

*The impact of secondary pests on *Bacillus thuringiensis* (Bt) crops*

Article

Accepted Version

Catarino, R., Ceddia, G., Areal, F. J. and Park, J. (2015) The impact of secondary pests on *Bacillus thuringiensis* (Bt) crops. *Plant Biotechnology Journal*, 13 (5). pp. 601-612. ISSN 1467-7652 doi: <https://doi.org/10.1111/pbi.12363> Available at <http://centaur.reading.ac.uk/39915/>

It is advisable to refer to the publisher's version if you intend to cite from the work. See [Guidance on citing](#).

To link to this article DOI: <http://dx.doi.org/10.1111/pbi.12363>

Publisher: Wiley-Blackwell

All outputs in CentAUR are protected by Intellectual Property Rights law, including copyright law. Copyright and IPR is retained by the creators or other copyright holders. Terms and conditions for use of this material are defined in the [End User Agreement](#).

www.reading.ac.uk/centaur

CentAUR

Central Archive at the University of Reading

Reading's research outputs online

Running Title: The Impact of Secondary Pests on *Bt* crops

The Impact of Secondary Pests on *Bt* Crops

Rui Catarino^{1*}; Graziano Ceddia²; Francisco J. Areal¹; Julian Park¹

¹School of Agriculture, Policy and Development, University of Reading, Reading, UK

²Department of Public Governance and Sustainable Development, MODUL University, Vienna, Austria

*Correspondence concerning this article should be addressed to Rui Catarino, Department of Agricultural and Food Economics, The University of Reading, Earley Gate, PO Box 236, Reading, RG6 6AR, Email: r.catarino@reading.ac.uk, Phone +44 (0) 118 378 5038

Word count: 7391

Keywords: Secondary Pest; Pest Outbreak; Ecological impact; Genetically Engineered Insect Resistance Crops; *Bacillus thuringiensis*.

Abstract

The intensification of agriculture and the development of synthetic insecticides enabled worldwide grain production to more than double in the last third of the 20th century. However, the heavy dependence and, in some cases, overuse of insecticides has been responsible for negative environmental and ecological impacts across the globe, such as a reduction in biodiversity, insect resistance to pesticides, negative effects on non-target species (e.g. natural enemies) and the development of secondary pests. The use of recombinant DNA technology to develop genetically engineered (GE) insect resistant crops could mitigate many of the negative side effects of pesticides. One such genetic alteration enables crops to express toxic crystalline (Cry) proteins from the soil bacteria *Bacillus thuringiensis* (*Bt*). Despite the widespread adoption of *Bt* crops, there are still a range of unanswered questions concerning longer term agro-ecosystem interactions. For instance, insect species that are not susceptible to the expressed toxin can develop into secondary pests and cause significant damage to the crop. Here we review the main causes surrounding secondary pest dynamics in *Bt* crops and the impact of such outbreaks. Regardless of the causes, if non-susceptible secondary pest populations exceed economic thresholds, insecticide spraying could become the immediate solution at farmers' disposal, and the sustainable use of this genetic modification technology may be in jeopardy. Based on the literature, recommendations for future research are outlined that will help to improve the knowledge of the possible long-term ecological trophic interactions of employing this technology.

1. Introduction

With the intensification of agriculture and development of synthetic insecticides in the mid-twentieth century, scientists and farmers regarded technological development as the solution to reduce pest losses and enhance food production (Oerke, 2006). Insecticide use has enabled worldwide grain production to more than double in the last third of the 20th century (Krebs et al., 1999). Conversely, the heavy dependence and overuse of insecticides has had many unintended consequences. Insecticides have been responsible for poisoning millions of people including numerous fatalities across the globe (Ecobichon, 2001; Jeyaratnam, 1990). Negative environmental impacts, such as a reduction in biodiversity, insect resistance to pesticides, negative effects on non-target species (e.g. natural enemies) and the development of secondary pests have also been attributed to the use of insecticides (Hardin et al., 1995; Matson et al., 1997; Vitousek et al., 1997). Even so in 2011 about 1.3 thousand tons of insecticidal active ingredients were used in the world (FAOSTAT, data 2011). The use of recombinant DNA technology to develop genetically engineered (GE) insect resistant crops could

mitigate many of pesticide's negative side effects. The expressing of toxic crystalline (Cry) proteins from the soil bacteria *Bacillus thuringiensis* (*Bt*) by *Bt* crops is one such genetic alteration. This comes with the hope of supporting an agricultural revolution that is more productive (Conway and Toenniessen, 1999) while maintaining healthy and functional ecosystems for future generations (Poppy and Sutherland, 2004; Tilman et al., 2001).

Overall, commercialized *Bt* crops have performed well against their target pests (Carrière et al., 2010; Tabashnik et al., 2008). Additionally, due to the high specificity and efficiency of *Bt* Cry toxins, it is generally accepted that any eventual detrimental impact on non-target organisms (NTO) is lower than that caused by broad-spectrum insecticides (Areal and Riesgo, 2015; Cattaneo et al., 2006; Marvier et al., 2007). The reduced use of insecticides may then allow for a higher diversity and density of beneficial arthropods (Lu et al., 2012; Naranjo, 2005). Also, in theory, the reduced reliance on insecticides enabled by *Bt* crops can lead to a reduction in farm operations with associated economic, environmental and social benefits (Areal et al., 2013; Wolfenbarger and Phifer, 2000). Still, regardless of worldwide adoption of *Bt* crops it remains a controversial technology which is surrounded by uncertainty, dividing the scientific community (e.g. the following debate: Andow et al., 2009; Lövei et al., 2009; Shelton et al., 2009). These uncertainties are mainly based on alleged methodological research faults concerning the potential long-term impacts of *Bt* crops, such as the development of insect resistance and the impact on NTOs (Garcia and Altieri, 2005; Lövei et al., 2009; Smale, 2006). Two arguments are often mentioned in connection with possible long-term impacts: i) ecological shifts can take several years to manifest (Ho et al., 2009); and ii) impacts of *Bt* crops vary temporally and spatially, which may not reflect the results obtained in laboratory studies (Andow et al., 2006; Lövei et al., 2009). A further concern is that other insect species that are not susceptible to the expressed toxin will develop into secondary pests and cause significant damage to the crop (Sharma and Ortiz, 2000; Wu and Guo, 2005). If these impacts materialise they will certainly affect other trophic chains which, according to the magnitude of the impact, could become of high economic and ultimately of ecological relevance.

This paper focuses on the development and effects of secondary pests on *Bt* crops. This issue, although of high importance, has to date received only limited attention in spite of Harper's warning (Harper, 1991, p.22) that, "ignoring secondary pests can lead to devastating crop damage that may continue over a considerable period of time". By reviewing the relevant literature, this paper has three main goals: i) to assess the main causes of secondary pest outbreaks when arising in association with *Bt* crops; ii) to review the impacts of these outbreaks, as currently understood; and iii) to provide recommendations for future research.

2. Characterisation of secondary pests in *Bt* crops

The concept of secondary pests is intrinsically linked with that of NTOs. NTOs in the broader context of GE crops include, “all living organisms that are not meant to be affected by newly expressed compounds in GE crops, and that can be potentially exposed, directly or indirectly, to the GE crop and/or its products in the agro-ecosystem where GE crops will be released or in adjacent habitats” (Arpaia, 2010, p.14). Although food webs in agro-ecosystems are typically simpler than those in natural habitats, they still include multi-trophic relationships (Altieri, 1999; Arpaia, 2010). In any given cropping system, numerous species and scores of ecosystem functions can be found, although only a few can cause major losses in crop yield or quality (Hooper et al., 2005; Matson et al., 1997). A lethal or sub-lethal effect of a *Bt* crop upon one or a group of NTOs might occur through direct exposure to the *Bt* toxin or indirectly due to changes in the ecosystem on which that species depends (Snow et al., 2005). In order to assess the impact of *Bt* crops on NTOs at different trophic levels, scientists would need to be acquainted with the majority of arthropod species prevalent in a given agro-ecosystem (Meissle et al., 2010). Lövei et al. (2009), in summarising published literature, concluded that stating that *Bt* crops will pose “no harm” to NTOs is still a premature conclusion due to the limited number of non-target species studied.

There are two relevant phenomena in agricultural systems that are considered as ecological backlash events: pest resurgence and outbreaks of secondary pests. The former refers to a situation in which a suppressed pest population unexpectedly rebounds following a pest control action, exceeding the economic injury level (Hardin et al., 1995). The latter, and the focus of this paper, refers to the emergence of a pest other than that originally targeted by an agricultural intervention (the ‘targeted’ or ‘primary’ pest), and can be seen as “replacement” for the primary pest (Hardin et al., 1995; Metcalf, 1980). According to the FIFRA Scientific Advisory Panel (1998), a secondary pest is a "non-targeted" pest that has historically posed small or negligible economic threat, but which could be affected directly by a dose expressed in a *Bt* crop, or indirectly through changes in insecticide use patterns. Berryman et al. (1987, p.3) defines outbreaks of secondary pests as ‘an explosive increase in the abundance of a particular species that occurs over a relatively short period of time’. Termed a “type II resurgence” by Metcalf (1986), this can arise when the primary pest is strongly affected by a pest management strategy, yet is replaced by another pest not affected by this strategy. The causes responsible for both pest resurgence and outbreaks of secondary pests are relatively similar which includes reduction in the number of natural enemies and removal of competitors (Hardin et al., 1995; Ripper, 1956). In the event of

a secondary pest outbreak, additional pest management interventions are required. In most cases this results in crop spraying with a broad-spectrum insecticide (Gross and Rosenheim, 2011).

3. Causes for secondary outbreaks in *Bt* crops

The employment of *Bt* crops might have non-intuitive negative effects on agricultural ecosystem interactions and on farm profits (Sharma and Ortiz, 2000; Wolfenbarger and Phifer, 2000). Secondary pests, which before were of minor importance, might now find favourable conditions and themselves become major pests (Lu et al., 2010). Three main drivers may trigger an outbreak of secondary pest species with the use of *Bt* crops: i) a reduction in broad-spectrum insecticide applications; ii) a reduction in natural enemy populations; or iii) a decrease in inter-specific competition with the target pest. Each of these is in turn explored below.

i) A reduction in broad-spectrum insecticide applications

The introduction of *Bt* technology, at least in the early years, brought significant decreases in insecticide application among adopters, considerably alleviating the negative impacts associated with such insecticides (Kouser and Qaim, 2011; Krishna and Qaim, 2012; Meissle et al., 2010). Despite warnings from several authors (e.g. Sharma and Ortiz, 2000; Wu and Guo, 2005) that some NTOs could appear in such numbers that they become key insect pests in *Bt* crop fields, specific measures to combat their population increases were not taken. Consequently there have been outbreaks of secondary pests which were previously controlled by the insecticide applications originally targeting the primary pest (Lu et al., 2010; Pemsil et al., 2011). This situation has been particularly evident in *Bt* cotton production in China. Less than 3 years after its introduction in 1998, several pest groups including whiteflies, plant hoppers, aphids, mirids and mealy bugs increased in number (Men et al., 2004 ; Yang et al., 2005a). Similarly in *Bt* maize there is evidence that several secondary pests have acquired higher levels of agronomic importance (Eizaguirre et al., 2010; Erasmus et al., 2010; Gray et al., 2009; Pérez-Hedo et al., 2012) (see section 4ii for further details). As a consequence in some cases farmers have had to re-commence the use of insecticide applications because *Bt* cropping systems have failed to control insect pest populations.

ii) Reduction of natural enemies

Agro-ecosystem biodiversity is important not only because of its fundamental ecological, environmental and anthropocentric value but also because it is vital to a healthy and sustainable agriculture (Hooper et al., 2005; Matson et al., 1997). The employment of *Bt* crops and the consequent reduction in

insecticide usage increases the significance of the function of natural enemies to control secondary pests (Naranjo, 2005). Natural enemies include predators, parasitoids and pathogens. Natural enemies are critical to ecosystem functioning by inhibiting the excessive multiplication of potential pests in agricultural systems through 'biological control' (Bianchi et al., 2006; Wilby and Thomas, 2002). Natural enemies alone may be sufficient in some cases to keep secondary pest populations under economic injury thresholds (Hardin et al., 1995; Wolfenbarger et al., 2008). Hence, a major concern related to the growing of *Bt* crops is their potential impact on the abundance of natural enemies (Marvier et al., 2007; Poppy and Sutherland, 2004). The selectivity of Cry toxins is not entirely known, with the potential for unintended effects on beneficial species which may influence other non-susceptible pests (Lövei et al., 2009). However, interactions between prey and natural enemies are extremely complex. Not all herbivores that feed on *Bt* plants take up the toxin, nor will all natural enemies be negatively affected by prey that have ingested the toxin (e.g. Dutton et al., 2002).

