

Recommendations on multiple testing adjustment in multi-arm trials with a shared control group

Article

Accepted Version

Howard, D. R., Brown, J. M., Todd, S. ORCID: <https://orcid.org/0000-0002-9981-923X> and Gregory, W. M. (2018) Recommendations on multiple testing adjustment in multi-arm trials with a shared control group. *Statistical Methods in Medical Research*, 27 (5). pp. 1513-1530. ISSN 0962-2802 doi: 10.1177/0962280216664759 Available at <https://centaur.reading.ac.uk/66407/>

It is advisable to refer to the publisher's version if you intend to cite from the work. See [Guidance on citing](#).

To link to this article DOI: <http://dx.doi.org/10.1177/0962280216664759>

Publisher: Sage

All outputs in CentAUR are protected by Intellectual Property Rights law, including copyright law. Copyright and IPR is retained by the creators or other copyright holders. Terms and conditions for use of this material are defined in the [End User Agreement](#).

www.reading.ac.uk/centaur

CentAUR

Central Archive at the University of Reading

Reading's research outputs online

Recommendations on multiple testing adjustment in multi-arm trials with a shared control group

Dena R Howard¹, Julia M Brown¹, Susan Todd², Walter M Gregory¹

¹Leeds Institute of Clinical Trials Research, University of Leeds, Leeds, United Kingdom

²Department of Mathematics and Statistics, University of Reading, Reading, United Kingdom

Correspondence to: Dena Howard

Author contact details:

Dena Howard

Leeds Institute of Clinical Trials Research

University of Leeds

Leeds

LS2 9JT

United Kingdom

Email: D.R.Cohen@leeds.ac.uk

Tel: 0113 343 1475

Fax: 0113 343 1471

Recommendations on multiple testing adjustment in multi-arm trials with a shared control group

Short Title

Recommendations on multiple testing adjustment in multi-arm trials

Abstract

Multi-arm clinical trials assessing multiple experimental treatments against a shared control group can offer efficiency advantages over independent trials through assessing an increased number of hypotheses. Published opinion is divided on the requirement for multiple testing adjustment to control the familywise type-I error rate (FWER). The probability of a false positive error in multi-arm trials compared to equivalent independent trials is affected by the correlation between comparisons due to sharing control data. We demonstrate that this correlation in fact leads to a reduction in the FWER, therefore FWER adjustment is not recommended solely due to sharing control data. In contrast, the correlation increases the probability of multiple false positive outcomes across the hypotheses, although standard FWER adjustment methods do not control for this. A stringent critical value adjustment is proposed to maintain equivalent evidence of superiority in two correlated comparisons to that obtained within

independent trials. FWER adjustment is only required if there is an increased chance of making a single claim of effectiveness by testing multiple hypotheses; not due to sharing control data. For competing experimental therapies, the correlation between comparisons can be advantageous as it eliminates bias due to the experimental therapies being compared to different control populations.

Keywords

multiple testing; multi-arm clinical trial; familywise error rate; type-I error; multiplicity; shared control group; correlated comparisons; alpha adjustment

1. Introduction

A typical confirmatory two arm trial would usually compare an experimental treatment against the current standard within the population of interest. For several reasons, it is advantageous to conduct multi-arm trials, in which a number of experimental treatments are compared to the current standard. Firstly, such trials are more efficient since they use the data collected on the control group more than once so fewer patients are required. Secondly, trial set-up times and costs can be reduced over running separate trials. Finally, increasing the number of experimental arms increases the chance of finding a successful treatment¹.

When testing a hypothesis in a phase III clinical trial, the chance of a false positive result, known as the type-I error, is required to be stringently controlled. The convention is to set this error, usually denoted by α , to be no greater than 5% by setting the one-sided p-value to <0.025 . If more than one hypothesis is to be tested within a set, or family, of hypotheses, the chance of a false positive conclusion occurring anywhere within that set is known as the familywise error rate (FWER). That is, the FWER is the probability of at least one false positive conclusion being declared anywhere within a family of hypotheses.

There are conflicting viewpoints within the literature on whether the relevant errors to control in the case of multi-arm trials are the individual type-I errors for each hypothesis, known as the pairwise error rates, or the overall FWER for all hypotheses combined. The crux of the issue is how ‘family’ should be defined; whether all hypotheses belong to a family simply because they share a protocol and control group, or whether a family is a set of hypotheses that are related in that they contribute towards a single claim of effectiveness.

The literature on requirement for multiplicity adjustment is often based on philosophical opinions, rather than statistical theory considering the actual effect on the type-I error rates of using a shared control group compared to running independent trials. Some literature, including a points to consider document from the European Agency for the Evaluation of Medicinal Products (EMA)², advocates strong control of the FWER for confirmatory claims because the hypotheses are being tested within a single experiment, regardless of the relatedness of the hypotheses^{3, 4}. Others argue that adjustment is not required in all cases, particularly although not exclusively, where the experimental arms do not contribute towards a single claim of effectiveness, because the design is essentially just running a number of different trials⁵⁻⁹. Although most of the literature agrees that the need for adjustment should be considered and justified in each case, including the International Conference on Harmonisation (ICH) E9 guidance of

Statistical Principles for Clinical Trials¹⁰, no literature has been found to give comprehensive guidance from a quantitative, rather than philosophical, perspective on which situations require type-I error adjustment and which do not.

Discussion points from the Statisticians in the Pharmaceutical Industry multiplicity expert group state that “the concern with multiplicity is that, if it is not properly handled, unsubstantiated claims for the effectiveness of a drug may be made as a consequence of an inflated rate of false positive conclusions”¹¹. In this manuscript, we therefore break down and quantitatively investigate the aspects of a multi-arm trial that affect the chance of different types of false positive errors on a claim of effectiveness in order to make informed recommendations on the need for adjustment. We begin by providing examples in Section 2, and describing the background to multiplicity concerns in multi-arm trials in Section 3. In Section 4 we investigate the effect of the shared control data on the chance of type-I errors. Section 5 shows the probabilities of type-I errors after applying common multiplicity adjustment methods in the case of shared control data, and we compare these to the errors had the hypotheses been assessed within independent trials. In Section 6 we suggest a correction to control for the increased probability of multiple type-I errors that advantage the experimental treatment, which may be necessary where more than one superior outcome within the

protocol could inform the same claim of effectiveness. The paper concludes with a discussion in Section 7.

2. Motivational examples

Three examples of multi-arm trials with different types of design and varying levels of relatedness between the hypotheses are considered in this article.

2.1 MRC COIN

The phase III MRC COIN trial¹² in previously untreated patients with colorectal cancer had three-arms and two primary hypotheses. The control treatment was chemotherapy with oxaliplatin and fluoropyrimidine (OxFP) given continuously (arm *Z*). One experimental arm included an additional therapy cetuximab to OxFP (arm *A*), and the other assessed the chemotherapy OxFP given intermittently (arm *B*). Patients were randomised to the three treatment arms with a 1:1:1 ratio, and the trial objective was to assess a difference in overall survival at two years for each of the comparisons, arm *A* vs *Z* and arm *B* vs *Z*.

