

Comparable reductions in hyperphoeainduced bronchoconstriction and markers of airway inflammation after supplementation with 6.2 and 3.1 g/d of long chain omega-3 polyunsaturated fatty acids in adults with asthma

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## Comparable reductions in hyperphoea-induced bronchoconstriction and markers of airway inflammation after supplementation with 6.2 and 3.1 g/d of long chain omega-3 polyunsaturated fatty acids in adults with asthma

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- **6 Authors:** Neil C. Williams<sup>1</sup>, Kirsty A. Hunter<sup>1</sup>, Dominick E. Shaw<sup>2</sup>, Kim G. Jackson<sup>3</sup>, Graham R.
- 7 Sharpe<sup>1</sup> and Michael A. Johnson<sup>1</sup>
- 8 <sup>1</sup>Exercise and Health Research Group, Sport, Health and Performance Enhancement (SHAPE)
- 9 Research Centre, Department of Sport Science, Nottingham Trent University, Nottingham, NG11
- 10 8NS, UK
- <sup>2</sup>Respiratory Research Unit, University of Nottingham, Nottingham, NG5 1PB, UK
- <sup>3</sup>Hugh Sinclair Unit of Human Nutrition, Department of Food and Nutritional Sciences, University
- 13 of Reading, Reading, RG6 6AP, UK
- 14
- 15 Corresponding Author: Dr Neil C. Williams, Exercise and Health Research Group, Sport, Health
- 16 and Performance Enhancement Research Centre, Department of Sports Science, School of Science
- 17 and Technology, Nottingham Trent University, Nottingham, NG11 8NS, UK
- 18 E-mail: <u>neil.williams@ntu.ac.uk</u>
- 19
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### 25 Abstract

Although high dose omega-3 polyunsaturated fatty acid (n-3 PUFA) supplementation 26 27 reduces exercise- and hyperphoea-induce bronchoconstriction (EIB/HIB), there are concurrent 28 issues with cost, compliance, and gastrointestinal discomfort. It is thus pertinent to establish the 29 efficacy of lower *n*-3 PUFA doses. Eight male adults with asthma and HIB and 8 controls without 30 asthma were randomly supplemented with two n-3 PUFA doses (6.2 g/d (3.7g EPA and 2.5g DHA)) 31 and 3.1 g/d (1.8g EPA and 1.3g DHA)) and a placebo, each for 21 days followed by 14 days 32 washout. A eucapnic voluntary hyperphoea (EVH) challenge was performed before and after treatments. Outcome measures remained unchanged in the control group. In the HIB group, the 33 peak fall in forced expiratory volume in 1 s (FEV<sub>1</sub>) after EVH at day 0 (-1005 (SD 520) mL, -30 34 35 (SD 18) %) was unchanged after placebo. The peak fall in  $FEV_1$  was similarly reduced from day 0 to day 21 of 6.2 g/d n-3 PUFA (-1000 (SD 460) mL, -29 (SD 17) % vs. -690 (SD 460) mL, -20 (SD 36 37 15) %) and 3.1 g/d n-3 PUFA (-970 (SD 480) mL, -28 (SD 18) % vs. -700 (SD 420) mL, -21 (SD 15) %) (P<0.001). Baseline fraction of exhaled nitric oxide was reduced by 24% (P=0.020) and 38 31% (P = 0.018) after 6.2 and 3.1 g/d n-3 PUFA, respectively. Peak increases in 9 $\alpha$ , 11B PGF<sub>2</sub> after 39 40 EVH were reduced by 65% (P=0.009) and 56% (P=0.041) after 6.2 and 3.1 g/d n-3 PUFA, 41 respectively. In conclusion, 3.1 g/d n-3 PUFA supplementation attenuated HIB and markers of 42 airway inflammation to a similar extent as a higher dose. Lower doses of n-3 PUFA thus represent a 43 potentially beneficial adjunct treatment for adults with asthma and EIB.

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#### 54 Introduction

Exercise-induced bronchoconstriction (EIB) is a prominent asthma phenotype affecting an 55 estimated 90% of asthma patients and up to 50% of elite athlete populations<sup>(1)</sup>. EIB is characterised 56 by transient airway narrowing during and/or after exercise<sup>(2)</sup> and is ascribed to airway drying 57 58 leading to degranulation of inflammatory cells and release of inflammatory mediators<sup>(3, 4)</sup>. Inhaled 59 corticosteroids and short- and long-acting  $\beta_2$ -agonists are effective therapies, but they are not curative and do not modify disease progression<sup>(5)</sup>. Furthermore, inhaled corticosteroids adherence is 60 notoriously poor and may have undesirable side effects, while chronic  $\beta_2$ -agonist use results in 61 tolerance<sup>(5, 6)</sup>. Development of therapies that modulate asthma immunopathology without adverse 62 side effects is therefore desirable. 63

64 One potential candidate therapy involves the omega-3 long chain polyunsaturated fatty acids (*n*-3 PUFA) eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA)<sup>(7)</sup>. Dietary 65 66 supplementation of n-3 PUFA increases cell membrane EPA and DHA content and reduces n-6arachidonic acid content<sup>(8)</sup>. This subsequently reduces the synthesis of the pro-inflammatory 67 arachidonic acid-derived eicosanoids cysteinyl leukotrienes and prostaglandins. EPA and DHA may 68 also increase synthesis of resolvin and protectin compounds through the cyclooxygenase and 69 70 lipoxygenase pathways which are involved in the resolution of inflammation. These mechanisms are thought to be central to the well-established anti-inflammatory effects of n-3 PUFA<sup>(8)</sup>, which 71 provide a rationale for the use of n-3 PUFA in asthma<sup>(7)</sup>. To date, however, the role of EPA and 72 DHA in the management of asthma and EIB remains uncertain<sup>(9-14)</sup>. Early research showed that 73 supplementation of 5.4 g/d n-3 PUFA (3.2 g/d EPA and 2.2 g/d DHA) for 10 weeks in adults with 74 75 asthma reduced leukotriene generation and neutrophil chemotactic responsiveness, but did not reduce bronchoconstriction after cycling exercise and a histamine challenge<sup>(9)</sup>. Similarly, 76 supplementation of 6.0 g/d n-3 PUFA (4.0 g/d EPA and 2.0 g/d DHA) for 3 weeks in adults with 77 asthma did not attenuate bronchoconstriction or markers of airway inflammation in response to a 78 mannitol challenge test<sup>(12)</sup>. Conversely, supplementation of 5.4 g/d *n*-3 PUFA (3.2 g/d EPA and 2.2 79 g/d DHA) for 3 weeks in elite athletes<sup>(10)</sup> and physically active asthmatic males<sup>(11, 15)</sup> abolished EIB. 80 Subsequent studies showed that supplementation of n-3 PUFA (3.2 g/d EPA and 2.0 g/d DHA) 81 reduced bronchoconstriction and markers of airway inflammation after eucapnic voluntary 82 hyperphoea (EVH)<sup>(16)</sup> and that *n*-3 PUFA (~3.2 g/d EPA and ~2.0 g/d DHA) was as effective as a 83 leukotriene modifier<sup>(17)</sup>. 84