Due to these complex uncertainties regarding ecological risks, many laboratory and field research studies have been conducted in order to evaluate the impact of *Bt* toxins on the natural enemies of potential secondary pests. While several laboratory studies reported no significant effects on natural enemies (e.g. Dutton et al., 2002; Li and Romeis, 2010; Meissle and Romeis, 2009), several others have indicated negative effects (e.g. García et al., 2012; González-Zamora et al., 2007; Hilbeck et al., 1998). Results from studies performed at a field level show similar variation; some found no significant impacts (e.g. Chen et al., 2006; Eckert et al., 2006; Pons et al., 2005), while other studies reported negative effects (e.g. Meissle et al., 2005; Obrist et al., 2006; Stephens et al., 2012). The overall dichotomy of results across the literature is striking (see Lang and Otto, 2010; Lövei and Arpaia, 2005; Lövei et al., 2009; Marvier et al., 2007; Wolfenbarger et al., 2008 for detailed reviews). The main source of uncertainty relates to the degree to which laboratory studies are of relevance to the complexity of field-scale agro-ecosystems (Lövei and Arpaia, 2005; Lövei et al., 2009; see also section 5). Natural enemies are often present in higher numbers in insecticide-free conventional fields than on *Bt* fields (Marvier et al., 2007; Naranjo, 2009). It is also widely accepted that the use of insecticides has larger direct negative effects on natural enemies than does the use of *Bt* crops (Cattaneo et al., 2006; Romeis et al., 2009; Wolfenbarger et al., 2008). Overall this suggests that in field settings, while *Bt* crops do have an impact on natural enemies, this is not as strong as the direct effect of insecticide.

The impact of *Bt* toxins on natural enemies can occur through direct and/or indirect effects (Romeis et al., 2006). Direct impacts might occur due to the ingestion of the insecticidal protein (Meissle et al., 2005; Obrist et al., 2006; Stephens et al., 2012). The mechanism of action of several available *Bt* toxins is still

unknown or inconclusive (Lövei and Arpaia, 2005; Lövei et al., 2009). Thus, it is conceivable that *Bt* toxins may cause similar negative effects on predators as they do on the target herbivores (Andow et al., 2006). In a recent study (Stephens et al., 2012), *Bt* proteins were passed from the Cry3Bb *Bt*-maize plant to the predator (*Harmonia axyridis*, a common coccinellid) via prey consumption (*Rhopalosiphum maidis*, the corn leaf aphid and *Rhopalosiphum padi*, the bird cherry-oat aphid), which significantly reduced their life span. Furthermore, although not yet demonstrated in the context of *Bt* crops, there is also concern regarding toxin bioaccumulation through the food chain, possibly driving cascade effects within the ecosystem (Chen et al., 2009). Indirect effects might manifest through reductions in prey/host populations or in the nutritional quality of the prey. Impacts of the toxin on herbivores may manifest at a sub-lethal level which can affect life parameters such as lifespan and fecundity (Meissle and Romeis, 2009; Romeis et al., 2004). There is evidence that the low nutritional quality of prey items after they have ingested *Bt* proteins has a significant impact on the performance, development and even survival of natural enemies (Dutton et al., 2002; Obrist et al., 2006; Stephens et al., 2012). Moreover, high mortality rates in the target species may cause a reduction of specialist natural enemies, which themselves can be important prey for generalist predators (Stephens et al., 2012). Additionally, prey species in general might migrate to non-*Bt* fields in search of preferable food resources (Daly and Buntin, 2005; Naranjo, 2005). Thus, if prey availability for secondary pest predators in *Bt* fields is scarce, predators might be encouraged to “migrate” to adjacent conventional crops, negatively affecting their abundance within *Bt* fields (Razze and Mason, 2012; Sisterson et al., 2007). As a result, any lethal or sub-lethal impacts on pest predators will disproportionately affect insect population dynamics. Hence, it may be possible that these negative impacts will permit the development of secondary pests in the crop itself or even in neighbouring crops (Gross and Rosenheim, 2011; Gutierrez et al., 2006). Understanding the direct and indirect effects of *Bt* cultivars on natural enemies is central for the management of insect pests since undoubtedly these insects play a major role in biological control of primary and secondary pests (Naranjo, 2009; Naranjo, 2011; Snyder et al., 2006).

iii) Species replacement

Competition may play an important role in the dynamics of herbivorous insects (Kaplan and Denno, 2007). However, the importance of replacement between primary and secondary pests has generally been ignored in conventional agriculture (Denno et al., 1995; Hardin et al., 1995), but especially in *Bt* cropping. *Bt* crops, as insecticides, are an artificially imposed disturbance on the ecosystem, hence it is not surprising that

niche rearrangement occurs (Catangui and Berg, 2006). It is possible that when a primary pest is successfully controlled by a *Bt* toxin, a non-susceptible species starts to utilise the newly available ecological resource (Gross and Rosenheim, 2011; Hardin et al., 1995). This situation occurs in cases where, prior to the pest management treatment, the primary pest is a dominant competitor species and the secondary pest is a weak competitor (Shivankar et al., 2007).

A notorious example of species replacement is the western bean cutworm (WBC) (*Striacosta albicosta* (Smith)) a noctuid moth native to West and Central America (Douglass et al., 1957). In the mid 1990's, the WBC began an expansion of range size which correlated with the introduction of transgenic maize. It has now effectively established itself as a major Lepidopteran pest of maize crops in some areas of the Corn Belt in the US and Canada (Dorhout and Rice, 2010; Lindroth et al., 2012; Michel et al., 2010). This secondary pest shows low susceptibility to most transgenic maize currently commercialized (Eichenseer et al., 2008). Transgenic crops expressing Cry1Ab and Cry9C toxins have larger populations of WBC compared to conventional maize (Catangui and Berg, 2006; Dorhout and Rice, 2010). It is possible that changes in cultural practices (e.g. conservation tillage and reduced insecticide use) due to the widespread adoption of *Bt* maize across these areas might have contributed to the WBC's rapid expansion (Hutchison et al., 2011). However, since the widespread planting of *Bt* maize hybrids has effectively eliminated intra-guild competition with the European corn borer (ECB) (*Ostrinia nubilalis*), and the corn earworm (CEW) (*Helicoverpa zea*) (Catangui and Berg, 2006; Dorhout and Rice, 2010) it is possible that an ecological opportunity opened for WBC (Catangui and Berg, 2006; Dorhout and Rice, 2010).

To date only one study appears to have been specifically conducted to assess the interaction between WBC and other species (Dorhout and Rice, 2010). CEW had a significant negative impact on WBC survival when both were fed on a meridic or isoline maize silk diet. CEWs are extremely aggressive by nature compared to the WBC (Douglass et al., 1957), and CEW larvae often kill WBC larvae even when the latter are present in larger numbers (Dorhout and Rice, 2010). However, when both pests were fed with a transgenic silk diet, WBC presented high survival rates (Dorhout and Rice, 2010). Competition with the ECB exists because of their similar feeding behaviour on the kernels in corn ears (Catangui and Berg, 2006). Hence, along with the high flight capacity of WBC (Michel et al., 2010), the reduction in direct competition very likely played a fundamental part in its territorial expansion.

Other examples of species replacement include the corn leafhopper (*Dalbulus maidis*) in maize in the absence of the target pest *Spodoptera frugiperda* in Argentina (Virla et al. (2010)). In Spain, the true armyworm

Mythimna unipuncta could have competitive advantage in the absence of both the Mediterranean and European corn borer (Eizaguirre et al., 2010; Malvar et al., 2004). In *Bt* cotton in the USA, stink bug pests, specifically *Nezara viridula* L. and *Euschistus servus* S., have recently become a severe problem in the absence of the target pests *H. zea* and *Heliothis virescens* (Zeilinger et al., 2011). *Helicoverpa armigera*, *Acantholeucania loreyi* and *Eublemma gayneri* could also gain competitive advantage following the displacement of *Busseola fusca* from *Bt* maize in South Africa (Van Wyk et al., 2007). As *Bt* cropping expands worldwide it is of critical importance to determine the key species – susceptible and non-susceptible pests – which might compete for resources within the same transgenic crop.

4. Impact of secondary pests on *Bt* crops

In the early years of *Bt* cropping there were reports of increased profitability in overall production due to 40-60% reductions in insecticide applications alongside increased crop yields, as compared to non-adopters (e.g. Bennett et al., 2004; Fitt, 2000; Huang et al., 2002; Pray et al., 2002; Qaim and Zilberman, 2003; Thirtle et al., 2003). There was also a reduction in human insecticide poisonings (Huang et al., 2002; Pray et al., 2002). Nonetheless, there were early concerns about the potential for secondary pest outbreaks due to the decrease in insecticide applications (Morse et al., 2005; Qaim, 2003; Wu et al., 2002). Here we focus on the development of secondary pests in two of the most important GE insect resistant crops, *Bt* maize and *Bt* cotton.

i) Cotton

From the worldwide 24.3 million hectares cropped with *Bt* cotton, India, China and USA account for 11.0, 4.2 and 4.1 million hectares respectively (James, 2013), with the adoption rate varying between 90 and 95% (James, 2013). The *Bt* cotton hectareage in Africa is increasing, for instance Burkina Faso and Sudan cropped 50% and 300% more *Bt* cotton, respectively, compared with 2012 (James, 2013).

In China, in some areas where the bollworm incidence is higher, the adoption is close to 100% (Xu et al., 2008) and undoubtedly, *Bt* cotton has reinvigorated Chinese cotton production. Historically, cotton and rice have required the largest number of insecticide applications in the world (Deguine et al., 2008). Until the end of the 20th century, insecticides were intensively applied to control the cotton bollworm (Wu and Guo, 2005). However in the early 1990s the effective control of this pest became problematic, the cotton bollworm became resistant to most insecticides due to their overuse (Deguine et al., 2008; Wu and Guo, 2005). Following the introduction of *Bt* technology in 1999, insecticide applications in *Bt* cotton fields dropped from about 61 kg/ha (20 applications) per year, to approximately 12 kg/ha (6.6 applications) per year (Huang et al., 2002).

