of corticosteroids to the eye. *Acta Ophthalmologica Scandinavica*, 80, 144-150 (2002).
- 34. Morrison PWJ, Connon CJ, Khutoryanskiy VV. Cyclodextrin-Mediated Enhancement of
- Riboflavin Solubility and Corneal Permeability, *Molecular Pharmaceutics*, 10, 756-762 (2013).
- 803 * Developed methods to enhance drug solubility and corneal permeability. Proposes a
- mechanism for corneal epithelial permeability enhancement using cyclodextrin.
- 805 35. Sahoo SK, Labhasetwar V. Nanotech approaches to drug delivery and imaging. Drug
- 806 Discovery Today, 8(24), 1112-1120 (2003).
- 36. Sahoo SK, Dilnawaz F, Krishnakumar S. Nanotechnology in ocular drug delivery. *Drug*
- 808 *Discovery Today*, 13(3-4), 144-151 (2008).
- 809 37. Gupta H, Aqil M, Khar RK, Bhatnagar A, Mittal G. Nanoparticles laden in situ gel for
- 810 sustained ocular drug delivery. Journal of Pharmacy and BioAllied Sciences. 5(2), 162-165
- 811 (2013).
- 38. Zhou HY, Hao JL, Wang S, Zheng Y, Zhang WS. Nanoparticles in ocular drug delivery.
- 813 International Journal of Ophthalmology. 6(3), 390-396 (2013).
- 39. Lai SK, Wang YY, Hanes J. Mucus-penetrating nanoparticles for drug and gene delivery to
- mucosal tissues. Advanced Drug delivery Reviews. 61(2), 158-171 (2009).
- 816 40. Das S, Suresh PK, Desmukh R. Design of eudragit RL 100 nanoparticles by
- nanoprecipitation method for ocular drug delivery. *Nanomedicine*. 6(2), 318-323 (2010).
- 41. Das S, Suresh PK. Drug delivery to the eye: special reference to nanoparticles. *International*
- 819 *Journal of Drug Delivery*. 2, 12-21 (2010).

- 820 42. Rangel-Yagui CO, Pessoa-Jr A, Tavares LC. Micellar solubilisation of drugs. Journal of
- Pharmacy and Pharmaceutical Sciences, 8(2), 147-163 (2005).
- 822 43. Qu X, Khutoryanskiy VV, Stewart A, Rahman S, Papahadjopoulos-Sternberg B, Dufes C,
- McCarthy D, Wilson CG, Lyons R, Carter KC, Schatzlein A, Uchegbu F. Carbohydrate-based
- micelle clusters which enhance hydrophobic drug bioavailability by up to 1 order of magnitude.
- 825 *Biomacromolecules*, 7(12), 3452-3459 (2006).
- 44. Kulkarni S, Gupta SP, Upmanya N, Tonpay SD. Solubility enhancement of water insoluble
- drug for ophthalmic formulation. *International Journal of Drug Delivery*, 3, 141-148 (2011).
- 45. Chung SH, Lee SK, Cristol SM, Lee ES, Lee DW, Seo KY, Kim EK. Impact of short-term
- 829 exposure of commercial eyedrops preserved with benzalkonium chloride on precorneal mucin.
- 830 *Molecular Vision*. 12, 415-421 (2006).
- 831 46. Burgalassi S, Chetoni P, Monti D, Saettone F. Cytotoxicity of potential ocular permeation
- enhancers evaluated on rabbit and human corneal epithelial cell lines. *Toxicology Letters*. 122, 1-
- 833 8 (2001).
- 47. Liu R, Liu Z, Zhang C, Zhang B. Gelucire44/14 as a novel absorption enhancer for drugs
- with different hydrophobicities: in vitro and in vivo improvement on transcorneal permeation.
- 836 *Journal of Pharmaceutical Sciences*, 100(8), 3186-3195 (2011).
- 837 ** Reveals a non-irritating strategy for permeability enhancement of topically applied
- 838 ocular drug delivery.
- 48. Antunes ABF, De Geest BG, Vervaet C, Remon JP. Gelucire 44/14 based immediate release
- 840 formulations for poorly water-soluble drugs. Drug Development and Industrial Pharmacy,
- 841 39(5), 791-798 (2013).