85 Given the lack of reported risk in taking n-3 PUFA, the American Thoracic Society/European Respiratory Society (ATS/ERS) guidelines suggest that the latter studies offer 86 some support for the consumption of n-3 PUFA by interested individuals with EIB<sup>(10, 11, 17)</sup>. In 87 contrast however, recent practice parameter  $^{(14)}$  cautions the recommendation for *n-3* PUFA use in 88 EIB based on more recent findings<sup>(12, 18)</sup>. When considering the practicalities associated with n-389 PUFA supplementation for asthma management, previous studies have used doses of 5.2-6.0 g/d. 90 Such doses require individuals to ingest between 8-20 capsules of commercial fish oil daily, which 91 has implications for cost, compliance, and gastrointestinal discomfort<sup>(19, 20)</sup>. In a single-blind study 92 an alternative flavoured beverage delivery was investigated (3.0 g/d EPA, 3.0 g/d DHA and 30 ug 93 of vitamin D3) but failed to show any attenuation in HIB or markers of airway inflammation 94 ( $F_ENO$ , urinary 9 $\alpha$ , 11 $\beta$  PGF<sub>2</sub> and cysteinyl leukotriene E4)<sup>(18)</sup>. However, no measure of compliance 95 to the treatment was made and participants included had mild asthma or did not have a physician 96 97 diagnosis of asthma. It is therefore pertinent to establish if lower doses of n-3 PUFA are effective in 98 reducing EIB. With respect to the simulation of EIB under laboratory conditions, an EVH challenge causes a highly reproducible hyperphoea-induced bronchoconstriction (HIB) (a surrogate for EIB) 99 in adults with asthma<sup>(21)</sup> which makes this an attractive challenge test to evaluate the effects of n-3100 PUFA treatments on airway hyper responsiveness. Thus, the aim of the current study was to 101 102 compare the effects of a 6.2 g/d n-3 PUFA dose with a half dose of 3.1 g/d n-3 PUFA on HIB and 103 markers of airway inflammation in adults with asthma. Provisional data from the study was previously part published in abstract form<sup>(22)</sup>. 104

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## 106 Methods

## 107 *Participants*

108 Sixteen non-smoking, recreationally active men (completing  $\geq$ 6-h of endurance exercise per 109 week) provided written, informed consent to participate in the study (Table 1). Eight participants 110 formed a HIB group and 8 formed a control group. Inclusion criteria for the HIB group were: physician diagnosis of asthma, a baseline FEV<sub>1</sub> >65% of predicted<sup>(23)</sup>, and a  $\geq$ 10% fall in FEV<sub>1</sub> 111 following initial EVH screening<sup>(24, 25)</sup>. The HIB group were on step 1-3 of the stepwise approach to 112 asthma control, indicating well-controlled asthma using either reliever medication (short acting  $\beta$ 2-113 114 agonsit) alone or in combination with controller medication (low dose inhaled corticosteroid and/or long-acting  $\beta$ 2-agonist<sup>(26)</sup> (Table 1). Inclusion criteria for the control group were: a baseline FEV<sub>1</sub> 115

116 >65% of predicted and a <10% fall in FEV<sub>1</sub> following initial EVH screening. Participants avoided 117 exercise for 24-h before an EVH test, and the HIB group ceased their medication as previously 118 descried<sup>(21, 24)</sup>. On EVH test days, participants abstained from caffeine and alcohol and arrived at the 119 laboratory >2-h post-prandial<sup>(27, 28)</sup>. Participants were instructed to abstain from consumption of *n*-3 120 PUFA supplements and eat no more than 2 oily fish meals per week for 3 weeks prior to the study 121 and throughout the study<sup>(11)</sup>. Participants were free from acute upper respiratory tract infections

- 122 throughout the study.
- 123 Experimental Design and Protocol

This study was conducted in accordance with the Declaration of Helsinki and all procedures were approved by the Nottingham Trent University Human Ethics Committee (Approval No. 186; Clinical trial No. ISRCTN80857707). The study adopted a counter balanced, double-blind, placebocontrolled crossover design over 14 consecutive weeks (Figure 1).

128 Participants were randomised (block randomisation) to receive three 21 day treatments each 129 separated by a 14 day washout period. All treatments involved daily oral consumption of 8 capsules 130 (CRODA International Plc, Snaith, UK). Four capsules were taken in the morning and four in the 131 afternoon with a recommendation to take with food. The three treatments were: 6.2 g/d n-3 PUFA (3.7 g EPA and 2.5 g DHA), 3.1 g/d n-3 PUFA (1.8 g EPA and 1.3 g DHA) (INCROMEGA<sup>TM</sup> 132 TG4030, CRODA, UK) and placebo (CRODAMOL<sup>™</sup> GTCC medium chain triglyceride, CRODA 133 International Plc, Snaith, UK) (Table 2). CRODAMOL<sup>TM</sup> GTCC was chosen as it is readily 134 oxidised in the liver so has little impact on human health-related biomarkers<sup>(29, 30)</sup>. Measurements 135 were taken at day 0 and 21 of each treatment period. The 6.2 g/d n-3 PUFA dose was comprised of 136 137 four n-3 PUFA capsules in the morning and four n-3 PUFA capsules in the afternoon. To ensure an 138 equal number of capsules were taken the 3.1 g/d n-3 PUFA comprised of two n-3 PUFA capsules 139 and two placebo capsules in the morning and two n-3 PUFA capsules and two placebo capsules in 140 the afternoon. All capsules (placebo and *n*-3 PUFA) were identical in appearance.