By 2002 this figure started to increase, reaching on average 15.6 kg/ha (10.7 applications) per year of insecticides, of which 4.7 kg were used against cotton bollworm, and the remaining against *lygus* bug and other pests (Pemsl et al., 2011). By 2005, farmers applied roughly the same amount against the cotton bollworm, but the amount sprayed against secondary pests had increased by 20%, to a total of 18.6 kg/ha (14.2 applications) per year (Pemsl et al., 2011). Within the space of approximately 10 years, the initial advantage of *Bt* crops had gone; Zhao et al. (2011) reported that *Bt* adopters were using on average between 16 and 22 insecticide applications, while conventional cottons farmer were using only 11 to 17 applications per year. Nowadays those insects once considered of minor relevance are actually the main concern of farmers' (Pemsl and Waibel, 2007). The drop in insecticide use and the ineffectiveness of *Bt* cotton against these secondary pests has led to a reversal of the ecological role of cotton (Li et al., 2011; Lu et al., 2010). Conventional cotton had been a population sink for the mirid bug secondary pest, while nowadays *Bt* cotton fields are a source of these pests (Lu et al., 2010). This has led to a situation where there are no major differences in the total quantity and expenditure in insecticide application between *Bt* and conventional cotton farmers (Yang et al., 2005b; Zhao et al., 2011). However, when comparing with the period prior to *Bt* adoption, farmers are generally not worse off. Cotton production is still effective and farmers are applying fewer sprayings in early season, with fewer cases of human poisoning (Huang et al., 2014). Moreover, a higher survival of generalist arthropod predators has been recorded (ladybirds, lacewings and spiders) providing additional biocontrol to neighbouring crops, such as maize and soybean (Huang et al., 2014; Lu et al., 2012).

Indian cotton farming is comparable with China, with numerous small scale farmers (Huang et al., 2002; Qaim et al., 2009). Recent evidence shows that secondary pests are now posing a major problem (Nagrare et al., 2009), with farmers battling against non-target insects (Stone, 2011). Ramaswami et al. (2012) found no significant difference between adopters and non-adopters in terms of insecticide use. This is consistent with Indian *Bt* farmers' perceptions, who attributed a total of 77% of cotton damage to aphids and other sucking pests and only 23% to the primary Lepidopteran pests, leading to 99% of the famers spraying against secondary pests (Stone, 2011). Elsewhere in the world similar issues to the Chinese and Indian cases have been reported in cotton. Adopting farmers are either still using significant numbers of insecticide applications in order to control secondary pests, or the damage caused by these pests has increased. Some examples include: South Africa (Hofs et al., 2006; Schnurr, 2012), Burkina Faso (Dowd-Uribe, 2014), Pakistan (Jaleel et al., 2014), Australia (Wilson et al., 2013), Brazil (Sujii et al., 2013) and Mexico (Traxler and Godoy-Avila, 2004).

In the USA for example, in the mid-southern and south-eastern cotton-producing regions, there has been a significant increase in the number of insects considered as secondary cotton pests, such as aphids, leafhoppers, mirid plant bugs and stinkbugs (Naranjo, 2011). The same author analysed National Cotton Council data, reporting that before *Bt* cotton adoption, farmers were applying an average of 17 applications per hectare, and this figure dropped post-adoption to five applications (a 71% reduction). Additionally, *Bt* cotton losses due to pest damage are around 5.4%, a decrease of 27% when compared to pre-1996 levels. Nonetheless, while insecticide use to control primary pests has decreased, insecticide applications used to control secondary pests such as plant bugs have nearly doubled to approximately four applications per hectare in order to achieve adequate control (Naranjo, 2011). In contrast to the other cases around the world, most of the secondary pests in the US are being effectively managed with sensible use of insecticides and other Integrated Pest Management (IPM) tactics (Naranjo and Ellsworth, 2009).

ii) Maize

The economic benefit of *Bt* maize associated with the regional suppression of specific pest populations is significant (Areal et al., 2013; Carpenter, 2010; Riesgo et al., 2012). Hutchison et al. (2010) estimated the cumulative benefits of controlling ECB with *Bt* maize over the last 14 years at \$6.8 billion for maize growers in the US Midwest, with an estimated 60% of this accruing to non-*Bt* maize growers. This is due to savings in insecticide applications because of the region-wide suppression of ECB populations. Presently, in the USA the most problematic secondary pest in *Bt* maize is the WBC (see section 3iii), causing up to 70% yield losses (Catangui and Berg, 2006). This value is not surprising, since only one WBC larvae per maize plant at dent stage can reduce yields by 232 kg/ha (Appel et al., 1993). Both the CEW and the fall armyworm are considered important secondary pests too. Their moderate survival rates in *Bt* maize expressing Cry1Ab and CryIF makes them economically important (Archer et al., 2001; Hardke et al., 2011; Storer et al., 2001).

Currently, the only *Bt* maize allowed for cultivation in Europe contains the transformation event MON810 (Monsanto Company) expressing Cry1Ab *Bt* toxin (EFSA, 2010), although several other events are under evaluation by the European Food Safety Authority (EFSA). This transgenic maize presents a highly efficient level of resistance to the two primary maize lepidopteran borers present in the EU, the Mediterranean and the European corn borer (Eizaguirre et al., 2010; González-Núñez et al., 2000). In European conditions, *Bt* Cry1Ab is not efficient against several secondary pests, such as the western corn rootworm and the true armyworm (Gray et al., 2009; Pérez-Hedo et al., 2012). The western corn rootworm was first noticed in Europe in the mid-1980s (Bača, 1994; Miller et al., 2005) and it has been spreading across the continent at an average

rate of 33 to 40 km per year (Gray et al., 2009; Meinke et al., 2009). Its presence is more common in central and eastern European countries and in the Po Valley in Italy where attributable yield losses of about 2–3% have been reported (Meissle et al., 2010).

Field research concerning secondary pests in Europe has mainly been conducted in Spain. Recent studies have revealed that the true armyworm is only mildly susceptible to *Bt* maize expressing the Cry1Ab toxin (González-Cabrera et al., 2013; Pilcher et al., 1997). Furthermore, field trials found no substantial differences in the number of true armyworm larvae per plant nor in their larval development between *Bt* and its isogenic variety (Eizaguirre et al., 2010; Pérez-Hedo et al., 2012). Hence, it is possible that the increasing use of transgenic maize expressing Cry1Ab toxin will further amplify the true armyworm's importance due to decreasing conventional insecticide applications. In time this could lead to it becoming a major pest (González-Cabrera et al., 2013; Pérez-Hedo et al., 2012).

In South Africa, *Bt* maize has the potential, when well-managed, to effectively control primary lepidopteran pests, such as *B. fusca*, *S. calamistis* and *C. partellus* (Kruger et al., 2012; Van den Berg and Van Wyk, 2007; Van Wyk et al., 2009). However, several important secondary pests are also present, including *A. segetum*, *H. armigera* and *A. loreyi* (Erasmus et al., 2010; Van Wyk et al., 2008; Van Wyk et al., 2009). Although these secondary pests may show some degree of susceptibility to Cry proteins (their densities are usually lower in *Bt*-maize fields compared to non-*Bt* fields) they are able to seriously damage the crop under field conditions (Van Wyk et al., 2007; Van Wyk et al., 2008). Similar importance is now given to *H. armigera* in China, Australia and South Africa (Tabashnik et al., 2003; Van Wyk et al., 2008) and to the corn leafhopper, an efficient vector of several plant pathogens, in Argentina (Bastos et al., 2007). Ecological explanations for higher attraction to *Bt* maize in some pest species have been found; for example chemical and/or morphological characteristics expressed by the *Bt* maize make it especially attractive to the corn leafhopper secondary pest (Bastos et al., 2007).

This review suggests that in both *Bt* maize and *Bt* cotton, the increased significance of secondary pests is intrinsically linked with insecticide use. On the one hand, as previously shown, decreases in insecticide applications can allow non-*Bt* susceptible insects to increase in numbers within the *Bt* crop. On the other hand, broad-spectrum insecticide spraying is the cheapest and most efficient solution for farmers avoiding severe crop damage due to a sudden pest outbreak of a non-target pest species. Several other issues are also linked with insecticide use in *Bt* crops, including pest resistance caused mainly by the lack of refuge strategies, weak

institutional structures, poor education and a lack of understanding concerning the technology (Dowd-Uribe, 2014; Morse et al., 2007; Yang et al., 2005a).

5. Outstanding issues

Regardless of the cause, if non-susceptible secondary pest populations exceed economic thresholds, the sustainability of the technology may be in jeopardy. If natural enemies are negatively affected by *Bt* maize, directly or indirectly, an ecological opportunity may appear for the emergence of a new pest which had previously been controlled through predation or parasitism. Consequently, insecticide spraying is the only immediate solution at farmers' disposal, which will disrupt the natural enemies' complex. If a secondary pest outbreak occurs due to an ecological opportunity arising from a drop in the density of a former major herbivore, the same immediate solution could be used with equivalent impacts. Hence, farmers growing *Bt* crops will potentially recommence running on the insecticide treadmill observed in the 20th century (van den Bosch, 1978) leading once again to the negative impacts of insecticides on the environment that it was hoped *Bt* crops would reduce (Krebs et al., 1999; Pemsl et al., 2011). There are serious disadvantages associated with over-use of pesticides, including human poisonings (Ecobichon, 2001; Jeyaratnam, 1990), the emergence of pest resistance (Metcalf, 1987) and natural enemy mortality (Hardin et al., 1995; Metcalf, 1987). Additionally, pests tend to increase their reproductive rate when stressed by sub-lethal quantities of a control agent; a phenomenon known as "hormoligosis" (Luckey, 1968; Morse, 1998). It appears that to date a potential "hormoligosis" effect of *Bt* crops has not been studied, even though this may be partly responsible for past outbreaks of secondary pests related to the misuse of insecticides (Cordeiro et al., 2013; Gross and Rosenheim, 2011; Guedes and Cutler, 2014). Research presented in this review suggests that secondary pests are eroding some of the economic and ecological benefits of *Bt* crops. Chinese cotton production is a clear example. In fact we have shown that, even in successful *Bt* cropping systems (such as *Bt* cotton in the USA), insecticide applications remain a strategically important method of controlling secondary pest outbreaks.