- 49. Uekama K, Hirayama F, Irie T. Cyclodextrin drug carrier systems. Chemical Reviews. 98,
- 843 2045-2076 (1998).
- 844 50. Tirucherai GS, Mitra AK. Effect of hydroxypropyl beta cyclodextrin complexation on
- aqueous solubility, stability, and corneal permeation of acyl ester prodrugs of ganciclovir. AAPS
- 846 *PharmSciTech*, 4(3), 1-12 (2003).
- 847 51. Kaur IP, Smitha R. Penetration enhancers and ocular bioadhesives: two new avenues for
- ophthalmic drug delivery. Drug Development and Industrial Pharmacy, 28(4), 353-369 (2002).
- 849 52. Shahwal VK. Ocular drug delivery: an overview. International Journal of Biomedical and
- 850 *Advance Research.* 2(5), 167-187 (2011).
- 851 53. Nazar H, Roldo M, Fatouros DG, van der Merwe SM, Tsibouklis J. Hydrogels in mucosal
- 852 delivery. *Therapeutic Delivery*, 3(4), 535-555 (2012).
- 853 54. Shaikh R, Raghu T, Singh R, Garland MJ, Woolfson AD, Donnelly F. Mucoadhesive drug
- delivery systems. *Journal of Pharmacy and BioAllied Sciences*, 3(1), 89-100 (2011).
- 855 55. Miller SC, Donovan MD. Effect of polaxomer 407 gel on miotic activity of pilocarpine
- nitrate in rabbits. *International Journal of Pharmaceutics*, 12(2-3), 147-152 (2004).
- 857 56. Mayol L, Quaglia F, Borzacchiello A, Ambrosio L, Rotonda MIL. A novel
- 858 polaxamers/hyaluronic acid in situ forming hydrogel for drug delivery: rheological,
- 859 mucoadhesive and in vitro release properties. European Journal of Pharmaceutics and
- 860 *Biopharmaceutics*. 70(1), 199-206 (2008).
- 57. Desai SD, Blanchard J, In vitro evaluation of pluronic F127 based controlled release ocular
- delivery systems for pilocarpine. *Journal Pharmaceutical Sciences*. 87(2), 226-230 (1998).

- 58. Ur-Rehman T, Tavelin S, Grobner G. Chitosan *in situ* gelation for improved drug loading
- and retention in polaxamer 407 gels. *International Journal of Pharmaceutics*. 409(1-2), 19-21
- 865 (2011).
- 866 59. Gratieri T, Gelfuso GM, Rocha EM, Sarmento VH, de Freitas O, Lopez RFV. A
- polaxamer/chitosan in situ forming gel with prolonged retention time for ocular delivery.
- European Journal of Pharmaceutics and Biopharmaceutics. 75(2), 186-193 (2010).
- 869 60. Gratieri T, Gelfuso GM, de Freitas O, Rocha EM, Lopez RFV. Enhancing and sustaining the
- 870 topical ocular delivery of fluconazole using chitosan solution and polaxamer/chitosan in situ
- forming gel. European Journal of Pharmaceutics and Biopharmaceutics. 79(2), 320-327 (2011).
- 872 61. Cao Y, Zhang C, Shen W, Cheng Z, Yu L, Ping Q. Poly(*N*-isopropylacrylamide)-chitosan as
- 873 thermosensitive in situ gel-forming system for ocular drug delivery. Journal of controlled
- 874 release. 120(3), 186-194 (2007).
- 875 62. Fujishige S. Phase transition of aqueous solutions of poly(*N*-isopropylmethacrylamide). *The*
- 876 *Journal of Physical Chemistry*, 93(8). 3311-3313 (1989).
- 877 63. Mishra DN, Gilhotra RM. Design and characterization of bioadhesive in-situ gelling ocular
- 878 inserts of gatifloxacin sesquihydrate. Daru Journal of Pharmaceutical Sciences, 16(1), 1-8
- 879 (2008)
- 880 64. Gurny R, Ibrahim H, Buri P. The development and use of in situ formed gels triggered by
- pH. In: Biopharmaceutics of Ocular Drug Delivery. Edman P (Ed), CRC Press, Florida, USA,
- 882 (1993).
- 883 65. Moorhouse R, Colegrove GT, Sandford PA. Baird JK, Kang KS. PS-60: a new gel-forming
- polysaccharide. In: Solution Properties of Polysaccharides. Brand DA (Ed), ACS Symposium
- 885 Series, Washington DC, USA, 111-124 (1981).