141 *Measurement of Pulmonary Function, EVH and Fraction of Exhaled Nitric Oxide (F<sub>E</sub>NO)* 

The EVH test was undertaken at day 0 and day 21 of each treatment and comprised 6 min of breathing dry gas at a target minute ventilation ( $\dot{V}_E$ ) of 85% of the predicted maximal voluntary ventilation (MVV) (30 x baseline FEV<sub>1</sub>). Pulmonary function (forced vital capacity, FVC; FEV<sub>1</sub>; peak expiratory flow, PEF; and forced expiratory flow 25-75%, FEF<sub>25-75%</sub>) was assessed according to ATS/ERS guidelines<sup>(31)</sup> in triplicate at baseline and in duplicate at 3, 6, 16, 20, and 30 min after

EVH, as previously described. The highest values recorded were used for analyses. Baseline  $F_ENO$ was measured (NIOX MINO; Aerocrine, Solna, Sweden) according to ATS/ERS guidelines<sup>(32)</sup> in

149 the HIB group only, as it is elevated in asthma patients but not in healthy  $controls^{(33)}$ .

## 150 Urinary 9α, $11\beta$ -PGF<sub>2</sub> Analysis

151 Participants provided a urine sample at baseline and at 12, 60, and 90 min after EVH. Urinary concentration of  $9\alpha$ ,  $11\beta$ -PGF<sub>2</sub>, a metabolite of prostaglandin D<sub>2</sub> was subsequently 152 153 determined by enzyme-linked immunosorbent assay (Caymen Chemicals, Ann Arbor, Michigan, 154 USA) and standardised for urinary creatinine concentration (ABX Pentra 400; Horiba, Northampton, UK) based on a kinetic method using alkaline picrate (Jaffe method)<sup>(34)</sup>. The 155 156 corrected  $9\alpha$ , 11 $\beta$ -PGF<sub>2</sub> was expressed as ng/mmol creatinine. The inter- and intra-assay coefficient 157 of variation was <15%. Two control participants and one HIB participant had samples at day 0 158 below the limit of detection for urinary  $9\alpha$ , 11 $\beta$ -PGF<sub>2</sub> (5 pg/ml) and were subsequently excluded 159 from analysis.

## 160 Neutrophil Phospholipid Fatty Acid Analysis

The neutrophil phospholipid fatty acid composition was assessed as a measure of 161 162 compliance to the treatments in addition to count of capsules returned after each treatment. 163 Neutrophil cells were isolated from 20 mL of whole venous blood (drawn at baseline from an 164 antecubital vein) through a 3 step purification protocol that consisted of dextran sedimentation 165 (Fisher Scientific, Leicestershire UK), hypotonic lysis and Ficoll Paque sedimentation (GE Healthcare, Buckinghamshire, UK)<sup>(35)</sup>. This method achieved up to 98% of pure neutrophils which 166 were stored at -80°C under nitrogen until extraction of phospholipids using previously described 167 methods<sup>(36)</sup>. Fatty acid composition was analysed by gas chromatography as previously 168 described<sup>(37)</sup>. To identify the fatty acid methyl esters (FAME), retention times were compared 169 170 against known standards, Supelco 37 component FAME mix and PUFA-3 menhaden oil (Sigma, 171 Dorset, UK). EPA, DHA, arachidonic acid and linoleic acid were expressed as a percentage of total 172 fatty acids.

#### **173** *Statistical Analysis*

The average minimum perceptible improvement in  $FEV_1$  in adults with asthma is 230 mL<sup>(38)</sup>, whereas the within participant standard deviation for the fall in  $FEV_1$  after EVH is 100 mL<sup>(21)</sup>. *A priori* sample size calculation revealed that with power = 0.90 and alpha = 0.05, a sample size of 7 in the HIB group would be required to detect a 230 mL improvement in the fall in  $FEV_1$ after EVH.

Data were analysed using SPSS (Chicago, IL). Following assessment for normality 179 180 (Shapiro-Wilks test, skewness and kurtosis), data were analysed using repeated measures ANOVA 181 and Bonferroni adjusted paired t-tests. Statistical significance was set at P < 0.05. Data presented 182 are mean (SD) unless otherwise stated. Within treatment percent differences (from day 0 to day 21) 183 for the fall in FEV<sub>1</sub> following EVH were calculated as: ((% fall day 0 - % fall day 21)/% fall day 0)184  $\times$  100. For the HIB group, the individual percent protection afforded by the *n*-3 PUFA treatments 185 compared with placebo was calculated for the % fall in FEV<sub>1</sub> following EVH as: ((% fall day 21 186 placebo - % fall day 21 treatment) / % fall day 21 placebo) × 100. In the HIB group, the overall 187 severity of HIB was determined by calculating the area under the curve for % fall in  $FEV_1$  after 188 EVH (AUC $_{0-30}$ ) using the trapezoidal rule.

189 Results

190 Pulmonary Function and Ventilation Rate during EVH

Baseline FEV<sub>1</sub> was lower in the HIB group (3.75 (SD 0.81) L) than the control group (4.63 (SD 0.37) L) (P = 0.017), and FVC tended to be lower in the HIB group (4.66 (SD 0.06) L) than the control group (5.26 (SD 0.39) L) (P = 0.051). There were no within group differences in FEV<sub>1</sub> or FVC measured at day 0 between the three treatments.