New stacked events expressing several *Bt* toxins may temporally overcome some of the drawbacks associated with secondary pests. Scientists are hopeful that these stacked crops will mitigate some of the concerns raised so far for single traits and still increase yields even further (e.g. Shi et al., 2013). From an ecological perspective, such expectations have yet to be proven as stacked events may equally cause faster changes in ecosystem processes, affecting the resilience of the system as a whole to adapt efficiently. Furthermore, some agro-ecosystem responses occur over a long time frame so only long-term studies could

effectively detect any effects (see Symstad et al., 2003 for a detailed discussion). For example, continuous exposure to a range of *Bt* toxins throughout the full season may affect prey species and food chains (Groot and Dicke, 2002), and the occurrence of resistance may be increased in pests with low susceptibility to *Bt* toxins over time (Brévault et al., 2013). This lack of certainty regarding ecological impacts and the complexity of agro-ecosystems have led to questions about the conclusions of several studies assessing the impacts of transgenic crops due to their simplistic methodological approaches (e.g. Andow et al., 2006; Dowd-Uribe, 2014; Glover, 2010a; Glover, 2010b; Kruger et al., 2012; Lövei et al., 2009; Smale, 2006; Stone, 2011). There is a risk that interactions evaluated over a short period fail to detect potential longer-term impacts (Kouser and Qaim, 2011; Pemsal et al., 2011). The occurrence of secondary pests is clearly linked with profitability, which in turn is affected by other important factors: quality of seeds (Xu et al., 2008); development of resistance (Kruger et al., 2012); farm size (Stone, 2011); regional, social and institutional variability (Dowd-Uribe, 2014; Smale, 2006) and farmers' knowledge/education, skills and wealth (Mancini et al., 2008; Yang et al., 2005a). For example, making an assumption that early adopters are similar in terms of managerial performance to late adopters or small-scale farmers may introduce a bias to the results (Croft et al., 2007; Morse et al., 2007; Stone, 2011). Similarly, differences in agricultural systems – such as irrigated versus non-irrigated fields in India (Qaim and Zilberman, 2003) – are important factors that are often omitted from research (Stone, 2011). It is likely that such systems have differences in pest abundance and insecticide use (Stone, 2011).

Studies assessing the impact of *Bt* crops on NTOs remain controversial. Ecological criticisms are mainly based on the reliability of data, poor replicability, low numbers of possible response variables and short temporal frames, and the studies often do not take into consideration environmental variability across regions (Andow et al., 2006; Lövei and Arpaia, 2005; Marvier, 2002; Shantharam et al., 2008). Laboratory studies are essential to assess the effects of *Bt* crops on NTOs provided they are assessed across the full variety of relevant ecological contexts (Lövei and Arpaia, 2005). In the laboratory or in controlled field cages, most non-field scale studies have assessed the direct impact that the toxin has on the predator, on the prey or on the impact of a predator through prey consumption (see for example Lang and Otto, 2010; Lövei et al., 2009; Marvier et al., 2007; Wolfenbarger et al., 2008). However, the relevance of these findings within the field agro-ecosystem is uncertain (Andow and Hilbeck, 2004; Lövei and Arpaia, 2005) and such studies often fail to account for indirect spatial and temporal effects on tritrophic population dynamics (Andow et al., 2006). Further, the occurrence and distribution of insect pests in crops are non-uniform, depending instead on factors such as the agro-climatic conditions, agro-ecology, anthropogenic interventions, introduction of new crops, pest control

management techniques, and other hard to define random factors (Baker et al., 2000; Sisterson et al., 2005; Velasco Pazos et al., 2007).

6. Conclusions

Like insecticides, *Bt* crops alter agro-ecosystem processes and functioning. In some cases this may lead to large and complex landscape-level effects on pest dynamics, a rearrangement of niches, and thus a possible outbreak of secondary pests. This review has explored the reasons for, and the results of, secondary pest outbreaks in *GE insect resistant* crops, with a focus on *Bt* maize and *Bt* cotton. Undoubtedly, *Bt* crops have led to several economic and environmental advantages, but many claim that those gains, although real, have been overemphasized (Smale, 2006; Stone, 2011). Although secondary pest outbreaks are a well-known phenomenon, they have generally been overlooked in transgenic cropping research. While *Bt* crops are highly efficient at controlling target pest levels, they may not be as effective at controlling other pests that have historically posed less or even no threat (Sharma and Ortiz, 2000). The three potential mechanisms related to secondary pest emergence: i) a reduction in broad-spectrum insecticide applications; ii) a reduction in control by natural enemies; and iii) a decrease in inter-specific competition with the target pest, have relevance for better *Bt* crop management.

It is evident from the literature that, due to lower insecticide applications, secondary pests that are not susceptible to the expressed toxin are becoming an increasing concern in some agro-ecosystems where *Bt* crops are grown. The potentially negative influence of *Bt* crops on natural enemies has generated considerable debate among scientists, although there appears to be agreement that a negative impact is conceivable (e.g. Andow et al., 2009; Lövei et al., 2009; Shelton et al., 2009). This impact can be direct through the ingestion of the toxin or indirect due to changes in the agro-ecosystem on which that species depends, such as reduced prey density (Andow et al., 2006; Snow et al., 2005). Less attention has been given to ecological opportunism by competitive species (but see Dorhout and Rice, 2010; Virla et al., 2010). Secondary pests may take several years to develop to a point where they actually become a major concern (Ho et al., 2009). Hence, it has been suggested that additional research is needed to evaluate the potential long-term effects of the wide-scale adoption of new *Bt* events and their impacts on ecosystems (Krishna and Qaim, 2012). It is important that the ecological relevance of such studies is properly acknowledged (Gatehouse et al., 2011), especially with regard to the impact on ecological services across the agricultural landscape and on the resilience of regional agro-

ecosystems (Tschardt et al., 2005). Based on this review of the literature, we conclude with five major issues that require further exploration:

1. Large-scale, multi-trophic and multi-species field studies in order to reveal the extent and potential of impacts on ecosystems (Lang and Otto, 2010) since: i) *Bt* toxins concentrations vary throughout the season depending on expressed toxins and the cultivar (Nguyen and Jehle, 2009; Showalter et al., 2009); ii) Interactions between *Bt* fields and adjacent ecosystems will surely occur (e.g. natural enemy migration or niche replacement) (Dorhout and Rice, 2010; Razzi and Mason, 2012), which might carry direct and/or indirect biotic impacts across the landscape (Lundgren et al., 2009); iii) In the case of stacked *Bt* crops, potential interactions between the expressed events may occur (Zhao et al., 2005) and resistance may be increased in pests with low susceptibility to *Bt* toxins (Brévault et al., 2013). Hence, laboratory and/or single species studies may fail to capture the wider trophic impacts that occurs in field environment (Lövei et al., 2009). Only a holistic knowledge of pests and the behaviour of natural enemies will enable the formulation of a sustainable IPM framework capable of effectively suppressing secondary pest outbreaks (Lundgren et al., 2009; Sisterson et al., 2007).

2. The baseline for risk analysis studies should be adjusted. Until now studies have used conventional cropping with insecticide treatments as the main basis for comparison of risk of *Bt* crops (Meissle et al., 2011; Sisterson et al., 2007). However, this comparison should be broadened to include other scenarios, such as organic or untreated cropping systems (Andow et al., 2006). The assessment of *Bt* cropping with other IPM strategies, such as crop rotation, tillage, selective insecticides and biological control (Deguine et al., 2008; Musser and Shelton, 2003; Vasileiadis et al., 2011) would be useful, especially taking into consideration the forecasted increase in global food demand (Park et al., 2011). Failing to take this into account might lead farmers to neglect other good farming practices (Bergé and Ricroch, 2010).

3. Economic studies should move towards a wider approach, taking into consideration farmers' heterogeneity (Glover, 2010a; Glover, 2010b). Assessing the mean yielding/profits of a crop within an entire country/region will likely be biased towards wealthier and better informed/educated farmers (Sanglestawai et al., 2014). This is especially relevant in developing countries, where institutional networks are weak, making the enforcement of laws, policies and agricultural recommendations less effective (Dowd-Urbe, 2014; Kruger et al., 2011; Kruger et al., 2012; Shantharam et al., 2008; Stone,

2011; Xu et al., 2008). As Stone (2011, page 395) states, “longitudinal, multi-village, multi-ethnic, probabilistically selected, ethnographically grounded studies that avoid bias are helpful”.

4. In order to identify possible secondary pests and other non-target effects of *Bt* crops with insecticidal properties, data are needed on which arthropod species occur in a given agro-ecosystem (Truter et al., 2014). Presently, several million hectares of crops with *Bt* traits are being grown, and we should take advantage of such “large-scale field research” opportunities. Hence, continuous, post-market, spatiotemporal monitoring is critical for rapid identification of the development of ecological problems. This could lead to timely regulatory decisions and the efficient deployment of mitigation measures (Sanvido et al., 2009; Smale, 2012; Waage and Mumford, 2008). Furthermore, we suggest that post-market monitoring could help build a robust spatiotemporal database of insect species according to their ecological functions and occurrence in the specific receiving environments. Such a methodical process would also help to select a number of relevant and practical surrogate species for detailed laboratory or field tests (Hilbeck et al., 2014).

5. The importance of spatially-dynamic, bio-mathematical and -economic multi-species models in pre and post GE crop risk-assessment research has been recognized for some time (e.g. Bohanec et al., 2008; Harper and Zilberman, 1989; Marino and Landis, 1996; Yang et al., 2009). Rigorous assessments of the present and future economic impacts, based on ecological constraints, are required to provide sound information to policy makers (Ascough II et al., 2008; Holmes et al., 2010; Keller et al., 2007; McDermott et al., 2013). By allowing the manipulation of key biological parameters with economic production and damage functions, it will be possible to analyse potential solutions under different IPM scenarios, real or hypothesised (Carrasco et al., 2010; DeJonge et al., 2012; Liang et al., 2012). A robust assessment of the effects of agro-ecosystem heterogeneity on pest population dynamics might be obtained when a geographic information system (GIS) approach is added to the model (Carrière et al., 2006). From these models it would be possible to assess which species are most likely to be susceptible to landscape or environmental changes (Maiorano et al., 2014; Petrovskii et al., 2014). It is therefore important to foster research collaborations between the fields of ecology, mathematics and economics (Codling, 2014; Crowder and Jabbour, 2014).

In summary, despite the widespread adoption of *Bt* crops and a continued increase in the area on which they are grown, there are still a number of unanswered questions associated with longer term agro-ecosystem interactions, for instance the impact of secondary pests. These may not be serious enough to undermine the

use of the technology, but do require further exploration so that practical and economically viable advice can be given to farmers and so that regulators are aware of potential issues and risks during a crop's approval phase.

Acknowledgments

We thank the three anonymous referees for their comments on previous drafts of this manuscript that greatly improved the paper. The research forms part of the 'Assessing and Monitoring the Impacts of Genetically Modified Plants on Agro-ecosystems' (AMIGA) project. The AMIGA project was funded by the European Commission under Framework Programme 7.