2.2 AMAGINE-1

The phase III AMAGINE-1 trial (clinicaltrials.gov identifier: NCT01708590) was run by Amgen / AstraZeneca from 2012 to 2015. The trial assessed the safety and efficacy of brodalumab taken every two weeks via subcutaneous injection at two doses (140 mg or 210 mg) compared with placebo in patients with moderate-to-severe plaque psoriasis. The primary hypotheses concerned the efficacy of brodalumab compared to placebo, as assessed by Static Physician Global Assessment (sPGA) score and improvement in Psoriasis Area and Severity Index (PASI) at 12 weeks.

2.3 Myeloma XI+ Intensive

The Myeloma XI Intensive trial (ClinicalTrials.gov Identifier: NCT01554852) at the University of Leeds opened to recruitment in 2010, comparing the current standard therapy CTD (cyclophosphamide, thalidomide and dexamethasone) with CRD (cyclophosphamide, lenalidomide and dexamethasone) in terms of progression-free survival (PFS) in newly diagnosed patients with Multiple Myeloma. It was anticipated that recruitment would take up to four years, with the required number of events occurring within three years after the close of recruitment. During recruitment, early evidence suggested a new therapy, carfilzomib, added to the existing CRD regime (CCRD) might improve efficacy. Since it was of interest to assess CCRD as soon as possible, the follow-on Myeloma XI+ intensive trial was designed without waiting for

the results of the original trial, and opened to recruitment in 2013 following on seamlessly from Myeloma XI. The Myeloma XI+ trial therefore compared the experimental therapy CCRD to the current standard control CTD and the previous experimental therapy CRD at a 2:1:1 randomisation in order to protect the trial in the case that CRD was found superior and superseded CTD as the standard therapy before the amended trial had completed and reported.

3. Background to multiplicity concerns in multi-arm trials

3.1 Shared control data

If two experimental treatments, say A and B , are to be compared against the current standard, say Z , in independent trials, it is accepted that there is no requirement for multiple testing adjustment. If these two hypotheses are instead assessed within the same protocol, where the data remain entirely independent and non-overlapping with separate control groups Z_1 and Z_2 , and the hypotheses are both powered separately and appropriately (figure 1a), it would be difficult to argue for multiple testing adjustment since there is no multiple use of any data. The sharing of a protocol or even a randomisation system in this case does not affect the probability of an error over that for independent trials. Westfall et al.¹³ report that it may be plausible that multiplicity

problems due to sharing a protocol could result from “selection effects” such as the method of assessment of the primary endpoint. However, this could just as easily occur in two independent trials led by the same trials team. Therefore it would seem that there is no additional reason for multiplicity concerns due to simply sharing a protocol, when separate pieces of confirmatory evidence are not required to be obtained from distinct teams.

[Insert Figure 1a]

Figure 1a. *Illustration of two separate hypotheses being tested within the same protocol. There is no overlap of the use of patients, so the questions are entirely independent. R denotes the point of randomisation.*

A trial of the design displayed in figure 1a may not make practical sense where the eligibility criteria and control group for both experimental treatment comparisons are the same. Efficiency can be greatly improved by comparing both experimental arms to the same group of control patients. If the treatment difference being sought is the same, then utilising a single control group offers a saving of 25% of the trial sample size for an even allocation ratio (figure 1b).

[Insert Figure 1b]

Figure 1b. *Illustration of a multi-arm design where two separate hypotheses are being asked within the same protocol and sharing the same control patients. R denotes the point of randomisation.*

For the design shown in figure 1b, the hypotheses can be addressed separately and can both be adequately powered. Given the logic that the use of the same protocol does not cause multiplicity concerns over the same hypotheses being tested in independent trials, the difference is around the shared use of the control data. The comparisons are no longer independent, but are correlated based on the shared comparator group¹⁴. The impact of this correlation can be formally quantified to inform the necessary adjustment so that the chances of errors do not exceed those for independent trials. Proschan et al.¹⁵ examine the effects of treatments being compared to a control within the same trial compared to independent trials, “in terms of the different distributions of the number of Type-I errors and power”. Senn¹⁶ considers the conditional probability of a type-I error under the null hypothesis of “concluding that a given dose is significant given that all other doses tested to date are significantly different from placebo” when comparing multiple doses against a placebo, as an alternate way of thinking about the probability of multiple errors within a family. He notes that “even where the probabilities of making at least one type-I error are controlled, conditional error rates may not be”, but does not make recommendations for multiple testing adjustment based on this. Very little

literature has been published assessing the effect of shared control data on the probabilities of type-I errors over those in independent trials, and this effect is rarely considered when assessing the requirement for multiplicity adjustment in multi-arm trials. Extending the work of Proschan et al. we further investigate the effect of correlation due to shared control data in detail within this manuscript.

3.2 Increased chance of making a single claim of effectiveness for a therapy

Section 3.1 highlights that a key statistical implication of running a single multi-arm trial compared to separate trials is due to multiple use of shared control data. However, another factor that could increase the chance of a false conclusion over that for independent trials is the ability to test more hypotheses than would otherwise have been assessed. The necessity for adjustment in this case is a largely philosophical, rather than necessarily statistical, argument that has been well addressed in the literature, albeit with varying opinions, and needs to be considered and justified on a trial-by-trial basis⁴⁻⁸.

On reviewing the literature, our opinion is that if the hypotheses contribute towards a single claim of effectiveness, for example because they assess different doses of the same therapy with any success leading to promotion of that therapy, the hypotheses are likely to be considered a ‘family’ and therefore FWER adjustment may be required. If

the hypotheses inform different claims of effectiveness, for example because they are assessing different experimental therapies, FWER control is likely to be an unnecessary penalty^{5, 6}. This does not contradict ICH E9 ‘Statistical Principles for Clinical Trials’¹⁰, which states that “adjustment should always be considered, and the details of any adjustment procedure or an explanation of why adjustment is not thought to be necessary should be set out in the analysis plan.” Hung and Wang¹⁷ discuss defining “a relevant family of hypotheses for which the type-I error needs to be properly controlled”, and recommend a “clinical decision tree”, determined in advance, to decide what aspects need to be protected from type-I error inflation. The decision on adjustment due to assessing multiple hypotheses should be made at the design stages for each trial and documented with full justification. This important consideration is incorporated into the recommendations on adjustment in multi-arm trials given in Section 7.