195 As expected, at day 0 there was a greater peak fall in  $FEV_1$  after EVH in the HIB group (pooled data: -29 (SD 17) %) than the control group (pooled data: -3 (SD 2) %) (P = 0.001). In the 196 197 control group, there was no effect of treatment or day on the peak fall in  $FEV_1$  after EVH. In the 198 HIB group, there was a treatment  $\times$  day interaction for the peak fall in FEV<sub>1</sub> after EVH (P = 0.011). 199 Further analyses revealed an effect of treatment for the peak fall in FEV<sub>1</sub> after EVH at day 21 (P =200 0.001). Specifically, the peak fall in FEV<sub>1</sub> after EVH was reduced by 34 (SD 14) % (-690 (SD 460) mL) after 6.2 g/d *n*-3 PUFA (mean difference = 310 (SD 150) mL, 95% CI = 185, 432 mL, P =201 202 0.001, effect size = 0.70), and by 30 (SD 11) % (-700 (SD 420) mL) after 3.1 g/d n-3 PUFA (mean difference = 270 (SD 120) mL, 95% CI = 170, 377 mL, P = 0.001, effect size = 0.58). The reduced 203 peak falls in FEV<sub>1</sub> after 6.2 g/d and 3.1 g/d n-3 PUFA were not different (P = 0.834) (Figure 2, 204 205 Table 4). The percent protection afforded by the n-3 PUFA treatments compared with placebo is 206 shown in Table 4. The peak fall in  $FEV_1$  was unchanged from day 0 to day 21 of placebo.

Figure 3 shows the percent change in  $FEV_1$  during 30 min recovery after EVH in the HIB group. At day 0, the percent change in  $FEV_1$  during recovery was not different between treatments. At day 21, the percent change in  $FEV_1$  was reduced for up to 20 and 30 min recovery after 6.2 g/d and 3.1 g/d *n*-3 PUFA, respectively. Furthermore, compared with placebo at day 21, the percent

change in FEV<sub>1</sub> was lower for up to 6 and 20 min recovery after 6.2 g/d and 3.1 g/d n-3 PUFA,
respectively.

In the HIB group, there was a treatment x day interaction for AUC<sub>0-30</sub> (P = 0.004). Further analysis revealed that at day 21 the AUC<sub>0-30</sub> was reduced after 6.2 g/d (-415 (SD 382), P = 0.002) and 3.1 g/d (-398 (SD 399), P = 0.001) *n*-3 PUFA compared with placebo (-595 (SD 424)). The

**216** AUC<sub>0-30</sub> at day 21 of 6.2 g/d and 3.1 g/d *n*-3 PUFA was not different (P = 0.751).

Consistent with our previous findings<sup>(21, 39)</sup> the peak fall in FEV<sub>1</sub> after EVH in the HIB 217 218 group was reproducible with no differences occurring between day 0 of the three treatments (within 219 participant CV = 7 (SD 4) %; measurement error = 84 mL; reproducibility = 231 mL; smallest 220 meaningful change = 115 mL). The peak fall in FVC showed similar outcomes. At day 0 of the 221 three treatments the peak fall in FVC was greater in the HIB group (pooled data: -992 (SD 604) mL; 222 -21 (SD 15) %) than the control group (pooled data: -140 (SD 90) mL; -3 (SD 2) %) (P = 0.004). In 223 the control group, the peak fall in FVC was unchanged after placebo and both doses of n-3 PUFA. 224 The peak fall in FVC after EVH at day 0 was reduced by 30 (SD 21) % after 6.2 g/d n-3 PUFA and 225 by 29 (SD 24) % after 3.1 g/d n-3 PUFA but was unchanged after placebo in the HIB group. The 226 reduced peak fall in FVC after 6.2 g/d and 3.1 g/d n-3 PUFA was not different (P = 0.847). In the 227 HIB group, the peak fall in PEF was unchanged after placebo and both n-3 PUFA treatments 228 (pooled data: -2.52 (SD 1.68) L/s). In the HIB group, the peak fall in FEF<sub>25-75%</sub> was reduced from day 0 (-1.55 (SD 0.37) L/s) to day 21 (-1.24 (SD 0.45) L/s) of 6.2 g/d n-3 PUFA (mean difference = 229 0.31 (SD 0.23) L/s, 95% CI = 0.12, 0.50 L/s, P = 0.006, effect size = 0.77). In the HIB group, the 230 231 peak fall in FEF<sub>25-75%</sub> was unchanged after 3.1g/d *n*-3 PUFA and placebo in the HIB group, and 232 after all treatments in the control group (P > 0.05).

Minute ventilation achieved during each of the six EVH trials did not differ in the HIB group (P > 0.05; between trial CV = 0.23) (pooled data: 109.5 ± 21.4 L/min; 78 ± 18% of MVV target) which was less compared with the control group (P = 0.001; between trial CV = 0.09) (pooled data: 133.4 ± 10.8 L/min; 60 ± 6% of MVV target).

237 Fraction of Exhaled Nitric Oxide

In the HIB group, there was a treatment × day interaction for  $F_ENO$  (P = 0.004). After 6.2 g/d *n*-3 PUFA,  $F_ENO$  was reduced by 24% from day 0 (48 (SD 33) ppb) to day 21 (35 (SD 28) ppb) (mean difference = 13 (SD 12) ppb, 95% CI = 3, 24 ppb, P = 0.020, effect size = 0.41). Similarly, after 3.1 g/d *n*-3 PUFA,  $F_ENO$  was reduced by 31% from day 0 (49 (SD 33) ppb) to day 21 (34 (SD

- 242 28) ppb) (mean difference = 15 (SD 14) ppb, 95% CI = 4, 27 ppb, P = 0.018, effect size = 0.46)
- 243 (Figure 4). The reduced  $F_ENO$  after 6.2 g/d and 3.1 g/d *n*-3 PUFA was not different (P = 0.491)
- (Figure 4).  $F_ENO$  remained unchanged from day 0 (47 (SD 26) ppb) to day 21 (47 (SD 27) ppb) of
- 245 placebo.

246 Urinary  $9\alpha$ ,  $11\beta PGF_2$ 

Baseline urinary  $9\alpha$ , 11ß PGF<sub>2</sub> did not differ between the HIB group (day 0 pooled data: 40.85 (SD 21.49) ng/mmol creatinine) and the control group (day 0 pooled data: 27.05 (SD 14.01) ng/mmol creatinine) (P = 0.133). There were no within-group differences in urinary  $9\alpha$ , 11ß PGF<sub>2</sub> measured before EVH at day 0 and day 21 of the three treatments.