References

- Altieri, M.A. (1999) The ecological role of biodiversity in agroecosystems. *Agric. Ecosyst. Environ.* **74**, 19-31.
- Andow, D.A. and Hilbeck, A. (2004) Science-based risk assessment for nontarget effects of transgenic crops. *BioScience* **54**, 637-649.
- Andow, D.A., Lövei, G.L. and Arpaia, S. (2006) Ecological risk assessment for *Bt* crops. *Nat. Biotechnol.* **24**, 749-751.
- Andow, D.A., Lövei, G.L. and Arpaia, S. (2009) Cry toxins and proteinase inhibitors in transgenic plants do have non-zero effects on natural enemies in the laboratory: Rebuttal to Shelton et al. 2009. *Environ. Entomol.* **38**, 1528-1532.
- Appel, L.L., Wright, R.J. and Campbell, J.B. (1993) Economic injury levels for western bean cutworm, *Loxagrotis albicosta* (Smith)(Lepidoptera: Noctuidae), eggs and larvae in field corn. *J. Kans. Entomol. Soc.*, 434-438.
- Archer, T.L., Patrick, C., Schuster, G., Cronholm, G., Bynum Jr, E.D. and Morrison, W.P. (2001) Ear and shank damage by corn borers and corn earworms to four events of *Bacillus thuriangiensis* transgenic maize. *Crop. Prot.* **20**, 139-144.
- Areal, F.J. and Riesgo, L. (2015) Probability functions to build composite indicators: A methodology to measure environmental impacts of genetically modified crops. *Ecol. Indic.* **52**, 498-516.
- Areal, F.J., Riesgo, L. and Rodriguez-Cerezo, E. (2013) Economic and agronomic impact of commercialized GM crops: a meta-analysis. *J. Agric. Sci.* **151**, 7-33.
- Arpaia, S. (2010) Genetically modified plants and “non-target” organisms: analysing the functioning of the agro-ecosystem. *Collection of Biosafety Reviews* **5**, 12-80.
- Ascough II, J.C., Maier, H.R., Ravalico, J.K. and Strudley, M.W. (2008) Future research challenges for incorporation of uncertainty in environmental and ecological decision-making. *Ecol. Model.* **219**, 383-399.
- Bača, F. (1994) New member of the harmful entomofauna of Yugoslavia *Diabrotica virgifera virgifera* LeConte (Coleoptera, Chrysomelidae). *Zaštita bilja* **45**, 125-131.
- Baker, R., Sansford, C., Jarvis, C., Cannon, R., MacLeod, A. and Walters, K. (2000) The role of climatic mapping in predicting the potential geographical distribution of non-indigenous pests under current and future climates. *Agric. Ecosyst. Environ.* **82**, 57-71.
- Bastos, C.S., Cardoso Galvao, J.C., Picanco, M.C., Gomes Pereira, P.R. and Cecon, P.R. (2007) Nutrient content affecting *Spodoptera frugiperda* and *Dalbulus maidis* occurrence in corn. *Insect Sci.* **14**, 117-123.
- Bennett, R., Ismael, Y., Morse, S. and Shankar, B. (2004) Reductions in insecticide use from adoption of *Bt* cotton in South Africa: impacts on economic performance and toxic load to the environment. *J. Agric. Sci.* **142**, 665-674.
- Bergé, J.B. and Ricroch, A.E. (2010) Emergence of minor pests becoming major pests in GE cotton in China: What are the reasons? What are the alternatives practices to this change of status? *GM Crops* **1**, 214-219.
- Berryman, A.A. (1987) The theory and classification of outbreaks. In: *Insect Outbreaks* (Barbosa, P. and Schultz, J. eds), pp. 3-30. San Diego: Academic Press.
- Bianchi, F.J.J.A., Booij, C.J.H. and Tscharntke, T. (2006) Sustainable pest regulation in agricultural landscapes: a review on landscape composition, biodiversity and natural pest control. *Proc. R. Soc. Lond. B* **273**, 1715-1727.
- Bohanec, M., Messéan, A., Scatasta, S., Angevin, F., Griffiths, B., Krogh, P.H., Žnidaršič, M. and Džeroski, S. (2008) A qualitative multi-attribute model for economic and ecological assessment of genetically modified crops. *Ecol. Model.* **215**, 247-261.
- Brévault, T., Heuberger, S., Zhang, M., Eilers-Kirk, C., Ni, X., Masson, L., Li, X., Tabashnik, B.E. and Carrière, Y. (2013) Potential shortfall of pyramided transgenic cotton for insect resistance management. *Proc. Natl. Acad. Sci. U.S.A.* **110**, 5806-5811.
- Carpenter, J.E. (2010) Peer-reviewed surveys indicate positive impact of commercialized GM crops. *Nat. Biotechnol.* **28**, 319-321.

- Carrasco, L.R., Mumford, J.D., MacLeod, A., Knight, J.D. and Baker, R.H.A. (2010) Comprehensive bioeconomic modelling of multiple harmful non-indigenous species. *Ecol. Econ.* **69**, 1303-1312.
- Carrière, Y., Crowder, D.W. and Tabashnik, B.E. (2010) Evolutionary ecology of insect adaptation to *Bt* crops. *Evol. Appl.* **3**, 561-573.
- Carrière, Y., Ellsworth, P.C., Dutilleul, P., Ellers-Kirk, C., Barkley, V. and Antilla, L. (2006) A GIS-based approach for areawide pest management: the scales of *Lygus hesperus* movements to cotton from alfalfa, weeds, and cotton. *Entomol. Exp. Appl.* **118**, 203-210.
- Catangui, M.A. and Berg, R.K. (2006) Western bean cutworm, *Striacosta albicosta* (Smith)(Lepidoptera: Noctuidae), as a potential pest of transgenic Cry1Ab *Bacillus thuringiensis* corn hybrids in South Dakota. *Environ. Entomol.* **35**, 1439-1452.
- Cattaneo, M.G., Yafuso, C., Schmidt, C., Huang, C., Rahman, M., Olson, C., Ellers-Kirk, C., Orr, B.J., Marsh, S.E. and Antilla, L. (2006) Farm-scale evaluation of the impacts of transgenic cotton on biodiversity, pesticide use, and yield. *Proc. Natl. Acad. Sci. U.S.A.* **103**, 7571-7576.
- Chen, M., Ye, G.-y., Liu, Z.-c., Fang, Q., Hu, C., Peng, Y.-f. and Shelton, A.M. (2009) Analysis of Cry1Ab toxin bioaccumulation in a food chain of *Bt* rice, an herbivore and a predator. *Ecotoxicology* **18**, 230-238.
- Chen, M., Ye, G.Y., Liu, Z.C., Yao, H.W., Chen, X.X., Shen, Z.C., Hu, C. and Datta, S.K. (2006) Field assessment of the effects of transgenic rice expressing a fused gene of Cry1ab and Cry1ac from *Bacillus thuringiensis* berliner on nontarget planthopper and leafhopper populations. *Environ. Entomol.* **35**, 127-134.
- Codling, E.A. (2014) Pest insect movement and dispersal as an example of applied movement ecology: Comment on “Multiscale approach to pest insect monitoring: Random walks, pattern formation, synchronization, and networks” by Petrovskii, Petrovskaya and Bearup. *Phys. Life Rev* **11**, 533-535.
- Conway, G. and Toenniessen, G. (1999) Feeding the world in the twenty-first century. *Nature* **402**, C55-C58.
- Cordeiro, E.M.G., de Moura, I.L.T., Fadini, M.A.M. and Guedes, R.N.C. (2013) Beyond selectivity: Are behavioral avoidance and hormesis likely causes of pyrethroid-induced outbreaks of the southern red mite *Oligonychus ilicis*? *Chemosphere* **93**, 1111-1116.
- Crost, B., Shankar, B., Bennett, R. and Morse, S. (2007) Bias from farmer self-selection in genetically modified crop productivity estimates: Evidence from indian data. *J. Agric. Econ.* **58**, 24-36.
- Crowder, D.W. and Jabbour, R. (2014) Relationships between biodiversity and biological control in agroecosystems: Current status and future challenges. *Biol. Control* **75**, 8-17.
- Daly, T. and Buntin, G.D. (2005) Effect of *Bacillus thuringiensis* transgenic corn for lepidopteran control on nontarget arthropods. *Environ. Entomol.* **34**, 1292-1301.
- Deguine, J.-P., Ferron, P. and Russell, D. (2008) Sustainable pest management for cotton production. A review. *Agron. Sustain. Dev.* **28**, 113-137.
- DeJonge, K.C., Ascough II, J.C., Ahmadi, M., Andales, A.A. and Arabi, M. (2012) Global sensitivity and uncertainty analysis of a dynamic agroecosystem model under different irrigation treatments. *Ecol. Model.* **231**, 113-125.
- Denno, R.F., McClure, M.S. and Ott, J.R. (1995) Interspecific interactions in phytophagous insects: competition reexamined and resurrected. *Annu. Rev. Entomol.* **40**, 297-331.
- Dorhout, D.L. and Rice, M.E. (2010) Intraguild competition and enhanced survival of western bean cutworm (Lepidoptera: Noctuidae) on transgenic Cry1Ab (MON810) *Bacillus thuringiensis* corn. *J. Econ. Entomol.* **103**, 54-62.
- Douglass, J., Ingram, J., Gibson, K. and Peay, W. (1957) The western bean cutworm as a pest of corn in Idaho. *J. Econ. Entomol.* **50**, 543-545.
- Dowd-Uribe, B. (2014) Engineering yields and inequality? How institutions and agro-ecology shape *Bt* cotton outcomes in Burkina Faso. *Geoforum* **53**, 161-171.
- Dutton, A., Klein, H., Romeis, J. and Bigler, F. (2002) Uptake of *Bt*-toxin by herbivores feeding on transgenic maize and consequences for the predator *Chrysoperla carnea*. *Ecol. Entomol.* **27**, 441-447.