4. The effect of correlation due to shared control data

As noted above, whilst the increased chance of making a single claim of effectiveness based on multiple hypotheses within a trial has been widely discussed in the literature, the effect of correlation due to multiple use of the shared control data has been less well addressed and does not appear to be widely understood, and therefore forms the main focus of this manuscript.

We begin by stating some key definitions before exploring the quantification of the effect of the shared control data.

4.1 Definitions of error regions

In two independent hypothesis tests, such as illustrated in the trial design given in figure 1a, the null hypothesis H_{0A} assesses therapy A against Z_1 , with test statistic X_A , and the null hypothesis H_{0B} assesses therapy B against Z_2 , with test statistic X_B , each with a two-sided significance level of 0.05. It can be assumed that the test statistics for each comparison follow a normal distribution when sample sizes are reasonable.

Figure 2 illustrates the joint density for the standardised test statistics in this setting, based on the probability density function of the standardised bivariate normal distribution with no correlation, since the tests are independent. The rejection regions for the hypothesis tests are the shaded areas around the outside of the square, as described by Fernandes and Stone¹⁴.

The probability of falling within just one of the four shaded rejection regions along the length of the edges is 2.5%. That is, the probability of concluding that either therapy is

either falsely inferior or falsely superior to its control therapy is 2.5%. The darker shaded corner regions represent the probability that both hypotheses have false positive outcomes, that is there are two type-I errors.

[Insert Figure 2]

Figure 2. *Rejection regions for two independent comparisons plotted on orthogonal axes, with the standardised test statistic for the null hypothesis H_{0A} being displayed horizontally, and H_{0B} displayed vertically.*

The amount of false positive error that falls in different shaded regions within figure 2 may have different implications depending on the goals of the trial. We have therefore defined various types of false positive error below, and these will first be quantified in the case of independent comparisons before exploring the case where there is shared control data.

Familywise Error Rate (FWER):

Recall that the FWER is the overall probability of at least one false positive conclusion anywhere within a defined set of trial hypotheses. It can be seen from figure 2 that in the case of two independent comparisons and a two-sided significance level of 0.05, if we

consider the two comparisons as a family, the FWER is equal to the total shaded region. This can be calculated by the sum of the four shaded regions around the edges, minus the darker shaded regions that are double counted in the four corners.

$$FWER = (4 * 0.025) - (4 * 0.025^2) = 0.0975$$

Note that the FWER can easily be confirmed for independent comparisons, as it can be described using a binomial distribution since each null hypothesis has a binary outcome associated with it. Define Y to be the random variable associated with the event that a type-I error occurs. In the independent case, with k comparisons and a probability α of finding a significant difference, the probability of exactly y type-I errors across the k comparisons ($y = 1, \dots, k$) can be expressed as:

$$P(Y = y) = \binom{k}{y} \alpha^y (1 - \alpha)^{k-y}$$

Since the FWER is the probability of at least one error,

$$FWER = p(Y > 0) = 1 - p(Y = 0) = 1 - (1 - \alpha)^k$$

So, with two independent comparisons and $\alpha = 0.05$ for each as illustrated in figure 2, the FWER is 0.0975, as expected.

Family Multiple Error Rate (FMER):

A second type of false positive error can be defined as the chance of multiple false positive findings across a family of hypotheses, which we call the Family Multiple Error Rate (FMER).

The overall error that exists in any family of hypotheses will always equal the sum of the errors for each hypothesis. With a family of two (null) hypotheses H_{0A} and H_{0B} respectively relating to the comparisons of therapies *A* and *B* with control, and $\alpha=0.05$ for each, the total error will be 0.1. By probability theory:

$$P(H_{0A}) + P(H_{0B}) = P(H_{0A} \cup H_{0B}) + P(H_{0A} \cap H_{0B})$$

$P(H_{0A})$ is the probability of a type-I error for the null hypothesis H_{0A} .

$P(H_{0B})$ is the probability of a type-I error for the null hypothesis H_{0B} .

$P(H_{0A} \cup H_{0B})$ is the overall chance of a type-I error, i.e. the FWER.

$P(H_{0A} \cap H_{0B})$ is the chance of more than one error occurring from the same pair of null hypotheses, which is the FMER.

In figure 2, the FMER is represented by the sum of the probabilities in the four dark shaded corner regions. In the case of two fully independent hypotheses tested in two separate trials, the FMER is $4 * 0.025^2 = 0.0025$. Therefore FWER + FMER is 0.1, as expected.

Note that the FMER is directly related to the conditional probability of a type-I error $P(H_{0B}|H_{0A})$, as discussed in Section 3.1, since:

$$P(H_{0B}|H_{0A}) = \frac{P(H_{0A} \cap H_{0B})}{P(H_{0A})}$$

However, the FMER is easier to interpret in the case of a multi-arm trial due to the hypotheses not necessarily having any sensible order.

Multiple Superior False Positives (MSFP):

In figure 2, the lower left corner signifies both false positives falling in the rejection region in favour of the control, thus declaring the experimental treatment significantly inferior (multiple inferior false positive outcomes), the upper right corner signifies both false positives falling in favour of the experimental treatments (multiple superior false positive (MSFP) outcomes), and the upper left and lower right corners signify one false positive favouring the control and the other an experimental treatment. The chance of MSFP errors could be important if the outcomes of the hypotheses inform a single claim of effectiveness, as discussed in Section 6. In the independent case with two hypotheses, the probability of a MSFP outcome is $0.025^2 = 0.000625$.

4.2 The effect of positive correlation due to the shared control group

Recall Figure 1b illustrating a typical three-arm trial design with two experimental arms, A and B , and a shared control group Z . Since both treatments are being compared to the same control data, the comparisons are not independent. That is, if the control group sample, by chance, perform worse than the true population, there is an increased probability that both therapies A and B will report a false positive outcome to conclude that they are superior. The test statistics are therefore positively correlated, since the outcomes for the control sample will affect both in the same way.

It has previously been shown that the positive correlation between the test statistics reduces the probability of making at least one type-I error (the FWER) over cases where there is no correlation, such as in independent trials; but the probability of making two or more errors is higher, such that “the conditional probability of a Type-I error on one comparison with control, given that a Type-I error has been made on another comparison with control, is substantially increased” in the correlated case^{15, 16}. We further quantify and expand on these findings in order to inform recommendations on the need for a multiple testing adjustment in multi-arm trials with correlated test statistics.

4.3 Calculating the correlation between the test statistics due to sharing control data

Recall from Section 4.1 that in the case of multi-arm trials with independent experimental therapies and a shared control group, the test statistics for the comparisons, X_A and X_B , can be assumed to follow standardised normal distributions when sample sizes are reasonable. Their joint distribution therefore follows a standardised bivariate normal with correlation ρ_{AB} . Relevant theory is given in Follmann et al.¹⁸.