251 At day 0, urinary 9a, 11B PGF<sub>2</sub> increased by 31.53 (SD 22.77) ng/mmol creatinine (pooled 252 data) after EVH in the HIB group (P = 0.030) but not in the control group (pooled data: 11.11 (SD 253 16.30) ng/mmol creatinine). At day 0, changes in  $9\alpha$ , 11B PGF<sub>2</sub> after EVH were different between 254 groups (P = 0.003). In the HIB group, subsequent analyses revealed an effect of treatment on the 255 peak increase in 9 $\alpha$ , 11B PGF<sub>2</sub> after EVH at day 21 (P = 0.014). Specifically, the peak increase in 256  $9\alpha$ , 11B PGF<sub>2</sub> after EVH was reduced after 6.2 g/d *n*-3 PUFA (mean difference = 11.75 (SD 7.42)) 257 ng/mmol creatinine, 95% CI = 5.54, 17.96 ng/mmol creatinine, P = 0.009, effect size = 0.50) and 258 3.1 g/d *n*-3 PUFA (mean difference = 12.86 (SD 14.90) ng/mmol creatinine, 95% CI = 0.40, 25.33 259 ng/mmol creatinine, P = 0.041, effect size = 0.40). The reduced peak increase in 9 $\alpha$ , 11B PGF<sub>2</sub> after EVH following 6.2 g/d and 3.1 g/d n-3 PUFA was not different (P = 0.377) (Figure 5). There was 260 261 no effect of placebo on the increase in urinary  $9\alpha$ , 11B PGF<sub>2</sub> after EVH.

## 262 Compliance Assessment - Neutrophil Phospholipid Fatty Acid Content, and Capsule Counts

263 Good compliance to all treatments was shown from the capsule counts in both the HIB and 264 control groups (Table 3). Further compliance evidence was provided from the neutrophil 265 phospholipid fatty acid content. In the HIB group, 6.2 g/d n-3 PUFA increased EPA (P = 0.002) and 266 DHA (P = 0.045) content, and reduced arachidonic acid (P = 0.005). In the HIB group, 3.1 g/d n-3 267 PUFA increased EPA content (P = 0.018) and reduced arachidonic acid (P = 0.009); there was a 268 trend for an increase in DHA content (P = 0.074). In the HIB group, linoleic acid content was 269 unchanged after 6.2 g/d and 3.1 g/d n-3 PUFA. Phospholipid fatty acid content was unchanged after 270 placebo in both groups. In the control group, 6.2 g/d n-3 PUFA tended to increase EPA (P = 0.054) 271 and DHA content (P = 0.087), whereas 3.1 g/d *n*-3 PUFA increased DHA content (P = 0.007) and 272 reduced arachidonic acid (P = 0.038); EPA content was unchanged (Table 3).

## 274 Discussion

275 Previous research has shown that high doses of n-3 PUFA (5.2-5.4 g/d) reduce the severity of HIB and EIB<sup>(10, 11, 15-17)</sup>. The present study demonstrates that a lower dose of 3.1 g/d n-3 PUFA is 276 equally effective in reducing HIB in adult men with asthma. The 6.2 and 3.1 g/d n-3 PUFA 277 278 treatments also resulted in similar reductions in baseline F<sub>E</sub>NO, and comparable suppression of 279 urinary  $9\alpha$ , 11β-PGF<sub>2</sub> after EVH. These findings suggest that 3.1 g/d *n*-3 PUFA could be used as an 280 adjunct therapy for physically active adults with asthma and EIB. The percent protection afforded 281 by 6.2 g/d n-3 PUFA (35 (SD 14) %) and 3.1 g/d n-3 PUFA (33 (SD 12) %) highlights the efficacy of *n*-3 PUFA as an adjunct therapy. The percent protection afforded by *n*-3 PUFA is comparable to 282 montelukast (50mg twice daily)<sup>(40)</sup> but less than the 60-70% protection afforded by short-acting  $\beta_2$ -283 agonists procaterol (10 micrograms/inhalation) and albuterol (90 micrograms/inhalation)<sup>(41)</sup>. Long-284 acting  $\beta_2$ -agonists also provide effective treatment for EIB,<sup>(42-44)</sup> although chronic treatment with 285 both long- and short-acting  $\beta_2$ -agonists can result in tolerance<sup>(45, 46)</sup>. Furthermore, compliance to the 286 287 lower 3.1 g/d n-3 PUFA dose may be improved due to reduced capsule numbers, reduce cost and 288 risk of gastrointestinal distress, which is commonly associated with bloating, and stomach upset. Indeed, two participants withdrew from the study when on the 6.2 g/d n-3 PUFA treatment due to 289 290 gastrointestinal distress, whereas full compliance and no participant withdrawals (capsule counts) 291 was observed with 3.1 g/d n-3 PUFA.

292 The EVH test is an indirect bronchial provocation test that is a suitable objective surrogate for identifying EIB<sup>(24)</sup>. We have previously reported that the EVH protocol used in the present study 293 elicits a highly reproducible fall in  $\text{FEV}_1^{(21, 39)}$  which is crucial when assessing treatment efficacy. 294 The ~290 mL (~32%) decrease in the post-EVH fall in FEV<sub>1</sub> after 6.2 g/d and 3.1 g/d n-3 PUFA 295 supplementation exceeds the minimum perceptible change of 230 mL<sup>(38)</sup> and is therefore clinically 296 relevant. Our findings are in agreement with previous studies showing an attenuation of HIB<sup>(17)</sup> and 297  $EIB^{(11)}$  after *n*-3 PUFA supplementation in adults with asthma. These findings collectively support 298 the ATS/ERS guidance<sup>(25)</sup> suggesting n-3 PUFA supplementation could be of benefit in individuals 299 300 with EIB. Our findings are also in broad agreement with work on n-3 PUFA supplementation in elite athletes with EIB<sup>(10)</sup>, and asthmatic patients with EIB and HIB<sup>(11, 15-17)</sup>. An interesting 301 difference is that previous work showed abolition of EIB after n-3 PUFA supplementation<sup>(10)</sup>. 302 303 whereas in the present study only 3 participants became non-diagnostic (Figure 2). This may be 304 partly due to inter-study differences in the severity of EIB: the peak fall in  $FEV_1$  in the present study (-30%) was ~50% greater than that reported in previous *n*-3 PUFA supplementation studies demonstrating a reduction in EIB/HIB<sup>(11, 15-17)</sup>.