- Eckert, J., Schuphan, I., Hothorn, L.A. and Gathmann, A. (2006) Arthropods on maize ears for detecting impacts of *Bt* maize on nontarget organisms. *Environ. Entomol.* **35**, 554-560.
- Ecobichon, D.J. (2001) Pesticide use in developing countries. *Toxicology* **160**, 27-33.
- EFSA (2010) EFSA Panel on Genetically Modified Organisms (GMO): Scientific opinion on the assessment of potential impacts of genetically modified plants on non-target organisms. *EFSA Journal* **8(11)**, 73.
- Eichenseer, H., Strohhahn, R. and Burks, J.C. (2008) Frequency and severity of western bean cutworm (Lepidoptera: Noctuidae) ear damage in transgenic corn hybrids expressing different *Bacillus thuringiensis* Cry toxins. *J. Econ. Entomol.* **101**, 555-563.
- Eizaguirre, M., Madeira, F. and López, C. (2010) Effects of *Bt* maize on non-target lepidopteran pests. *IOBC/WPRS Bulletin* **52**, 49-55.
- Erasmus, A., Rensburg, J.B.J.V. and Berg, J.V.D. (2010) Effects of *Bt* Maize on *Agrotis segetum* (Lepidoptera: Noctuidae): A Pest of Maize Seedlings. *Environ. Entomol.* **39**, 702-706.
- FAOSTAT (data 2011) Food and agriculture organization of the united nations statistics division - Pesticides (use) Database. Rome: FAOSTAT. <http://faostat3.fao.org/download/R/RP/E>, (accessed: 15 September 2014).
- FIFRA Scientific Advisory Panel (1998) Transmittal of the final report of the FIFRA scientific advisory panel subpanel on *Bacillus thuringiensis* (*Bt*) plant-pesticides and resistance management. 9–10 Feb. 1998. *Docket No. OPPTS-00231* **59**.
- Fitt, G.P. (2000) An Australian approach to IPM in cotton: integrating new technologies to minimise insecticide dependence. *Crop. Prot.* **19**, 793-800.
- García, M., Ortego, F., Castañera, P. and Farinós, G.P. (2012) Assessment of prey-mediated effects of the coleopteran-specific toxin Cry3Bb1 on the generalist predator *Atheta coriaria* (Coleoptera: Staphylinidae). *Bull. Entomol. Res.* **102**, 293-302.
- Garcia, M.A. and Altieri, M.A. (2005) Transgenic crops: implications for biodiversity and sustainable agriculture. *Bull. Sci. Technol. Soc.* **25**, 335-353.
- Gatehouse, A.M.R., Ferry, N., Edwards, M.G. and Bell, H.A. (2011) Insect-resistant biotech crops and their impacts on beneficial arthropods. *Phil. Trans. R. Soc B* **366**, 1438-1452.
- Glover, D. (2010a) Exploring the resilience of *Bt* cotton's 'pro-poor success story'. *Dev. Change* **41**, 955-981.
- Glover, D. (2010b) Is *Bt* cotton a pro-poor technology? A review and critique of the empirical record. *J. Agrar Change* **10**, 482-509.
- González-Cabrera, J., García, M., Hernández-Crespo, P., Farinós, G.P., Ortego, F. and Castañera, P. (2013) Resistance to *Bt* maize in *Mythimna unipuncta* (Lepidoptera: Noctuidae) is mediated by alteration in Cry1Ab protein activation. *Insect Biochem. Mol. Biol.* **43**, 635-643.
- González-Núñez, M., Ortego, F. and Castañera, P. (2000) Susceptibility of Spanish populations of the corn borers *Sesamia nonagrioides* (Lepidoptera: Noctuidae) and *Ostrinia nubilalis* (Lepidoptera: Crambidae) to a *Bacillus thuringiensis* endotoxin. *J. Econ. Entomol.* **93**, 459-463.
- González-Zamora, J.E., Camúñez, S. and Avilla, C. (2007) Effects of *Bacillus thuringiensis* Cry toxins on developmental and reproductive characteristics of the predator *Orius albidipennis* (Hemiptera: Anthocoridae) under laboratory conditions. *Environ. Entomol.* **36**, 1246-1253.
- Gray, M.E., Sappington, T.W., Miller, N.J., Moeser, J. and Bohn, M.O. (2009) Adaptation and invasiveness of western corn rootworm: intensifying research on a worsening pest. *Annu. Rev. Entomol.* **54**, 303-321.
- Groot, A.T. and Dicke, M. (2002) Insect-resistant transgenic plants in a multi-trophic context. *Plant J.* **31**, 387-406.
- Gross, K. and Rosenheim, J.A. (2011) Quantifying secondary pest outbreaks in cotton and their monetary cost with causal-inference statistics. *Ecol. Appl.* **21**, 2770-2780.
- Guedes, R.N.C. and Cutler, G.C. (2014) Insecticide-induced hormesis and arthropod pest management. *Pest Manag. Sci.* **70**, 690-697.
- Gutierrez, A.P., Adamczyk, J.J., Ponsard, S. and Ellis, C. (2006) Physiologically based demographics of *Bt* cotton–pest interactions: II. Temporal refuges, natural enemy interactions. *Ecol. Model.* **191**, 360-382.

- Hardin, M.R., Benrey, B., Coll, M., Lamp, W.O., Roderick, G.K. and Barbosa, P. (1995) Arthropod pest resurgence: an overview of potential mechanisms. *Crop. Prot.* **14**, 3-18.
- Hardke, J.T., Leonard, B.R., Huang, F. and Jackson, R. (2011) Damage and survivorship of fall armyworm (Lepidoptera: Noctuidae) on transgenic field corn expressing *Bacillus thuringiensis* Cry proteins. *Crop. Prot.* **30**, 168-172.
- Harper, C.R. (1991) Predator-prey systems in pest management. *Northeast. J. Agric. Resour. Econ.* **20**, 15-23.
- Harper, C.R. and Zilberman, D. (1989) Pest externalities from agricultural inputs. *American J. Agric. Econ.* **71**, 692-702.
- Hilbeck, A., Baumgartner, M., Fried, P.M. and Bigler, F. (1998) Effects of transgenic *Bacillus thuringiensis* corn fed prey on mortality and development time of immature *Chrysoperla carnea* (Neuroptera: Chrysopidae). *Environ. Entomol.* **27**, 480-487.
- Hilbeck, A., Weiss, G., Oehen, B., Römbke, J., Jänsch, S., Teichmann, H., Lang, A., Otto, M. and Tappeser, B. (2014) Ranking matrices as operational tools for the environmental risk assessment of genetically modified crops on non-target organisms. *Ecol. Indic.* **36**, 367-381.
- Ho, P., Zhao, J.H. and Xue, D. (2009) Access and control of agro-biotechnology: *Bt* cotton, ecological change and risk in China. *J. Peasant Stud.* **36**, 345-364.
- Hofs, J.-L., Fok, M. and Vaissayre, M. (2006) Impact of *Bt* cotton adoption on pesticide use by smallholders: A 2-year survey in Makhatini Flats (South Africa). *Crop. Prot.* **25**, 984-988.
- Holmes, T.P., Liebhold, A.M., Kovacs, K.F. and Von Holle, B. (2010) A spatial-dynamic value transfer model of economic losses from a biological invasion. *Ecol. Econ.* **70**, 86-95.
- Hooper, D.U., Chapin, F.S., Ewel, J.J., Hector, A., Inchausti, P., Lavorel, S., Lawton, J.H., Lodge, D.M., Loreau, M., Naeem, S., Schmid, B., Setälä, H., Symstad, A.J., Vandermeer, J. and Wardle, D.A. (2005) Effects of biodiversity on ecosystem functioning: a consensus of current knowledge. *Ecol. Monogr.* **75**, 3-35.
- Huang, J., Mi, J., Chen, R., Su, H., Wu, K., Qiao, F. and Hu, R. (2014) Effect of farm management practices in the *Bt* toxin production by *Bt* cotton: evidence from farm fields in China. *Transgenic Res.* **23**, 397-406.
- Huang, J., Rozelle, S., Pray, C. and Wang, Q. (2002) Plant biotechnology in China. *Science* **295**, 674-676.
- Hutchison, W.D., Burkness, E.C., Mitchell, P.D., Moon, R.D., Leslie, T.W., Fleischer, S.J., Abrahamson, M., Hamilton, K.L., Steffey, K.L., Gray, M.E., Hellmich, R.L., Kaster, L.V., Hunt, T.E., Wright, R.J., Pecinovsky, K., Rabaey, T.L., Flood, B.R. and Raun, E.S. (2010) Areawide suppression of European corn borer with *Bt* maize reaps savings to non-*Bt* maize growers. *Science* **330**, 222-225.
- Hutchison, W.D., Hunt, T.E., Hein, G.L., Steffey, K.L., Pilcher, C.D. and Rice, M.E. (2011) Genetically engineered *Bt* corn and range expansion of the Western bean cutworm (Lepidoptera: Noctuidae) in the United States: A response to Greenpeace Germany. *J. Int. Pest Manag.* **2**, B1-B8.
- Jaleel, W., Saeed, S., Naqqash, M.N. and Zaka, S.M. (2014) Survey of *Bt* cotton in Punjab Pakistan related to the knowledge, perception and practices of farmers regarding insect pests. *Int. J. Agric. Crop Sci.* **7**, 10.
- James, C. (2013) Global status of commercialised biotech/GM crops: 2013, ISAAA Brief No. 46. International Service for the Acquisition of Agri-Biotech Applications, Ithaca, NY. ISBN 978-1-892456-55-9.
- Jeyaratnam, J. (1990) Acute pesticide poisoning: a major global health problem. *World Health Statistics Quarterly* **43**, 139-144.
- Kaplan, I. and Denno, R.F. (2007) Interspecific interactions in phytophagous insects revisited: a quantitative assessment of competition theory. *Ecol. Lett.* **10**, 977-994.
- Keller, R.P., Lodge, D.M. and Finnoff, D.C. (2007) Risk assessment for invasive species produces net bioeconomic benefits. *Proc. Natl. Acad. Sci. U.S.A.* **104**, 203-207.
- Kouser, S. and Qaim, M. (2011) Impact of *Bt* cotton on pesticide poisoning in smallholder agriculture: A panel data analysis. *Ecol. Econ.* **70**, 2105-2113.
- Krebs, J.R., Wilson, J.D., Bradbury, R.B. and Siriwardena, G.M. (1999) The second silent spring? *Nature* **400**, 611-612.

- Krishna, V.V. and Qaim, M. (2012) *Bt* cotton and sustainability of pesticide reductions in India. *Agric. Syst.* **107**, 47-55.
- Kruger, M., Rensburg, J.R.J.V. and Berg, J.V.D. (2011) Resistance to *Bt* maize in *Busseola fusca* (Lepidoptera: Noctuidae) from Vaalharts, South Africa. *Environ. Entomol.* **40**, 477-483.
- Kruger, M., Van Rensburg, J. and Van den Berg, J. (2012) Transgenic *Bt* maize: farmers' perceptions, refuge compliance and reports of stem borer resistance in South Africa. *J. Appl. Entomol.* **136**, 38-50.
- Lang, A. and Otto, M. (2010) A synthesis of laboratory and field studies on the effects of transgenic *Bacillus thuringiensis* (*Bt*) maize on non-target Lepidoptera. *Entomol. Exp. Appl.* **135**, 121-134.
- Li, G., Feng, H., McNeil, J.N., Liu, B., Chen, P. and Qiu, F. (2011) Impacts of transgenic *Bt* cotton on a non-target pest, *Apolygus lucorum* (Meyer-Dür) (Hemiptera: Miridae), in northern China. *Crop. Prot.* **30**, 1573-1578.
- Li, Y. and Romeis, J. (2010) *Bt* maize expressing Cry3Bb1 does not harm the spider mite, *Tetranychus urticae*, or its ladybird beetle predator, *Stethorus punctillum*. *Biol. Control* **53**, 337-344.
- Liang, J., Tang, S. and Cheke, R.A. (2012) An integrated pest management model with delayed responses to pesticide applications and its threshold dynamics. *Nonlinear Anal. Real World Appl.* **13**, 2352-2374.
- Lindroth, E., Hunt, T.E., Skoda, S.R., Culy, M.D., Lee, D. and Foster, J.E. (2012) Population genetics of the western bean cutworm (Lepidoptera: Noctuidae) across the United States. *Ann. Entomol. Soc. Am.* **105**, 685-692.
- Lövei, G. and Arpaia, S. (2005) The impact of transgenic plants on natural enemies: a critical review of laboratory studies. *Entomol. Exp. Appl.* **114**, 1-14.
- Lövei, G.L., Andow, D.A. and Arpaia, S. (2009) Transgenic insecticidal crops and natural enemies: a detailed review of laboratory studies. *Environ. Entomol.* **38**, 293-306.
- Lu, Y., Wu, K., Jiang, Y., Guo, Y. and Desneux, N. (2012) Widespread adoption of *Bt* cotton and insecticide decrease promotes biocontrol services. *Nature* **487**, 362-365.
- Lu, Y., Wu, K., Jiang, Y., Xia, B., Li, P., Feng, H., Wyckhuys, K.A.G. and Guo, Y. (2010) Mirid bug outbreaks in multiple crops correlated with wide-scale adoption of *Bt* cotton in China. *Science* **328**, 1151-1154.
- Luckey, T. (1968) Insecticide Hormoligosis. *J. Econ. Entomol.* **61**, 7-12.
- Lundgren, J.G., Gassmann, A.J., Bernal, J., Duan, J.J. and Ruberson, J. (2009) Ecological compatibility of GM crops and biological control. *Crop. Prot.* **28**, 1017-1030.
- Maiorano, A., Cerrani, I., Fumagalli, D. and Donatelli, M. (2014) New biological model to manage the impact of climate warming on maize corn borers. *Agron. Sustain. Dev.* **34**, 609-621.
- Malvar, R., Butrón, A., Alvarez, A., Ordas, B., Soengas, P., Revilla, P. and Ordas, A. (2004) Evaluation of the European Union maize landrace core collection for resistance to *Sesamia nonagrioides* (Lepidoptera: Noctuidae) and *Ostrinia nubilalis* (Lepidoptera: Crambidae). *J. Econ. Entomol.* **97**, 628-634.
- Mancini, F., Termorshuizen, A.J., Jiggins, J.L.S. and van Bruggen, A.H.C. (2008) Increasing the environmental and social sustainability of cotton farming through farmer education in Andhra Pradesh, India. *Agric. Syst.* **96**, 16-25.
- Marino, P.C. and Landis, D.A. (1996) Effect of landscape structure on parasitoid diversity and parasitism in agroecosystems. *Ecol. Appl.* **6**, 276-284.
- Marvier, M. (2002) Improving risk assessment for nontarget safety of transgenic crops. *Ecol. Appl.* **12**, 1119-1124.
- Marvier, M., McCreedy, C., Regetz, J. and Kareiva, P. (2007) A meta-analysis of effects of *Bt* cotton and maize on nontarget invertebrates. *Science* **316**, 1475-1477.
- Matson, P.A., Parton, W.J., Power, A. and Swift, M. (1997) Agricultural intensification and ecosystem properties. *Science* **277**, 504-509.
- McDermott, S.M., Irwin, R.E. and Taylor, B.W. (2013) Using economic instruments to develop effective management of invasive species: insights from a bioeconomic model. *Ecol. Appl.* **23**, 1086-1100.