Dunnett¹⁹ notes that the correlation between the test statistics is directly linked to the allocation ratio, as follows:

$$\rho_{AB} = \frac{1}{\sqrt{\left(\frac{n_Z}{n_A} + 1\right)\left(\frac{n_Z}{n_B} + 1\right)}}$$

where n_i is the sample size in group i ($i = A, B, Z$).

If the allocation ratio is 1:1:1, the correlation is 0.5. For an allocation of 2:1:1 in favour of control, the correlation is 0.333. With 1:2:2, the correlation is 0.667.

4.4 Calculating the FWER, FMER and MSFP assuming a multivariate normal distribution, incorporating correlation

The various false positive errors of potential interest in a multi-arm trial can be calculated based on the assumption of the joint distribution of the test statistics following a standardised multivariate normal distribution. The R program in Appendix 1 computes these probabilities in the case of two or three experimental therapies, allowing varying correlation, in order to calculate the error density in each of the rejection regions.

The effect of the correlation on the rejection regions in the case of two experimental treatments is illustrated in figure 3. As the correlation increases, the proportion of error in the lower left and upper right corners, indicating false positive outcomes in the same direction for both hypotheses, also increases. That is, if the shared control group performs better or worse than expected, there is a greater chance of an error in both of the hypotheses in the same direction, as expected. The effect of this correlation on the different types of errors described in Section 4.1 can easily be calculated by solving the probabilities of the outcomes falling within the relevant rejection regions.

[Insert Figure 3]

Figure 3. *Illustration of rejection regions for:*

a) two hypotheses, each with individual control data, $\rho=0$

b) two hypotheses with shared control data and 2:1:1 randomisation, $\rho=0.333$

c) two hypotheses with shared control data and 1:1:1 randomisation, $\rho=0.5$

d) two hypotheses with shared control data and 1:2:2 randomisation, $\rho=0.667$

4.5 FWER, FMER and MSFP comparison for multi-arm trials with a shared control group compared to independent trials

The R program in Appendix 1a can be used to calculate the FWER, FMER and MSFP rates for different levels of correlation based on the allocation ratio in three-arm trials with two experimental arms and a shared control. The code has also been extended to calculate the probabilities for four-arm trials with three experimental arms and a shared control, provided in Appendix 1b. These probabilities are shown in table 1.

	Independent case (Separate trials)	Dependent case 2:1:1(:1) (2 to control)	Dependent case 1:1:1(:1)	Dependent case 1:2:2(:2) (1 to control)
Correlation (ρ)	0	0.333	0.5	0.667
Reject H_0 for each individual hypothesis (A, B or C)	0.050	0.050	0.050	0.050
Three-arm trial (hypotheses A and B)				
FWER: Reject at least one H_0, A or B	0.0975	0.0946	0.0908	0.0849

FMER: Reject both H_0's A and B (in any direction)	0.0025	0.0054	0.0093	0.0151
MSFP: Reject both H_0's in favour of treatments A and B	0.00063	0.00267	0.00462	0.00753
Four-arm trial (hypotheses A, B and C)				
FWER: Reject at least one H_0, A, B or C	0.1426	0.1348	0.1254	0.1124
FMER2: Reject at least two H_0's (in any direction)	0.0072	0.0141	0.0214	0.0301
FMER3: Reject all three H_0's A, B and C (in any direction)	0.0001	0.0011	0.0032	0.0076
MSFP2: Reject at least two H_0's in favour of A, B or C	0.0018	0.0069	0.0107	0.0150
MSFP3: Reject all three H_0's in favour of A, B and C	0.00002	0.00056	0.00160	0.00378

Table 1. FWER, FMER and MSFP comparisons for three and four arm trials with a shared control group and varying allocation ratios, compared to independent 1:1 randomised trials ($\alpha=0.05$ for each hypothesis)

FWER:

The FWER is lower in all cases with shared control data than the equivalent error when assessing two independent trials. That is, the correlation between the test statistics

reduces the overall probability of a type-I error occurring across either of the hypotheses over the case where there is no shared control data, as also shown by Proschan et al.¹⁵.

FMER:

In a multi-arm trial with two hypotheses, the chance of multiple errors has increased from 0.25% for independent trials to 0.93% in the case with even allocation, an increase of 3.7 times. The message stays the same as the number of hypotheses increases; in the case with three hypotheses and even allocation, the chance of any two errors is now over 2%, which is not trivial. Similar increases are found with unequal allocation ratios and the trend across the resultant correlations from these changing allocation ratios can be clearly seen.

The increase in the FMER is due to the increased chance of an error occurring within the correlated comparisons in the same direction. This is caused by a chance deviation in the outcome for the control sample from the outcome for the true population. The probability of multiple type-I errors in opposite directions has decreased, as expected from figure 3, but to a lesser extent than the increase in the chance of errors in the same direction. Recall that the total error (FWER + FMER) is fixed, thus the increased FMER explains the reduction in the FWER.

MSFP:

With two hypotheses, the MSFP rate has increased from 0.06% in independent trials to 0.46% in the multi-arm case with even allocation, an increase of 7.7 times. With three hypotheses, the chance of any two superior false positive outcomes has increased by nearly 6 times to over 1%, and the chance of three MSFPs is substantially greater than in the independent case, although the probability is very small at 0.16%. Again, similar patterns and trends are seen for other allocation ratios. This is intuitively obvious since a chance ‘bad’ outcome in the control sample compared to the true population would increase the chances of false positives in both hypotheses, but the magnitude of this effect is now apparent, and is not trivial.

5. An investigation of the effect of multiplicity adjustment methods

Many multiplicity adjustment procedures have been devised to strongly control the FWER for a number of tests within a family of hypotheses. The adjustment methods considered here are: Bonferroni²⁰, a simple, conservative and popular adjustment method; Holm²¹ and Hochberg²², closed testing methods based on a hierarchical strategy of testing the outcomes ordered by significance; Dunnett’s t ¹⁹, a parametric

method that adjusts the Bonferroni boundaries to control the probability of observing a significant result under H_0 at 0.05; and Dunnett and Tamhane²³, an adjusted Hochberg step-up multiple test procedure in which the rejection levels are adjusted to account for the correlation so that the final FWER is 0.05.

Table 2 shows the effects of applying these adjustment methods on the various error rates, using the example of a three arm trial with 1:1:1 allocation ratio in which the two experimental arms are compared to a shared control group.