Unlike the rather consistent finding that pharmacological therapy reduces EIB<sup>(42-44)</sup>, the 307 efficacy of n-3 PUFA remains controversial. Our findings contrast those of Arm et al.<sup>(9)</sup> and 308 Brannan et al.<sup>(12)</sup> who despite using relatively high n-3 PUFA doses (5.4-6.0 g/d), reported no 309 change in bronchial hyperresponsiveness to inhaled histamine and mannitol, respectively. Arm et 310 al.<sup>(9)</sup> reported no change in airway resistance (fall in FEV<sub>1</sub> was not measured) after a cycling 311 exercise challenge, although their data are confounded by: (1) the suggestion that low-grade, non-312 pharmaceutical n-3 PUFA was used<sup>(47)</sup>; (2) low statistical power due to the small number of 313 participants (n = 6) performing the exercise; (3) exercise being performed at ambient temperature 314 315 and humidity, which may result in insufficient environmental stress; and (4) the low exercise 316 intensity (60-130 W) which likely (data not measured/reported) elicited only a modest increase in  $\dot{V}_{E}$ . The efficacy of *n*-3 PUFA supplementation may also partly depend on the choice of bronchial 317 318 provocation test. Direct bronchial provocation tests such as inhaled histamine act directly on receptors on the airway smooth muscle causing contraction<sup>(48)</sup> and may not, therefore, fully reveal 319 320 the potential anti-inflammatory effects of n-3 PUFA. Indirect bronchial provocation tests such as 321 mannitol and EVH both alter the tonicity and volume of the airway surface liquid thereby promoting the release of bronchoconstrictive inflammatory mediators<sup>(49, 50)</sup>. However, mannitol and 322 EVH may exert different levels of osmotic and mechanical stress on the airways. Specifically, 323 mannitol induces dehydration stress primarily in the proximal airways<sup>(50, 51)</sup>, whereas the high  $\dot{V}_E$ 324 during EVH results in the dehydration stress being extended to peripheral airways<sup>(52)</sup> which are 325 326 increasingly recruited to heat and humidify inspired air. Furthermore, EVH also elicits airflowinduced shear stress and high transepithelial pressure gradients<sup>(53)</sup>. These differences may 327 predispose the airway epithelium to greater damage after EVH than mannitol; indeed, EVH tends to 328 329 result in greater increases in urinary Clara cell protein 16 concentration, a marker of airway epithelial damage<sup>(54, 55)</sup>. This is significant because n-3 PUFA supplementation attenuates the 330 increase in urinary Clara cell protein concentration after EVH, which suggests that reduced HIB 331 may be partly due to reduced airway epithelial damage<sup>(56)</sup>. Differences in the intensity of the 332 stimulus to the airways may thus explain the divergent effects of n-3 PUFA supplementation on 333 334 bronchial hyperresponsiveness to EVH and mannitol.

Recently, Price et al.<sup>(18)</sup> reported no change in the severity of HIB or markers of airway inflammation ( $F_ENO$ , urinary 9 $\alpha$ , 11 $\beta$  PGF<sub>2</sub> and cysteinyl leukotriene E4) in recreationally active

337 adults supplemented daily with a combined treatment of 30  $\mu$ g vitamin D3 with *n*-3 PUFA (3.0 g EPA and 2.0 g DHA). Unfortunately, objective treatment compliance measures (e.g. neutrophil 338 339 phospholipid DHA and EPA content) were not taken in this study. The fall in  $FEV_1$  after EVH was 340 also modest (~16%) compared with the ~30% fall observed at day 0 in the present study, and 6 341 from 10 of the participants did not have physician diagnosed asthma (and consequently no asthma medication). Two participants in Price et al.<sup>(18)</sup> also became non-diagnostic for HIB after the 342 placebo treatment. It may be that *n*-3 PUFA supplementation is thus more effective in individuals 343 344 with diagnosed asthma and more severe HIB, as observed in the present study.

345 The increase in urinary  $9\alpha$ ,  $11\beta$ -PGF<sub>2</sub> after EVH at day 0 in the HIB group is similar in magnitude to that reported previously in adults with asthma performing exercise or EVH<sup>(10, 57)</sup>. The 346 347 reduced increase in  $9\alpha$ , 11B-PGF<sub>2</sub> after EVH was comparable after 6.2 g/d and 3.1 g/d *n*-3 PUFA supplementation. Previous work also reports a reduced increase in  $9\alpha$ ,  $11\beta$ -PGF<sub>2</sub> after EVH 348 following n-3 PUFA supplementation<sup>(10)</sup>. The increased EPA and DHA cell membrane content 349 following n-3 PUFA supplementation is likely to result in dual inhibition of arachidonic acid-350 351 dependent cyclooxygenase and 5-lipoxygenase pathways which are responsible for eicosanoid generation<sup>(7)</sup>. This may explain the decrease in  $9\alpha$ ,  $11\beta$ -PGF<sub>2</sub>, a metabolite of the arachidonic acid-352 derived pro-inflammatory eicosanoid prostaglandin- $D_2$ . The osmotic changes in the airways 353 following EVH results in mast cell activation and release of PGD2 during bronchoconstriction<sup>(49, 50)</sup>. 354 Suppression of mast cell activation, and reduced eicosanoid generation, may thus partially explain 355 356 reduced HIB after *n*-3 PUFA supplementation.