- 886 66. Rupenthal ID, Alany RG, Green CR. Ion-activated in situ gelling systems for antisense
- oligodeoxynucleotide delivery to the ocular surface. *Molecular Pharmaceutics*, 8(6), 2282-2290
- 888 (2011).
- 889 67. Sultana Y, Aquil M, Ali A. Ion-activated, Gelrite®-based in situ ophthalmic gels of
- pefloxacin mesylate: comparison with conventional eye drops. Drug Delivery. 13(3) 215-219
- 891 (2006).
- 892 68. Smidsrod O, Skjakbraek G. Alginate as immobilization matrix for cells. Trends in
- 893 Biotechnology. 8, 71-78 (1990).
- 894 69. Yadav VK, Gupta AB, Kumar R, Yadav JS, Kumar B. Mucoadhesive polymers: means of
- improving the mucoadhesive properties of drug delivery system. *Journal of Chemical and*
- 896 *Pharmaceutical Research.* 2(5), 418-432 (2010).
- 897 70. Boddupalli BM, Mohammed ZNK, Nath RA, Banji D. Mucoadhesive drug delivery system:
- 898 An overview. Journal of advanced Pharmaceutical technology and research. 1(4), 381-387
- 899 (2010).
- 900 71. Khutoryanskiy VV. Advances in mucoadhesion and mucoadhesive polymers.
- 901 *Macromolecular Bioscience*, 11(6), 748-764 (2011).
- 902 72. Smart JD. The basics and underlying mechanisms of mucoadhesion. *Advanced Drug*
- 903 Delivery Reviews. 57(11), 1556-1568 (2005).
- 904 73. Baudner BC, O'Hagan DT. Bioadhesive delivery systems for mucosal vaccine delivery.
- 905 *Journal of Drug Targeting*. 18(10), 752-770 (2010).
- 906 74. Vinod KR, Reddy T, Sandhya S, Banji D, Reddy V. Critical review on mucoadhesive drug
- 907 delivery system. Hygeia, Journal for Drugs and Medicines. 4(1), 7-28 (2012).

- 908 75. Khutoryanskaya OV, Morrison PWJ, Seilkanov SK, Mussin MN, Ozmukhametova EK,
- Rakhypbekov TK, Khutoryanskiy VV. Hydrogen-bonded complexes and blends of poly(acrylic
- acid) and methylellulose: Nanoparticles and mucoadhesive films for ocular delivery of
- 911 riboflavin. *Macromolecular Bioscience*. 14(2), 225-234 (2014).
- 912 76. Hans ML, Lowman AM. Biodegradable nanoparticles for drug delivery and targeting.
- 913 Current opinion in Solid State Materials Science. 6, 319-327 (2002).
- 77. Ibrahim HK, El-Leithy IS, Makky AA. Mucoadhesive nanoparticles as carrier systems for
- prolonged ocular delivery of gatifloxacin / prednisolone biotherapy. *Molecular Pharmaceutics*,
- 916 7(2), 576-585 (2010).
- 78. Kassem MA, Rahman AAA, Ghorab MM, Ahmed MB, Khalil RM. Nanosuspension as an
- ophthalmic delivery system for certain glucocorticoid drugs. *International Journal of*
- 919 *Pharmaceutics*. 340(1-2), 126-133 (2007).
- 920 79. De Campos AM, Sanchez A, Gref R, Calvo P, Alonso MJ. The effect of a PEG versus a
- 921 chitosan coating on the interaction of drug colloidal carriers with ocular mucosa. European
- 922 Journal of Pharmaceutical Sciences. 20(1), 73-81 (2003).
- 923 80. Hironaka K, Inokuchi Y, Tozuka Y, Shimazawa M, Hara H, Takeuchi H. Design and
- evaluation of a liposomal delivery system targeting the posterior segment of the eye. *Journal of*
- 925 *Controlled Release.* 136(3), 247-253 (2009).
- 926 81. Inokuchi Y, Hironaka K, Fujisawa T, Tozuka Y, Tsuruma K, Shimazawa M, Takeuchi H,
- 927 Hara, H. Physicochemical properties affecting retinal drug / coumarin-6 delivery from
- 928 nanocarrier systems via eyedrop administration. Investigative Ophthalmology and Visual
- 929 Science. 51(6), 3162-3170 (2010).