	Independent case	Dependent case, 1:1:1 allocation					
		Un-adjusted	Bonferro ni	Holm	Hochberg	Dunnett's t	Adjusted Hochberg
Reject H_0 for individual comparison (A or B)	0.0500	0.0500	0.0250	0.0271	0.0286	0.0271	0.0296
FWER: Reject at least one H_0, A or B	0.0975	0.0908	0.0465	0.0465	0.0480	0.0502	0.0500
FMER: Reject both H_0's A and B (in any direction)	0.0025	0.0093	0.0035	0.0077	0.0093	0.0039	0.0093
MSFP: Reject both H_0's in favour of treatments A and B	0.00063	0.00462	0.00176	0.00385	0.00462	0.00197	0.00462

Table 2. *FWER, FMER and MSFP comparisons for three arm trials with two hypotheses ($\alpha=0.05$ for each), a shared control group and even allocation ratio, after applying various multiple testing adjustments*

FWER:

All adjustment methods control the FWER at 0.05 or less, as expected. In all cases, the chance of rejecting the null hypothesis for each individual comparison has taken a penalty compared to running independent trials. The Dunnett's t and Adjusted Hochberg methods account for the effect of the correlation due to the shared control data on the FWER, in order to make less conservative adjustments than the other methods.

FMER:

Although adjustment methods control the probability of falsely rejecting at least one hypothesis, no method fully controls the chance of multiple errors occurring within the same set of hypotheses to be what it would have been if the hypotheses had been assessed in independent trials.

With Bonferroni and Dunnett's t , the probabilities of multiple errors are reduced towards those in independent trials, but the Holm, Hochberg and Adjusted Hochberg methods based on the closed testing principle offer very little or no protection of the FWER over no adjustment. The first step of a step-up procedure is to accept all hypotheses if the least significant is <0.05 , so it can easily be seen why this is the case. Intuitively, if the critical level is set lower, as with the Bonferroni method, fewer null hypotheses will be rejected in all comparisons, and therefore the chance of both being rejected will also decrease. However, since a 'bad' outcome in the shared control sample would still affect both comparisons by increasing the chances of a false positive error, it makes sense that a higher proportion of paired hypotheses are both rejected when they share a common control.

MSFP:

Since the adjustment methods do not control the FWER, they also do not offer full protection against the chance of MSFP outcomes. After applying the Bonferroni and Dunnett's t corrections, the chance of two superior false positive errors is still inflated by approximately 3 times over that with independent trials, and following the Hochberg adjustments this rises to over 7 times.

The above results highlight that multiple testing adjustment methods only control the probability of the overall FWER to that for a single hypothesis. They do not offer control over the chance of multiple false positive errors, which is the probability that is increased over that had the hypotheses been assessed in independent trials.

6. Controlling the probability of multiple superior false positive outcomes

We have shown that in the case where two superior hypotheses may both be used to jointly inform a claim of effectiveness, the overall chance of both having a false positive outcome in favour of the experimental treatments (MSFP) is inflated in a multi-arm trial over that chance occurring in independent trials. In addition, applying multiple testing correction methods do not reduce the chance of MSFP outcomes to the same level as in two independent trials.

The FDA guidance on ‘Providing Clinical Evidence of Effectiveness for Human Drug and Biological Products’²⁴ suggests that it is feasible for multiple hypotheses from within a single study to be accepted as evidence of effectiveness if the trial is designed appropriately. In this case, it would be important that the probability of multiple conclusions of superiority (MSFP) is not inflated over that for independent studies. The example of the AMAGINE-1 trial described in Section 2.2 assesses two doses of an experimental treatment against placebo. If these doses were investigated in independent

trials, both trial outcomes may be used to inform a claim of effectiveness, but the penalty for assessing these within a multi-arm trial in terms of inflation of the MSFP rate has not been investigated or quantified. This is a similar issue to that discussed by Shun et al²⁵, in which they require the overall ‘positive rejection region’ in a large pivotal trial to be controlled to the same level as in two smaller pivotal trials.

6.1 Critical values to control the MSFP rate in the case of three arms

In two independent trials, the chance of two superior false positive outcomes is 0.000625 (Section 4.1). Since the joint distribution can be described using a bivariate normal (Section 4.4), this can be used to obtain the exact critical value that returns a probability of 0.000625. This principle is similar to the work of Follmann et al., which relies on the multivariate normal assumption of the test statistics to estimate critical values that strongly protect the type-I error rate in the case of multi-armed trials with interim looks¹⁸. The R code to calculate the critical value is provided in Appendix 2.

In the 1:1:1 case, the critical value required to protect the MSFP rate at 0.000625 is 0.0118. In the 2:1:1 case it is 0.0195, and in the 1:2:2 case it is 0.0069, as shown in Table 3. That is, if two hypotheses are assessed in a multi-arm trial with a shared control group, and are to be used to jointly inform a claim of effectiveness; in order to

control the probability of two superior false positive outcomes to the level in independent trials, the p-values for both hypotheses are required to be less than these adjusted critical values. It can be seen that with this level of control, the FWER is reduced to much lower than 5%.

	Independent case No adjustment	Dependent case No adjustment	Dependent case 2:1:1 $\alpha = 0.0195$	Dependent case 1:1:1 $\alpha = 0.0118$	Dependent case 1:2:2 $\alpha = 0.0069$
Reject H_0 for individual comparison (A or B)	0.050	0.050	0.0195	0.0118	0.0069
FWER: Reject at least one H_0, A or B	0.0975	0.0908	0.0377	0.0224	0.0125
MSFP: Reject both H_0's in favour of treatments A and B	0.000625	0.00462	0.000623	0.000624	0.000628

Table 3. Adjusted critical values to control the chance of a MSFP error in a three-arm trial to that for two independent 1:1 randomised trials

6.2 The effect of MSFP control on the power and sample size

If a trial is designed to allow two superior outcomes to be used as evidence to inform a single claim of effectiveness, the power is required to be maintained for each hypothesis

as it would for independent trials, requiring an increased sample size. As an example, take a confirmatory trial with a survival primary endpoint and analysis based on the log-rank test for equality based on an exponential survival distribution. Assume that the estimated median survival in the control group is 36 months, and that a clinically relevant difference would be an improvement to 48 months ($HR=0.75$). In a two-arm trial, with a recruitment period of 48 months and an additional 36 month follow-up period, 408 patients are required per arm (1:1) to achieve 508 events for 90% power with a type-I error rate of 5%.

If there are two experimental arms of interest in the population, a three-arm trial may be considered rather than two independent trials. In the scenario of running independent trials, the total sample size for the two trials assuming 1:1 allocations would be 1632, and in the multi-arm trial this is reduced to 1224 with no adjustment. Adjusting to control the chance of MSFP outcomes reduces the power from 90% to 77%, and to account for this loss in power the sample size would need to be increased by 37% to 1680, which makes the multi-arm trial slightly larger than running two independent trials. There may still be benefits of running a multi-arm trial, however, in terms of reducing the total number of patients receiving the control therapy as well as the time and cost of only needing to set-up and run a single trial.

7. Discussion

Multi-arm trials can be efficient and therefore advantageous over running independent trials. However, there are conflicting views in the literature on how to appropriately control the probability of a false positive error. A lack of proper control of the FWER could lead to an unacceptable chance of an ineffective treatment being recommended to be taken forward into practice; but unnecessary control of the FWER could affect the efficiency of a trial, requiring increased patient numbers and resources. FWER adjustment without increasing the sample size to maintain power could lead to a superior treatment being denied. Each of these scenarios raises ethical concerns.