357 The comparable reduction in baseline  $F_ENO$  after 6.2 g/d and 3.1 g/d n-3 PUFA supplementation concurs with previous work<sup>(17, 58)</sup> and is indicative of reduced baseline eosinophilic 358 359 airway inflammation. Increased F<sub>E</sub>NO can also result from elevated expression of inducible nitric oxide synthase in T-cells, macrophages, airway epithelial cells, and other inflammatory cells within 360 the airways<sup>(59, 60)</sup>. Elevated expression of inducible nitric oxide synthase can be induced by certain 361 pro-inflammatory cytokines such as TNF- $\alpha^{(59, 60)}$  probably via the activation of nuclear factor-kappa 362  $\beta$  transcription factor<sup>(61)</sup>. The *n*-3 and *n*-6 PUFAs modulate nuclear factor-kappa  $\beta$  transcription 363 factor activation<sup>(62)</sup> and presumably therefore TNF- $\alpha$  expression, which may explain the observed 364 fall in F<sub>E</sub>NO after *n*-3 PUFA supplementation in the present study. Support for this argument comes 365 366 from work showing that 5.4 g/d n-3 PUFA suppresses circulating plasma TNF-α in athletes with EIB<sup>(10)</sup>. The comparable reduction in both baseline  $F_ENO$  and the post-EVH increase in  $9\alpha$ , 11β-367 PGF<sub>2</sub> after 6.2 g/d and 3.1 g/d n-3 PUFA supplementation suggests that both doses increased the n-368

369 3 PUFA content of the phospholipid bilayer of cell membranes to influence the inflammatory 370 response. Furthermore, and consistent with previous reports<sup>(17, 56)</sup>, the lower  $F_ENO$  observed after *n*-371 3 PUFA supplementation in the present study may have resulted from an increase in airway pH, 372 which is considered a determinant of  $F_ENO$  and airway inflammation<sup>(63)</sup>.

373 Limitations of the current study include no run-in period to assess habitual *n*-3 PUFA intake, the exclusion of females, the comparatively low participant numbers, and the wide range of asthma 374 375 phenotypes suggested by the heterogeneous fall in  $FEV_1$  and large variation in inflammatory 376 markers after EVH. However, our cohort provides evidence for the use of n-3 PUFA in more severe HIB than previous literature<sup>(10, 11)</sup>. Future work may explore whether particular asthma phenotypes 377 378 and HIB/EIB severity respond preferentially to n-3 PUFA supplementation. Furthermore, given the 379 existing inter-study differences regarding the efficacy of *n*-3 PUFA for the management of EIB, further studies using larger sample sizes are warranted. 380

In conclusion, 3.1 g/d *n*-3 PUFA supplementation effectively reduced HIB in men with asthma to a similar extent as a higher dose. Lower doses of *n*-3 PUFA thus represent a potentially beneficial adjunct treatment for adults with asthma and EIB, whilst also reducing the burden of cost, compliance and potential gastrointestinal distress. Further studies are needed to elucidate if the bronchial provocation test used and the severity of asthma and HIB/EIB affect the degree of protection afforded by *n*-3 PUFA supplementation.

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N.C.W., K.A.H., D.E.S., G.R.S., and M.A.J designed the research; N.C.W., K.A.H., G.R.S.,
and M.A.J conducted research; N.C.W., and K.G.J provided essential reagents and conducted
analysis of urine and blood samples; N.C.W., G.R.S., and M.A.J analysed data. N.C.W., K.A.H.,
G.R.S., and M.A.J wrote paper; N.C.W., K.A.H., D.E.S., K.G.J., G.R.S., and M.A.J., contributed to
reviewing and approval of the final manuscript.

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- 401 None of the other authors had a personal or financial conflict of interest.
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- 405 **TABLE 1**. Individual and mean (SD) anthropometric data, baseline pulmonary function, and
- 406 medication.

				FVC (L	.)	FEV <sub>1</sub> (	L)	
	Age (y)	Height (cm)	Body Mass (kg)	n	%	п	%	Medications
HIB								
1	21	177	75	5.54	106	4.54	103	S
2	44	178	87	4.22	94	3.17	87	S, BUD+FORM
3	21	173	58	3.78	76	2.77	65	S, BUD+FORM
4	38	180	77	4.62	95	3.80	95	S
5	38	173	83	4.43	91	3.49	90	S
6	28	177	68	4.91	96	4.31	100	S
7	22	181	82	5.70	107	5.10	114	S
8	31	173	75	4.10	91	2.84	76	S, BEC
Mean (SD)	30 (9)	177 (3)	76 (9)	4.66 (0.68)	95 (10)	3.75 (0.84)*	91 (16)	
Control								
1	30	183	80	5.30	98	4.15	92	
2	20	184	80	5.93	106	5.18	110	
3	20	178	69	4.61	88	4.49	101	
4	28	170	84	4.87	103	4.18	104	
5	26	184	70	5.23	93	4.90	104	
6	23	189	92	5.34	91	5.00	102	
7	27	177	69	5.31	103	4.45	103	
8	27	181	88	5.46	101	4.69	104	
Mean (SD)	25 (4)	181 (6)	79 (9)	5.26 (0.39)	98 (7)	4.63 (0.38)	103 (7)	

- 407 BEC = beclomethasone; BUD = budesonide; FORM = formoterol;  $FEV_1$  = forced expiratory volume in 1 second; FVC
- 408 = forced vital capacity; S = salbutamol. \*Difference between HIB and control group (P = 0.017).

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- TABLE 2. Fatty acid composition (% total fatty acids) of the *n*-3 PUFA (INCROMEGA<sup>TM</sup>) 412
- TG4030) and placebo (CRODAMOL<sup>TM</sup> GTCC, medium chain triglycerides) treatments. Individual 413 fatty acids making up  $\geq 1\%$  are shown.
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			% total fatty acids			
	Fatty Acid	Name	n-3 PUFA (INCROMEGA™ TG4030)	Placebo (CRODAMOL <sup>™</sup> GTCC)		
	8:0	Caprylic acid		56		
	10:0	Capric acid		43		
	18:4 <i>n</i> -3	Octadecatetraenoic acid	3			
	20:4 <i>n</i> -6	Arachidonic acid	3			
	20:4 <i>n</i> -3	Eicosatetraenoic acid	2			
	20:5 n-3	Eicosapentaenoic acid	45			
	21:5 <i>n</i> -3	Heneicosapentaenoic acid	-2			
	22:5 n-3	Docosapentaenoic acid	6			
	22:6 n-3	Docosahexaenoic acid	31			
	Total		92	99		
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**TABLE 3.** Treatment adherence (%) based on capsule count, and fatty acid composition of
neutrophil extracts expressed as a percent weight of total fatty acids at day 0 and day 21 of each
treatment in HIB and control groups. Data are mean (SD).