- Meinke, L.J., Sappington, T.W., Onstad, D.W., Guillemaud, T., Miller, N.J., Komáromi, J., Levay, N., Furlan, L., Kiss, J. and Toth, F. (2009) Western corn rootworm (*Diabrotica virgifera virgifera* LeConte) population dynamics. *Agric. For. Entomol.* **11**, 29-46.
- Meissle, M., Mouron, P., Musa, T., Bigler, F., Pons, X., Vasileiadis, V., Otto, S., Antichi, D., Kiss, J. and Pálincás, Z. (2010) Pests, pesticide use and alternative options in European maize production: Current status and future prospects. *J. Appl. Entomol.* **134**, 357-375.
- Meissle, M. and Romeis, J. (2009) The web-building spider *Theridion impressum* (Araneae: Theridiidae) is not adversely affected by *Bt* maize resistant to corn rootworms. *Plant Biotech. J.* **7**, 645-656.
- Meissle, M., Romeis, J. and Bigler, F. (2011) *Bt* maize and integrated pest management-a European perspective. *Pest Manag. Sci.* **67**, 1049-1058.
- Meissle, M., Vojtech, E. and Poppy, G. (2005) Effects of *Bt* maize-fed prey on the generalist predator *Poecilus cupreus* L. (Coleoptera: Carabidae). *Transgenic Res.* **14**, 123-132.
- Men, X., Ge, F., Edwards, C.A. and Yardim, E.N. (2004) The influence of pesticide applications on *Helicoverpa armigera* Hübner and sucking pests in transgenic *Bt* cotton and non-transgenic cotton in China. *Crop. Prot.* **24**, 319-324.
- Metcalfe, R. (1980) Changing role of insecticides in Crop Protection. *Annu. Rev. Entomol.* **25**, 219-256.
- Metcalfe, R. (1987) Benefit/risk considerations in the use of pesticides. *Agric. Hum. Values* **4**, 15-25.
- Michel, A.P., Krupke, C.H., Baute, T.S. and Difonzo, C.D. (2010) Ecology and management of the western bean cutworm (Lepidoptera: Noctuidae) in corn and dry beans. *J. Int. Pest Manag.* **1**, A1-A10.
- Miller, N., Estoup, A., Toepfer, S., Bourguet, D., Lapchin, L., Derridj, S., Kim, K.S., Reynaud, P., Furlan, L. and Guillemaud, T. (2005) Multiple transatlantic introductions of the western corn rootworm. *Science* **310**, 992-992.
- Morse, J.G. (1998) Agricultural implications of pesticide-induced hormesis of insects and mites. *Hum. Exp. Toxicol.* **17**, 266-269.
- Morse, S., Bennett, R. and Ismael, Y. (2007) Inequality and GM crops: A case-study of *Bt* cotton in India. *AgBioForum* **10**, 44-50.
- Morse, S., Bennett, R.M. and Ismael, Y. (2005) Genetically modified insect resistance in cotton: some farm level economic impacts in India. *Crop. Prot.* **24**, 433-440.
- Musser, F.R. and Shelton, A.M. (2003) *Bt* sweet corn and selective insecticides: Impacts on pests and predators. *J. Econ. Entomol.* **96**, 71-80.
- Nagrare, V., Kranthi, S., Biradar, V., Zade, N., Sangode, V., Kakde, G., Shukla, R., Shivare, D., Khadi, B. and Kranthi, K. (2009) Widespread infestation of the exotic mealybug species, *Phenacoccus solenopsis* (Tinsley)(Hemiptera: Pseudococcidae), on cotton in India. *Bull. Entomol. Res.* **99**, 537-541.
- Naranjo, S.E. (2005) Long-term assessment of the effects of transgenic *Bt* cotton on the abundance of nontarget arthropod natural enemies. *Environ. Entomol.* **34**, 1193-1210.
- Naranjo, S.E. (2009) Impacts of *Bt* crops on non-target invertebrates and insecticide use patterns. *CAB Reviews: perspectives in agriculture, veterinary science, nutrition and natural resources* **4**, 1-11.
- Naranjo, S.E. (2011) Impacts of *Bt* transgenic cotton on integrated pest management. *J. Agric. Food Chem.* **59**, 5842.
- Naranjo, S.E. and Ellsworth, P.C. (2009) The contribution of conservation biological control to integrated control of *Bemisia tabaci* in cotton. *Biol. Control* **51**, 458-470.
- Nguyen, H.T. and Jehle, J.A. (2009) Expression of Cry3Bb1 in transgenic corn MON88017. *J. Agric. Food Chem.* **57**, 9990-9996.
- Obrist, L.B., Dutton, A., Albajes, R. and Bigler, F. (2006) Exposure of arthropod predators to Cry1Ab toxin in *Bt* maize fields. *Ecol. Entomol.* **31**, 143-154.
- Oerke, E.-C. (2006) Crop losses to pests. *J. Agric. Sci.* **144**, 31-43.
- Park, J., McFarlane, I., Phipps, R. and Ceddia, G. (2011) The role of transgenic crops in sustainable development. *Plant Biotech. J* **9**, 2-21.
- Pemsl, D. and Waibel, H. (2007) Assessing the profitability of different Crop. Prot. strategies in cotton: Case study results from Shandong Province, China. *Agric. Syst.* **95**, 28-36.

- Pemsl, D.E., Voelker, M., Wu, L. and Waibel, H. (2011) Long-term impact of *Bt* cotton: findings from a case study in China using panel data. *Int. J. Agr. Sustain.* **9**, 508-521.
- Pérez-Hedo, M., López, C., Albajes, R. and Eizaguirre, M. (2012) Low susceptibility of non-target Lepidopteran maize pests to the *Bt* protein Cry1Ab. *Bull. Entomol. Res.* **102**, 737.
- Petrovskii, S., Petrovskaya, N. and Bearup, D. (2014) Multiscale approach to pest insect monitoring: Random walks, pattern formation, synchronization, and networks. *Phys. Life Rev.* 467-525.
- Pilcher, C.D., Rice, M.E., Obrycki, J.J. and Lewis, L.C. (1997) Field and laboratory evaluations of transgenic *Bacillus thuringiensis* corn on secondary lepidopteran pests (Lepidoptera: Noctuidae). *J. Econ. Entomol.* **90**, 669-678.
- Pons, X., Lumbierres, B., Lopez, C. and Albajes, R. (2005) Abundance of non-target pests in transgenic *Bt*-maize: A farm scale study. *European Journal of Entomology* **102**, 73.
- Poppy, G.M. and Sutherland, J.P. (2004) Can biological control benefit from genetically-modified crops? Tritrophic interactions on insect-resistant transgenic plants. *Physiol. Entomol.* **29**, 257-268.
- Pray, C., Huang, J., Hu, R. and Rozelle, S. (2002) Five years of *Bt* cotton in China—the benefits continue. *Plant J.* **31**, 423-430.
- Qaim, M. (2003) *Bt* cotton in India: Field trial results and economic projections. *World Dev.* **31**, 2115-2127.
- Qaim, M., Subramanian, A. and Sadashivappa, P. (2009) Commercialized GM crops and yield. *Nat. Biotech.* **27**, 803-804.
- Qaim, M. and Zilberman, D. (2003) Yield effects of genetically modified crops in developing countries. *Science* **299**, 900-902.
- Ramaswami, B., Pray, C.E. and Lalitha, N. (2012) The spread of illegal transgenic cotton varieties in India: Biosafety regulation, monopoly, and enforcement. *World Dev.* **40**, 177-188.
- Razze, J.M. and Mason, C.E. (2012) Dispersal behavior of neonate European corn borer (Lepidoptera: Crambidae) on *Bt* corn. *J. Econ. Entomol.* **105**, 1214-1223.
- Riesgo, L., Areal, F. and Rodriguez-Cerezo, E. (2012) How can specific market demand for non-GM maize affect the profitability of *Bt* and conventional maize? A case study for the middle Ebro Valley, Spain. *Span. J. Agric. Res.* **10**, 867-876.
- Ripper, W. (1956) Effect of pesticides on balance of arthropod populations. *Annu. Rev. Entomol.* **1**, 403-438.
- Romeis, J., Dutton, A. and Bigler, F. (2004) *Bacillus thuringiensis* toxin (Cry1Ab) has no direct effect on larvae of the green lacewing *Chrysoperla carnea* (Stephens) (Neuroptera: Chrysopidae). *J. Insect Physiol.* **50**, 175-183.
- Romeis, J., Meissle, M. and Bigler, F. (2006) Transgenic crops expressing *Bacillus thuringiensis* toxins and biological control. *Nat. Biotechnol.* **24**, 63-71.
- Romeis, J., Meissle, M., Raybould, A. and Hellmich, R. (2009) Impact of insect-resistant transgenic crops on above-ground non-target arthropods. In: *Environmental impact of genetically modified crops* (Ferry, N. and Gatehouse, A.M.R. eds), pp. 165-198. UK: CAB International.
- Sanglestawai, S., Rejesus, R.M. and Yorobe, J.M. (2014) Do lower yielding farmers benefit from *Bt* corn? Evidence from instrumental variable quantile regressions. *Food Policy* **44**, 285-296.
- Sanvido, O., Romeis, J. and Bigler, F. (2009) An approach for post-market monitoring of potential environmental effects of *Bt*-maize expressing Cry1Ab on natural enemies. *J. Appl. Entomol.* **133**, 236-248.
- Schnurr, M.A. (2012) Inventing Makhathini: Creating a prototype for the dissemination of genetically modified crops into Africa. *Geoforum* **43**, 784-792.
- Shantharam, S., Sullia, S. and Shivakumara Swamy, G. (2008) Peer review contestations in the era of transgenic crops. *Curr. Sci.* **95**, 25.
- Sharma, H. and Ortiz, R. (2000) Transgenics, pest management, and the environment. *Curr. Sci.* **79**, 421-437.
- Shelton, A.M., Naranjo, S.E., Romeis, J., Hellmich, R.L., Wolt, J.D., Federici, B.A., Albajes, R., Bigler, F., Burgess, E.P. and Dively, G.P. (2009) Setting the record straight: a rebuttal to an erroneous analysis on transgenic insecticidal crops and natural enemies. *Transgenic Res.* **18**, 317-322.