- 930 82. Seatone MF, Salminen L. Ocular inserts for topical delivery. *Advanced Drug Delivery*
- 931 Reviews. 16(1), 95-106 (1995).
- 932 83. Rathore KS, Nema RK. Review on ocular inserts. International Journal of PharmTech
- 933 Research. 1(2), 164-169 (2009).
- 84. Kuno N, Fujii S. Recent advances in ocular drug delivery systems. *Polymers*. 3(1), 193-221
- 935 (2011).
- 936 85. Resch MD, Resch BE, Csizmazia E, Imre L, Nemeth J, Revesz P, Csanyi E. Permeability of
- 937 human amniotic membrane to ofloxacin in vitro. Investigative Ophthalmology and Visual
- 938 Science. 51(2), 1024-1027 (2010).
- 939 86. Resch MD, Resch BE, Csizmazia E, Imre L, Nemeth J, Revesz PS, Csanyi E. Drug
- 940 Reservoir Function of Human Amniotic Membrane. Journal of Ocular Pharmacology and
- 941 Therapeutics. 27(4), 323-326 (2011).
- 942 87. Karthikeyen D, Bhowmick M, Pandey VP, Nandhakumar J, Sengottuvelu S, Sonkar S,
- 943 Sivakumar T. The concept of ocular inserts as drug delivery systems: An Overview. Asian
- 944 *Journal of Pharmaceutics*. 2(4), 192-200 (2008).
- 945 88. Colo GD, Zambito Y, Burgalassi S, Serafini A, Seattone MF. Effect of chitosan on in vitro
- 946 release and ocular delivery of ofloxacin from erodible inserts based on poly(ethylene oxide).
- 947 International Journal of Pharmaceutics. 248(1-2), 115-122 (2002).
- 948 89. Hornof M, Weyenberg W, Ludwig A, Bernkop-Schnurch A. Mucoadhesive ocular insert
- based on thiolated poly(acrylic acid): development and in vivo evaluation in humans. *Journal of*
- 950 *Controlled Release.* 89(3), 419-428 (2003).

- ** Developed a well-tollerated cross-linked mucoadhesive drug delivery system based on
- 952 thiolated poly(acrylic acid) which offer sustained drug release with good resistance to
- 953 elimination.
- 954 90. Pandit JK, Harikumar SL, Mishra DN, Balasubramaniam J. Effect of physical cross-linking
- on in vitro and ex vivo permeation of indomethacin from polyvinyl alcohol ocular inserts.
- 956 Indian Journal of Pharmaceutical Sciences. 65(2), 146-151 (2003).
- 957 91. Rajasekaran A, Sivakumar V, Karthika K, Preetha JP, Abirami T. Design and evaluation of
- 958 polymeric controlled release natamycin ocular inserts. Kathmandu University Journal of
- 959 Science, Engineering and Technology. 6(1), 108-115 (2010).
- 960 92. Wichterle O, Lim D. Cross-linked hydrophilic polymers and articles made therefrom. US
- 961 3220960 (1965).
- 962 93. Hiratani H, Alvarez-Lorenzo C. Timolol uptake and release by imprinted soft contact lenses
- made of N,N- diethylacrylamide and methacrylic acid. Journal of Controlled Release. 83(2),
- 964 223-230 (2006).
- 94. White CJ, Byrne ME. Molecularly imprinted therapeutic contact lenses. Expert Opinion on
- 966 Drug Delivery Reviews. 7(6), 765-780 (2010).
- 967 95. Salian VD, Vaughan AD, Byrne ME. The role of living/controlled radical polymerization in
- 968 the formation of improved imprinted polymers. Journal of Molecular Recognition. 25(6), 361–
- 969 369 (2012).
- 970 96. Byrne ME, Park K, Peppas NA. Molecular imprinting within hydrogels. *Advanced Drug*
- 971 Delivery Reviews. 54(1), 149-161 (2002).
- 972 97. Alvarez-Lorenzo C, Yanez F, Barreiro-Iglesias R, Concheiro A. Imprinted soft contact
- lenses as norfloxacin delivery systems. *Journal of Controlled Release*. 113(3), 236-244 (2006).