False positive error rates may be affected in multi-arm trials compared to independent trials due to either: correlation between the comparisons caused by the shared use of the control data; or an increased chance of making a claim of effectiveness because of an increased ability to test a family of hypotheses. It is a common misconception that FWER adjustment is necessary due to sharing control data. When considering the designs illustrated in Figures 1a and 1b, some might assume that the overall FWER for the family of hypotheses, H_{0A} and H_{0B} , would be larger in Figure 1b where there is a common control group. However, we have confirmed that the FWER is in fact smaller in Figure 1b than in Figure 1a. The common control group instead has the effect of

increasing the chances of more than one false positive outcome within the family of hypotheses; although FWER adjustment methods do not control for this. The necessity for a FWER adjustment is therefore only dependent on whether assessing multiple hypotheses within a multi-arm trial has increased the chance of making a single claim of effectiveness.

We have formalised the implications of running a multi-arm trial with shared control data on the probabilities of various types of false positive errors, considering the effects of multiple testing adjustment methods on these probabilities, in order to make informed recommendations on the requirement for adjustment. The findings are summarised below. In addition, a flow diagram to aid the determination of the requirement for a multiple testing adjustment in a multi-arm trial is provided in Figure 4. Note that the decisions on the need for error control with respect to the interpretation of the trial results should be agreed and documented in advance in the protocol and statistical analysis plan.

[Insert Figure 4]

Figure 4. *Decision diagram to determine the requirement for a multiple testing adjustment in multi-arm trials*

7.1 Familywise Error Rate (FWER) in the case where each hypothesis informs a different claim of effectiveness

Consider the case in which multiple hypotheses are being assessed in the same trial with a shared control group, but each hypothesis informs a different claim of effectiveness. An obvious example is where the experimental arms assess entirely different therapies. A further example is the MRC COIN trial¹² introduced in Section 2.1. OxFP is present in all treatment arms, however since one primary hypothesis addresses the addition of an experimental therapy to OxFP and the other addresses a reduction in duration of OxFP therapy, these do not contribute towards the same claim of effectiveness. In this case, the chance of a false positive outcome for either claim of effectiveness is not increased by the presence of the other hypothesis.

Since we have confirmed that the correlation between comparisons due to the shared control data has the effect of reducing the overall chance of at least one false positive finding (FWER) over that chance had the same questions been asked in independent hypothesis tests, FWER adjustment is not necessary due to the shared control sample. These findings remain valid in the case of imbalanced randomisations, and also where there are more than two experimental therapies.

Therefore, adjusting to control the FWER in this case is an unnecessary penalty. It does not make sense here to consider the type-I error rate for the trial as a whole since any claim of effectiveness is only informed by a single hypothesis test. Note that if the experimental therapies are competing against each other for approval in the trial population, the correlation due to the shared control group is an advantage. If in two independent trials one of the control samples performs worse than the true population, the associated experimental group has an increased chance of being significant and taken forward. However, in the equivalent multi-arm case, bias due to comparisons to different control samples is removed. It is more likely that efficacious experimental therapies would be considered against each other directly without the influence of variations in the control samples.

7.2 Familywise Error Rate (FWER) in the case where the hypotheses inform a single claim of effectiveness

We have shown that the FWER is not inflated due to running a multi-arm trial with a shared control group. However, it should be noted that FWER adjustment may be required if the efficiency of running a multi-arm trial leads to more hypotheses being included than would have otherwise been assessed in independent trials. If these hypotheses inform the same claim of effectiveness, for example if the experimental

arms assess different doses or combinations of the same experimental therapy, the overall chance of a false positive result anywhere within the family of hypotheses is clearly increased over that for a single hypothesis. In this case, there is general agreement in the literature that FWER control is recommended⁷ since the type-I error rate can be considered for the claim of effectiveness as a whole, rather than for each individual hypothesis. Note that it also follows that the power in this case can be considered to be the overall chance of observing at least one true positive outcome, and this will also be increased by testing multiple hypotheses. Therefore, the penalty caused by applying the FWER adjustment may be compensated to some extent by the gain in overall power.

An example here is the AMAGINE-1 trial (Section 2.2) assessing two doses of brodalumab compared to placebo. Since a rejection of the primary hypotheses for each comparison could lead to a claim of effectiveness for brodalumab, there are two chances for a false positive result with respect to that claim, and therefore FWER adjustment is recommended.

7.3 Familywise Error Rate (FWER) in the case where all hypotheses are required to have superior outcomes in order to make a claim of effectiveness

If all trial hypotheses are required to be positive in order to make a claim of effectiveness, there is essentially only one overarching hypothesis being tested. In this case, there is only one chance for an overall false positive outcome for the trial, so the chance of ‘at least one’ error cannot be inflated, and therefore no FWER adjustment is necessary.

For example, the Myeloma XI+ Intensive trial (Section 2.3) compared the four-drug regime CCRD against the current standard control CTD, as well as the previously assessed three-drug regime CRD (which is CCRD excluding carfilzomib). Since CCRD will only be recommended for approval if it is better than both CTD and CRD, both hypotheses are required to be significant in order to recommend CCRD for use in practice. In this case, no adjustment is required.

7.4 Multiple Superior False Positives (MSFP) in the case where multiple superior outcomes could be used as separate pieces of evidence towards a single claim of effectiveness

In Section 7.2 we discuss the case where multiple hypotheses are being assessed in the same trial with a shared control group, with each hypothesis being tested and reported individually. If superiority in more than one of these hypotheses could contribute as

separate pieces of evidence towards a claim of effectiveness for a therapy, a more stringent adjustment is required for the evidence to be equivalent to that obtained from two independent trials, as discussed in Section 6.

We have shown that the correlation due to the shared control group increases the chance of falsely declaring more than one experimental treatment group to be superior to the shared control group (MSFP) in a multi-arm trial over that chance had the hypotheses been assessed in independent trials. Standard multiple testing adjustment methods do not adequately control for this. We have proposed a critical value adjustment to control the chance of MSFP outcomes in a three-arm trial in order for the evidence to be equivalent to that obtained from two independent trials.

Care should always be taken in reporting and interpretation if more than one hypothesis within a multi-arm trial with shared control group is positive.

Funding

DRH is funded by a Doctoral Research Fellowship from the National Institute for Health Research (NIHR) [reference number DRF-2012-05-364]. This paper presents independent research funded by the NIHR. The views expressed in this publication are

those of the authors and not necessarily those of the NHS, the NIHR or the Department of Health.