- Shi, G., Chavas, J.-P. and Lauer, J. (2013) Commercialized transgenic traits, maize productivity and yield risk. *Nat. Biotech.* **31**, 111-114.
- Shivankar, V.J., Shyam, S. and Rao, C.N. (2007) Secondary pest resurgence. In: *Encyclopedia of Pest Management, Volume II* (Pimentel, D. ed) pp. 597–601. CRC Press.
- Showalter, A.M., Heuberger, S., Tabashnik, B.E. and Carrière, Y. (2009) A primer for using transgenic insecticidal cotton in developing countries. *J. Insect Sci.* **9**, 1-39.
- Sisterson, M.S., Carrière, Y., Dennehy, T.J. and Tabashnik, B.E. (2005) Evolution of resistance to transgenic crops: interactions between insect movement and field distribution. *J. Econ. Entomol.* **98**, 1751-1762.
- Sisterson, M.S., Carrière, Y., Dennehy, T.J. and Tabashnik, B.E. (2007) Nontarget effects of transgenic insecticidal crops: Implications of source-sink population dynamics. *Environ. Entomol.* **36**, 121-127.
- Smale, M. (2012) Rough terrain for research: studying early adopters of biotech crops. *AgBioForum* **15**, 114-124.
- Smale, M., Zambrano, P., & Cartel, M. (2006) Bales and balance: A review of the methods used to assess the economic impact of *Bt* cotton on farmers in developing economies. *AgBioForum* **9**, 195-212.
- Snow, A.A., Andow, D.A., Gepts, P., Hallerman, E.M., Power, A., Tiedje, J.M. and Wolfenbarger, L. (2005) Genetically engineered organisms and the environment: Current status and recommendations. *Ecol. Appl.* **15**, 377-404.
- Snyder, W.E., Snyder, G.B., Finke, D.L. and Straub, C.S. (2006) Predator biodiversity strengthens herbivore suppression. *Ecol. Lett.* **9**, 789-796.
- Stephens, E.J., Losey, J.E., Allee, L.L., DiTommaso, A., Bodner, C. and Breyre, A. (2012) The impact of Cry3Bb *Bt*-maize on two guilds of beneficial beetles. *Agric. Ecosyst. Environ.* **156**, 72-81.
- Stone, G.D. (2011) Field versus farm in Warangal: *Bt* cotton, higher yields, and larger questions. *World Dev.* **39**, 387-398.
- Storer, N.P., Van Duyn, J.W. and Kennedy, G.G. (2001) Life history traits of *Helicoverpa zea* (Lepidoptera: Noctuidae) on non-*Bt* and *Bt* transgenic corn hybrids in Eastern North Carolina. *J. Econ. Entomol.* **94**, 1268-1279.
- Sujii, E.R., Togni, P.H.B., de A Ribeiro, P., de A Bernardes, T., Milane, P., Paula, D.P., Pires, C.S.S. and Fontes, E.M.G. (2013) Field evaluation of *Bt* cotton crop impact on nontarget pests: Cotton aphid and boll weevil. *Neotrop. Entomol.* **42**, 102-111.
- Symstad, A.J., Chapin, F.S., Wall, D.H., Gross, K.L., Huenneke, L.F., Mittelbach, G.G., Peters, D.P.C. and Tilman, D. (2003) Long-term and large-scale perspectives on the relationship between biodiversity and ecosystem functioning. *BioScience* **53**, 89-98.
- Tabashnik, B.E., Carrière, Y., Dennehy, T.J., Morin, S., Sisterson, M.S., Roush, R.T., Shelton, A.M. and Zhao, J.-Z. (2003) Insect resistance to transgenic *Bt* crops: Lessons from the laboratory and field. *J. Econ. Entomol.* **96**, 1031-1038.
- Tabashnik, B.E., Gassmann, A.J., Crowder, D.W. and Carrière, Y. (2008) Insect resistance to *Bt* crops: evidence versus theory. *Nat. Biotechnol.* **26**, 199-202.
- Thirtle, C., Beyers, L., Ismael, Y. and Piesse, J. (2003) Can GM-technologies help the poor? The impact of *Bt* cotton in Makhathini Flats, KwaZulu-Natal. *World Dev.* **31**, 717-732.
- Tilman, D., Fargione, J., Wolff, B., D'Antonio, C., Dobson, A., Howarth, R., Schindler, D., Schlesinger, W.H., Simberloff, D. and Swackhamer, D. (2001) Forecasting agriculturally driven global environmental change. *Science* **292**, 281-284.
- Traxler, G. and Godoy-Avila, S. (2004) Transgenic cotton in Mexico. *AgBioForum* **7**, 57-62.
- Truter, J., Hamburg, H.V. and Berg, J.V.D. (2014) Comparative diversity of arthropods on *Bt* maize and non-*Bt* maize in two different cropping systems in South Africa. *Environ. Entomol.* **43**, 197-208.
- Tscharntke, T., Klein, A.M., Kruess, A., Steffan-Dewenter, I. and Thies, C. (2005) Landscape perspectives on agricultural intensification and biodiversity–ecosystem service management. *Ecol. Lett.* **8**, 857-874.
- Van den Berg, J. and Van Wyk, A. (2007) The effect of *Bt* maize on *Sesamia calamistis* in South Africa. *Entomol. Exp. Appl.* **122**, 45-51.

- van den Bosch, R. (1978) The pesticide conspiracy. *University of California Press, Berkeley*, p. 226.
- Van Wyk, A., Van den Berg, J. and Van Hamburg, H. (2007) Selection of non-target Lepidoptera species for ecological risk assessment of *Bt* maize in South Africa. *Afr. Entomol.* **15**, 356-366.
- Van Wyk, A., Van den Berg, J. and Van Hamburg, H. (2008) Diversity and comparative phenology of Lepidoptera on *Bt* and non-*Bt* maize in South Africa. *Int. J. Pest Manag.* **54**, 77-87.
- Van Wyk, A., Van den Berg, J. and Van Rensburg, J. (2009) Comparative efficacy of *Bt* maize events MON810 and *Bt*11 against *Sesamia calamistis* (Lepidoptera: Noctuidae) in South Africa. *Crop. Prot.* **28**, 113-116.
- Vasileiadis, V.P., Sattin, M., Otto, S., Veres, A., Pálincás, Z., Ban, R., Pons, X., Kudsk, P., van der Weide, R., Czembor, E., Moonen, A.C. and Kiss, J. (2011) Crop. Prot. in European maize-based cropping systems: Current practices and recommendations for innovative Integrated Pest Management. *Agric. Syst.* **104**, 533-540.
- Velasco Pazos, P., Revilla Temiño, P., Monetti, L., Butrón Gómez, A.M., Ordás Pérez, A. and Malvar Pintos, R.A. (2007) Corn borers (Lepidoptera: Noctuidae; Crambidae) in northwestern Spain: population dynamics and distribution. *Maydica* **52**, 195-203.
- Virla, E.G., Casuso, M. and Frias, E.A. (2010) A preliminary study on the effects of a transgenic corn event on the non-target pest *Dalbulus maidis* (Hemiptera: Cicadellidae). *Crop. Prot.* **29**, 635-638.
- Vitousek, P.M., Mooney, H.A., Lubchenco, J. and Melillo, J.M. (1997) Human domination of Earth's ecosystems. *Science* **277**, 494-499.
- Waage, J.K. and Mumford, J.D. (2008) Agricultural biosecurity. *Phil. Trans. R. Soc. Lond. B* **363**, 863-876.
- Wilby, A. and Thomas, M.B. (2002) Natural enemy diversity and pest control: patterns of pest emergence with agricultural intensification. *Ecol. Lett.* **5**, 353-360.
- Wilson, L., Downes, S., Khan, M., Whitehouse, M., Baker, G., Grundy, P. and Maas, S. (2013) IPM in the transgenic era: a review of the challenges from emerging pests in Australian cotton systems. *Crop Pasture Sci.* **64**, 737-749.
- Wolfenbarger, L.L., Naranjo, S.E., Lundgren, J.G., Bitzer, R.J. and Watrud, L.S. (2008) *Bt* crop effects on functional guilds of non-target arthropods: a meta-analysis. *PLoS One* **3**, e2118.
- Wolfenbarger, L.L. and Phifer, P.R. (2000) The ecological risks and benefits of genetically engineered plants. *Science* **290**, 2088-2093.
- Wu, K. and Guo, Y. (2005) The evolution of cotton pest management practices in China. *Annu. Rev. Entomol.* **50**, 31-52.
- Wu, K., Li, W., Feng, H. and Guo, Y. (2002) Seasonal abundance of the mirids, *Lygus lucorum* and *Adelphocoris* spp. (Hemiptera: Miridae) on *Bt* cotton in northern China. *Crop. Prot.* **21**, 997-1002.
- Xu, N., Fok, M., Bai, L. and Zhou, Z. (2008) Effectiveness and chemical pest control of *Bt*-cotton in the Yangtze River Valley, China. *Crop. Prot.* **27**, 1269-1276.
- Yang, J., Wang, Z.-R., Yang, D.-L., Yang, Q., Yan, J. and He, M.-F. (2009) Ecological risk assessment of genetically modified crops based on cellular automata modeling. *Biotechnol. Adv.* **27**, 1132-1136.
- Yang, P., Iles, M., Yan, S. and Jolliffe, F. (2005a) Farmers' knowledge, perceptions and practices in transgenic *Bt* cotton in small producer systems in Northern China. *Crop. Prot.* **24**, 229-239.
- Yang, P., Li, K., Shi, S., Xia, J., Guo, R., Li, S. and Wang, L. (2005b) Impacts of transgenic *Bt* cotton and integrated pest management education on smallholder cotton farmers. *Int. J. Pest Manag.* **51**, 231-244.
- Zeilinger, A.R., Olson, D.M. and Andow, D.A. (2011) Competition between stink bug and heliothine caterpillar pests on cotton at within-plant spatial scales. *Entomol. Exp. Appl.* **141**, 59-70.
- Zhao, J.-Z., Cao, J., Collins, H.L., Bates, S.L., Roush, R.T., Earle, E.D. and Shelton, A.M. (2005) Concurrent use of transgenic plants expressing a single and two *Bacillus thuringiensis* genes speeds insect adaptation to pyramided plants. *Proc. Natl. Acad. Sci. U.S.A.* **102**, 8426-8430.
- Zhao, J.H., Ho, P. and Azadi, H. (2011) Benefits of *Bt* cotton counterbalanced by secondary pests? Perceptions of ecological change in China. *Environ. Monit. Assess.* **173**, 985-994.