- 974 98. Hyatt AJT, Rajan MS, Burling K, Ellington MJ, Tassoni A, Martin KR. Release of
- vancomycin and gentamicin from contact lens versus a fibrin coating applied to a contact lens.
- 976 Investigative Ophthalmology and Visual Science. 53(4), 1946-1952 (2012).
- 977 99. Ciolino JB, Hoare TR, Iwata NG, Behlau I, Dohlman CH, Langer R, Kohane DS. A drug-
- 978 eluting contact lens. Investigative Ophthalmology and Visual Science. 50(7), 3346-3352
- 979 (2009).
- 980 100. Ciolino JB, Hudson SP, Mobbs AN, Hoare TR, Iwata NG, Fink GR, Kohane DS. A
- 981 prototype antifungal contact lens. Investigative Ophthalmology and Visual Science. 52(9),
- 982 6286-6291 (2011).
- 983 101. Gulsen D, Chauhan A. Ophthalmic drug delivery through contact lenses. *Investigative*
- 984 *Ophthalmology and Visual Science*. 45(7), 2342-2347 (2004).
- 985 102. Gulsen D, Chauhan A. Dispersion of microemulsion drops in HEMA hydrogel: a potential
- 986 ophthalmic drug delivery vehicle. International Journal of Pharmaceutics. 292(1-2), 95-117
- 987 (2005).
- 988 103. Jung HJ, Chauhan A. Temperature sensitive contact lenses for triggered ophthalmic drug
- 989 delivery. *Biomaterials*. 33(7), 2289-2300 (2012).
- 990 104. dos Santos JFR, Couceiro R, Concheiro A, Torres-Labandeira JJ, Alvarez-Lorenzo C.
- 991 Poly(hydroxyethyl methacrylate-co-methacryalated-β-cyclodextrin) hydrogels: Synthesis,
- 992 cytocompatibility, mechanical properties and drug loading/release properties. Acta
- 993 *Biomaterialia*. 4(3), 745-755 (2008).
- 994 105. dos Santos JFR, Alvarez-Lorenzo C, Silva M, Balsa L, Couceiro J, Torres-Labandeira JJ,
- 995 Concheiro A. Soft contact lenses functionalized with pendant cyclodextrins for controlled drug
- 996 delivery. *Biomaterials*. 30(7), 1348-1355 (2009).

- 997 106. Xu J, Li X, Sun F. Cyclodextrin-containing hydrogels for contact lenses as a platform for
- 998 drug incorporation and release. *Acta Biomaterialia*. 6(2), 486-493 (2010).
- 999 107. Guadana R, Jwala J, Boddu SHS, Mitra AK. Recent perspectives in ocular drug delivery.
- 1000 Pharmaceutical Research. 26(5), 1197-1216 (2009).
- 1001 108. Edelhauser HF, Rowe-Redelman CL, Robinson MR, Dawson DG, Chader GJ, Grossniklaus
- HE,Rittenhouse KD, Wilson CG, Weber DA, Kuppermann BD, Csaky KG, Olsen TW,
- Kompella UB, Holers VM, Hageman GS, Gilger BC, Campochiaro PA, Whicup SM, Wong WT.
- 1004 Ophthalmic drug delivery systems for the treatment of retinal diseases: basic research to clinical
- applications. *Investigative Ophthalmology and Visual Science*. 51(11), 5403-5420 (2010).
- 1006 109. Williams, GA, Haller JA, Kuppermann BD, Blumenkranz MS, Weinberg DV, Chou C,
- 1007 Whitcup SM. Dexamethasone posterior-segment drug delivery system in the treatment of
- 1008 macular edema resulting from uveitis or Irvine-Gass syndrome. American Journal of
- 1009 Ophthalmology. 147(6), 1048-1054 (2009).
- 1010 110. Haller JA, Kuppermann BD, Blumenkranz MS, Williams AD, Weinberg DV, Chou C,
- 1011 whitcup SM. Randomized controlled trial of an intravitreous dexamethasone drug delivery
- system in patients with diabetic macular edema. JAMA Ophthalmology. 28(3), 289-296 (2010).
- 1013 111. Bergeles C, Kummer MP, Kratochvil BE, Framme C, Nelson BJ. Steerable intravitreal
- inserts for drug delivery: in vitro and ex vivo mobility experiments. *Medical Image Computing*
- 1015 *and Computer Assisted Intervention.* 14(1), 33-40 (2011).
- 1016 112. Zhang G, Feng X, Wabner K, Fandrey C, Nagwi A, Weidmann T, Olsen TW. Intraocular
- 1017 nanoparticle drug delivery: a pilot study using an aerosol during pars plana vitrectomy.
- 1018 Investigative Ophthalmology and Visual Science. 48(11), 5243-5249 (2007).