References

1. Parmar MKB, Carpenter J and Sydes MR. More multiarm randomised trials of superiority are needed. *The Lancet*. 2014; 384: 283-4.
2. CPMP (Committee for Proprietary Medicinal Products). Point to Consider on multiplicity issues in clinical trials. EMEA (European Medicines Agency), 2002.
3. Wason J, Magirr D, Law M and Jaki T. Some recommendations for multi-arm multi-stage trials. *Statistical Methods in Medical Research*. 2012.
4. Bender R and Lange S. Adjusting for multiple testing—when and how? *Journal of Clinical Epidemiology*. 2001; 54: 343-9.
5. Freidlin B, Korn EL, Gray R and Martin A. Multi-Arm Clinical Trials of New Agents: Some Design Considerations. *Clinical Cancer Research*. 2008; 14: 4368-71.
6. Proschan MA and Waclawiw MA. Practical Guidelines for Multiplicity Adjustment in Clinical Trials. *Controlled Clinical Trials*. 2000; 21: 527-39.
7. Wason JM, Stecher L and Mander A. Correcting for multiple-testing in multi-arm trials: is it necessary and is it done? *Trials*. 2014; 15: 364.
8. Cook RJ and Farewell VT. Multiplicity Considerations in the Design and Analysis of Clinical Trials. *Journal of the Royal Statistical Society Series A (Statistics in Society)*. 1996; 159: 93-110.

9. Rothman KJ. No Adjustments Are Needed for Multiple Comparisons. *Epidemiology*. 1990; 1: 43-6.
10. ICH (International Conference on Harmonisation). Statistical Principles for Clinical Trials E9. 1998.
11. Phillips A, Fletcher C, Atkinson G, et al. Multiplicity: discussion points from the Statisticians in the Pharmaceutical Industry multiplicity expert group. *Pharmaceutical Statistics*. 2013; 12: 255-9.
12. Maughan TS, Adams RA, Smith CG, et al. Addition of cetuximab to oxaliplatin-based first-line combination chemotherapy for treatment of advanced colorectal cancer: results of the randomised phase 3 MRC COIN trial. *The Lancet*. 377: 2103-14.
13. Westfall P and Bretz F. Multiplicity in Clinical Trials. *Encyclopedia of Biopharmaceutical Statistics*, Third Edition. Taylor & Francis, 2014, p. 889-96.
14. Fernandes N and Stone A. Multiplicity adjustments in trials with two correlated comparisons of interest. *Statistical Methods in Medical Research*. 2011; 20: 579-94.
15. Proschan M and Follman D. Multiple comparisons with control in a single experiment ver. *The American Statistician*. 1995; 49: 144.
16. Senn SS. *Statistical issues in drug development*. John Wiley & Sons, 1997.
17. Hung HMJ and Wang S-J. Challenges to multiple testing in clinical trials. *Biometrical Journal*. 2010; 52: 747-56.

18. Follmann DA, Proschan MA and Geller NL. Monitoring Pairwise Comparisons in Multi-Armed Clinical Trials. *Biometrics*. 1994; 50: 325-36.
19. Dunnett CW. A Multiple Comparison Procedure for Comparing Several Treatments with a Control. *Journal of the American Statistical Association*. 1955; 50: 1096-121.
20. Abdi H. Bonferroni and Sidak corrections for multiple comparisons. In: Salkind NJ, (ed.). *Encyclopedia of Measurement and Statistics*. Thousand Oaks, CA: Sage, 2007, p. 103-7.
21. Holm S. A simple sequentially rejective multiple test procedure. *Scandinavian journal of statistics*. 1979: 65-70.
22. Hochberg Y. A sharper Bonferroni procedure for multiple tests of significance. *Biometrika*. 1988; 75: 800-2.
23. Dunnett CW and Tamhane AC. A Step-Up Multiple Test Procedure. *Journal of the American Statistical Association*. 1992; 87: 162-70.
24. FDA. *Providing Clinical Evidence of Effectiveness for Human Drug and Biological Products*. Rockville, MD1998.
25. Shun Z, Chi E, Durrleman S and Fisher L. Statistical consideration of the strategy for demonstrating clinical evidence of effectiveness—one larger vs two smaller pivotal studies *Statistics in Medicine* 2005; 24:1619–1637. *Statistics in Medicine*. 2005; 24: 1652-6.

Appendix 1a: R code to calculate the probabilities for the rejection regions based on two correlated comparisons, assuming a bivariate normal distribution.

```
#Install library first use
setInternet2(TRUE)
install.packages("mvtnorm")
library(mvtnorm)

#Bivariate normal case (2 experimental arms)

#set correlation
corr <- 0

#correlation matrix
corrmatrix <- matrix(c(1,corr,corr,1),ncol=2,byrow=TRUE)

#Exactly 1 error (calculate probabilities for the edges
#excluding the corners of the square)

# Left hand side
leftside <- pmvnorm(lower=c(-Inf,-1.96), upper=c(-1.96,1.96), corr =
corrmatrix )

# Right hand side
rightside <- pmvnorm(lower=c(1.96,-1.96), upper=c(Inf,1.96), corr =
corrmatrix )

# Top edge
topside <- pmvnorm(lower=c(-1.96,1.96), upper=c(1.96,Inf), corr =
corrmatrix )

# Bottom edge
bottomside <- pmvnorm(lower=c(-1.96,-Inf), upper=c(1.96,-1.96), corr =
corrmatrix )

# Total chance of exactly 1 error
oneonly=leftside+rightside+topside+bottomside

#Exactly 2 errors (calculate probabilities in each of the 4 corners of
the square)

# Lower left corner
lowleft <- pmvnorm(lower=c(-Inf,-Inf), upper=c(-1.96,-1.96), corr =
corrmatrix )
```

```

# Lower right corner
lowright <- pmvnorm(lower=c(1.96,-Inf), upper=c(Inf,-1.96), corr =
corrmat )

# Upper Left corner
upleft <- pmvnorm(lower=c(-Inf,1.96), upper=c(-1.96,Inf), corr =
corrmat )

# Upper Right corner
upright <- pmvnorm(lower=c(1.96,1.96), upper=c(Inf,Inf), corr =
corrmat )

# Total chance of exactly 2 errors
twoonly <- lowleft+lowright+upleft+upright

# FWER
FWER <- oneonly+twoonly

# Probability of any two errors (FMER)
FMER <- twoonly

# MSFP probability of two superior false positives
MSFP <- upright

#Output results
corrmat
FWER
FMER
MSFP

```

Appendix 1b: R code to calculate the probabilities for the rejection regions based on three correlated comparisons, assuming a trivariate normal distribution.

```
# Install library first use
setInternet2(TRUE)
install.packages("mvtnorm")
library(mvtnorm)

# Trivariate normal case (3 experimental arms)

#set correlation
corr <- 0.5

#correlation matrix
corrmat <-
matrix(c(1,corr,corr,corr,1,corr,corr,corr,1),ncol=3,byrow=TRUE)

#Exactly 1 error (illustrated by the 6 side face of a cube minus the
upper and lower 5% around the edges)

Oneonly1 <- pmvnorm(lower=c(-1.96,-1.96,-Inf), upper=c(1.96,1.96,-
1.96), corr = corrmat )
Oneonly2 <- pmvnorm(lower=c(-1.96,-1.96,1.96), upper=c(1.96,1.96,Inf),
corr = corrmat )
Oneonly3 <- pmvnorm(lower=c(-Inf,-1.96,-1.96), upper=c(-
1.96,1.96,1.96), corr = corrmat )
Oneonly4 <- pmvnorm(lower=c(1.96,-1.96,-1.96), upper=c(Inf,1.96,1.96),
corr = corrmat )
Oneonly5 <- pmvnorm(lower=c(-1.96,-Inf,-1.96), upper=c(1.96,-
1.96,1.96), corr = corrmat )
Oneonly6 <- pmvnorm(lower=c(-1.96,1.96,-1.96), upper=c(1.96,Inf,1.96),
corr = corrmat )

# Total chance of exactly 1 error
Oneonly=Oneonly1+Oneonly2+Oneonly3+Oneonly4+Oneonly5+Oneonly6

#Exactly 2 errors (illustrated by the 12 edges of a cube minus the
upper and lower 5% in the corners)

# the 3 edges that corner the triple rejection in favour of control
(lower left front)
onlyllx <- pmvnorm(lower=c(-1.96,-Inf,-Inf), upper=c(1.96,-1.96,-
1.96), corr = corrmat )
onlylly <- pmvnorm(lower=c(-Inf,-1.96,-Inf), upper=c(-1.96,1.96,-
1.96), corr = corrmat )
```



```

onlyllz <- pmvnorm(lower=c(-Inf,-Inf,-1.96), upper=c(-1.96,-
1.96,1.96), corr = corrmat )

# the 3 edges that corner the triple rejection in favour of the
experimental arms (upper right back)
onlyurx <- pmvnorm(lower=c(-1.96,1.96,1.96), upper=c(1.96,Inf,Inf),
corr = corrmat )
onlyury <- pmvnorm(lower=c(1.96,-1.96,1.96), upper=c(Inf,1.96,Inf),
corr = corrmat )
onlyurz <- pmvnorm(lower=c(1.96,1.96,-1.96), upper=c(Inf,Inf,1.96),
corr = corrmat )

#Off edges (l=lower u=upper f=front b=back l=left r=right):
onlylrz <- pmvnorm(lower=c(1.96,-Inf,-1.96), upper=c(Inf,-1.96,1.96),
corr = corrmat )
onlyfry <- pmvnorm(lower=c(1.96,-1.96,-Inf), upper=c(Inf,1.96,-1.96),
corr = corrmat )
onlylbx <- pmvnorm(lower=c(-1.96,-Inf,1.96), upper=c(1.96,-1.96,Inf),
corr = corrmat )
onlyufx <- pmvnorm(lower=c(-1.96,1.96,-Inf), upper=c(1.96,Inf,-1.96),
corr = corrmat )
onlyulz <- pmvnorm(lower=c(-Inf,1.96,-1.96), upper=c(-1.96,Inf,1.96),
corr = corrmat )
onlybly <- pmvnorm(lower=c(-Inf,-1.96,1.96), upper=c(-1.96,1.96,Inf),
corr = corrmat )

# Total chance of exactly 2 errors
Twoonly <-
onlyllx+onlylly+onlyllz+onlyurx+onlyury+onlyurz+onlylrz+onlyfry+onlylb
x+onlyufx+onlyulz+onlybly

# Exactly 3 errors (Calculate probabilities in each of the 8 corners
of a cube)
xlylzl <- pmvnorm(lower=c(-Inf,-Inf,-Inf), upper=c(-1.96,-1.96,-1.96),
corr = corrmat )

x2ylzl <- pmvnorm(lower=c(1.96,-Inf,-Inf), upper=c(Inf,-1.96,-1.96),
corr = corrmat )
xly2zl <- pmvnorm(lower=c(-Inf,1.96,-Inf), upper=c(-1.96,Inf,-1.96),
corr = corrmat )
xlylzl <- pmvnorm(lower=c(-Inf,-Inf,1.96), upper=c(-1.96,-1.96,Inf),
corr = corrmat )

x2y2zl <- pmvnorm(lower=c(1.96,1.96,-Inf), upper=c(Inf,Inf,-1.96),
corr = corrmat )
x2ylzl <- pmvnorm(lower=c(1.96,-Inf,1.96), upper=c(Inf,-1.96,Inf),
corr = corrmat )
xly2zl <- pmvnorm(lower=c(-Inf,1.96,1.96), upper=c(-1.96,Inf,Inf),
corr = corrmat )

```

```

x2y2z2 <- pmvnorm(lower=c(1.96,1.96,1.96), upper=c(Inf,Inf,Inf), corr
= corrmat )

# Total chance of exactly 3 errors
Threeonly <- x1y1z1+x2y1z1+x1y2z1+x1y1z2+x2y2z1+x2y1z2+x1y2z2+x2y2z2

#FWER - the overall error region of the sides, edges and corners
FWER=Oneonly+Twoonly+Threeonly

# Probability of at least any two errors
twoerr <- Twoonly+Threeonly

# Probability of three errors - sum of the corner regions
threeerr <- Threeonly

# Two MSFP - probability of at least two superior false positives
# Sum of the 3 edges meeting the upper right back
#(i.e. two false positives along the plane of the third distribution)
# and the upper right corner

TwoMSFP <- onlyyurx+onlyyury+onlyyurz+x2y2z2

# Three MSFP - probability of three positive false positives
ThreeMSFP <- x2y2z2

#Output results
corrmat
FWER
twoerr
threeerr
TwoMSFP
ThreeMSFP

```

Appendix 2: R code to calculate the rejection region required to control the probability of two MSFP errors based on correlated comparisons to that for independent trials, assuming a multivariate normal distribution.

```
# Install library first use
setInternet2(TRUE)
install.packages("mvtnorm")
library(mvtnorm)

#The MSFP is the upper right corner of the rejection regions in
#figure 2 (based on the standardised bivariate normal).
#The MSFP needs to be controlled at 0.000625 (0.025**2)

#set correlation
corr <- 0.5

#correlation matrix
corrmat <- matrix(c(1,corr,corr,1),ncol=2,byrow=TRUE)

#Solve the critical value for the upper right corner
#equalling 0.000625
upperx <- qmvnorm(p=0.000625,tail=c("upper.tail"),corr=corrmat)
uppertail <- upperx$quantile
adjcval <- 2*(1-pnorm(uppertail))
adjcval
```