- 1019 113. Kleinmann G, Apple DJ, Chew J, Hunter B, Stevens S, Larson S, Mamalis N, Olson RJ.
- 1020 Hydrophilic acrylic intraocular lens as a drug-delivery system for fourth-generations
- fluoroquinolones. Journal of cataract refractive Surgery. 32(10), 1717-1721 (2006).
- 1022 114. Rayner C-flex® / Superflex® Monofocal IOL. Online, UrL:
- http://www.rayner.com/products/c-flex-superflex [Accessed: 24/02/2014].
- 1024 115. Davis LT, Kumar N, Nijm LM, Ulanski LJ, Tu, EY, Fiscella RG, Peterson RJ, Glickman
- 1025 RD. An adaptable HPLC method for the analysis of frequently used antibiotics in ocular
- 1026 samples. *Journal of Chromatography B.* 878(26), 2421-2426 (2010).
- 1027 116. STAAR Surgical. Online, Url: http://staar.com/products/collamer-iols/ [Accessed:
- 1028 24/02/2014].
- 1029 117. Duarte AR, Simplico AL, Vega-Gnzalez A, Subra- Paternault P, Coimbra P, Gil MH, De
- 1030 Sousa HC. Impregnation of an intraocular lens for ophthalmic drug delivery. Current Drug
- 1031 Delivery. 5(2), 102-107 (2008).
- 1032 118. Conway BR. Recent patents on ocular drug delivery systems. Recent Patents on Drug
- 1033 *Delivery and Formulation*. 2(1), 1-8 (2008).
- 1034 119. Vitrasert®. Online, Url: http://www.bioportfolio.com/resources/drug/18877/Vitrasert.html
- 1035 [Accessed: 24/02/2014].
- 1036 120. Retisert®. Online, Url: http://www.psivida.com/products-retisert.html [Accessed:
- 1037 24/02/2014].
- 1038 121. Medidur insert technology. Mruthyunjaya P, Jaffe GJ. Online, Url:
- http://www.retinalphysician.com/printarticle.aspx?articleID=100862 [Accessed: 24/02/2014].
- 1040 122. Sivaprasad S, Mcluskey P, Lightman S. Intravitreal steroids in the management of macular
- oedema. Acta Ophthalmologica Scandinavica. 84(6), 722-733 (2006).

- 1042 123. Stuart A. The promise of implantable drug delivery systems. *American Academy of*
- 1043 Ophthalmology. Online, URL: http://www.aao.org?publications/eyenet/201003/feature.cfm
- 1044 [Accessed: 14/06/2014].
- 1045 124. Posurdex implant technology. Kuppermann BD. (2007). Online, Url:
- http://www.retinalphysician.com/printarticle.aspx?articleID=100863 [Accessed: 24/02/2014].
- 1047 125. Tan DT, Chee SP, Lim I, Lim AS. Randomised clinical trial of a new dexamethasone
- delivery system (Surodex) for treatment of post-cataract surgery inflammation. *Ophthalmology*.
- 1049 106(2), 223-231 (1999).
- 1050 126. Gulati V, Pahuia S, Fan S, Toris CB. An experimental steroid responsive model of ocular
- inflammation in rabbits using an SLT frequency doubled Q switched Nd:YAG laser. Journal of
- 1052 *Ocular Pharmacology and Therapeutics*. 29(7), 663-669 (2013).
- 1053 127. Cheng Y, Xu Z, Ma M, Xu T. Dendrimers as drug carriers:applications in different routes
- of drug administration. *Journal of Pharmaceutical Sciences*. 97(1), 123-143 (2008).
- 1055 128. Duan X, Sheardown H. Dendrimer cross-linked collagen as a corneal tissue engineering
- scaffold: mechanical properties and corneal epithelial cell interactions. *Biomaterials*. 27(26),
- 1057 4608-4617 (2006).
- 1058 129. Patel CC, Roy H. Pars plana vitrectomy. (2013). Online. Url:
- http://emedicine.medscape.com/article/1844160-overview#a01 [Accessed: 27/02/14].
- 1060 130. Banerjee, Carvalho E. Nanoparticle in-situ gels as vitreous humor substitutes for ocular
- 1061 diseases. WO2011135400A1 (2011).