	Plac	cebo	6.2 g/d	n-3 PUFA	PUFA 3.1 g/d	
HIB	Day 0	Day 21	Day 0	Day 21	Day 0	Day 21
20:4 <i>n</i> -6 Arachidonic Acid	6.52 (0.82)	6.34 (0.62)	6.03 (0.78)	5.37 (0.80)**	6.35 (0.92)	5.26 (0.99)**
20:5 <i>n</i> -3 EPA	0.72 (0.17)	0.75 (0.19)	0.77 (0.19)	1.39 (0.38)**	0.59 (0.29)	1.05 (0.31)*
22:6 <i>n</i> -3 DHA	2.15 (0.33)	2.20 (0.37)	2.06 (0.33)	2.39 (0.31)*	2.14 (0.40)	2.29 (0.35)
Adherence (%)		93 (4)		94 (4)		95 (4)
Control						
20:4 n-6 Arachidonic Acid	5.18 (1.02)	5.09 (1.34)	5.20 (0.63)	4.94 (0.56)	5.73 (1.29)	5.02 (1.01)*
20:5 <i>n</i> -3 EPA	0.80 (0.19)	0.79 (0.22)	0.70 (0.20)	0.93 (0.12)	0.73 (0.21)	0.84 (0.10)
22:6 <i>n</i> -3 DHA	1.98 (0.31)	2.14 (0.45)	2.02 (0.28)	2.30 (0.30)	1.83 (1.40)	2.22 (0.26)**
Adherence (%)		93 (6)		92 (5)		93 (4)

431 DHA = Docosahexaenoic acid; EPA = Eicosapentaenoic acid; HIB = Hyperphoea-induced bronchoconstriction.

Difference between day 0 and day 21 within treatment (\* P < 0.05; \*\* P < 0.01). 

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445	<b>TABLE 4</b> . Individual and mean (SD) peak % fall in FEV <sub>1</sub> at day 0 and day 21 of each treatment
446	and percent protection afforded by <i>n</i> -3 PUFA. Data shown for HIB group only.

	Peak % fall in FEV <sub>1</sub>						Percent protection affe	orded by n-3 PUFA
	Placebo		6.2 g/day <i>n</i> -3 PUFA		3.1 g/day <i>n</i> -3 PUFA		treatments for peak % fall in $\ensuremath{FEV}_1$	
HIB	Day 0	Day 21	Day 0	Day 21	Day 0	Day 21	6.2 g/day <i>n</i> -3 PUFA	3.1 g/day <i>n</i> -3 PUFA
1	-12.58	-13.10	-13.02	-6.54	-14.95	-9.98	50	24
2	-63.22	-64.81	-61.04	-51.36	-60.32	-49.84	21	23
3	-48.73	-47.40	-48.20	-33.70	-48.92	-32.83	29	31
4	-32.90	-28.10	-28.50	-19.54	-30.45	-22.91	30	18
5	-27.02	-32.64	-29.71	-14.98	-26.44	-23.12	54	29
6	-19.72	-18.01	-17.33	-11.67	-18.41	-10.50	35	42
7	-17.74	-16.21	-16.05	-8.64	-13.04	-8.63	47	47
8	-14.83	-15.82	-15.71	-13.67	-13.78	-7.90	14	50
Mean (SD)	-30 (18)	-30 (18)	-29 (17)	-20 (15)	-28 (18)	-21 (15)	35 (14)	33 (12)

- Percent protection afforded by the *n*-3 PUFA treatments calculated as: ((% fall day 21 placebo % full day 21 placebo % full day 21 placebo %
- fall day 21 treatment) / % fall day 21 placebo)  $\times$  100.

- 450 Figure Captions
- FIGURE 1 Participant flow diagram. HIB = hyperphoea-induced bronchoconstriction; CTRL = controls; PUFA = polyunsaturated fatty acid; GI = gastrointestinal.
- 453 FIGURE 2 Peak falls in forced expiratory volume in 1 s (FEV1) before and after placebo (A), 6.2 g/d n-3 PUFA (B),
- and 3.1 g/d n-3 PUFA (C) in the HIB group (n = 8). Bars represent mean (SD) and identical symbols represent the same HIB participant. \*Day 0 v. day 21 (P = 0.001).
- 456 FIGURE 3 The percent change in forced expiratory volume in 1 second (FEV<sub>1</sub>) after eucapnic voluntary hyperphoea
- 457 in participants with hyperphoea-induced bronchoconstriction (HIB). A 10% fall in FEV<sub>1</sub> (shown by the dashed line) is
- 458 diagnostic of HIB. Closed circles, day 0 of placebo treatment; open circles, day 21 of placebo treatment; closed
- 459 squares, day 0 of 6.2 g/d n-3 PUFA treatment; open squares, day 21 of 6.2 g/d n-3 PUFA; closed triangles, day 0 of 3.1
- 460 g/d n-3 PUFA; open triangles, day 21 of 3.1 g/d n-3 PUFA. Significant differences indicated by letters: a, 6.2 g/d n-3
- 461 PUFA vs. placebo; b, 3.1 g/d *n*-3 PUFA vs. placebo; c, day 0 vs. day 21 of 6.2 g/d *n*-3 PUFA; d, day 0 vs. day 21 of 3.1
- 462 g/d *n*-3 PUFA. Single letters, P < 0.05; \*P < 0.01; \*\*P < 0.001.
- 463 FIGURE 4 Baseline fraction of exhaled nitric oxide (FENO) before and after placebo (A), 6.2 g/d n-3 PUFA (B), and
- 3.1 g/d n-3 PUFA (C) in the HIB group (n = 8). Bars represent mean (SD) and identical symbols represent the same
- 465 HIB participant. \*Day 0 v. day 21 (P < 0.05).

- 466 FIGURE 5 - Peak increase in urinary  $9\alpha$ , 11B PGF2 after EVH in the HIB group (n = 8). Filled and open bars represent 467 day 0 and day 21, respectively, of the treatment. Data are mean (SD). \*Day 0 v. day 21 (P < 0.05468 469 470 References 471 1. Carlsen K, Anderson S, Bjermer L et al. (2008) Exercise induced asthma, respiratory and allergic disorders in elite 472 athletes: epidemiology, mechanisms and diagnosis: Part I of the report from the Joint Task Force of the European 473 Respiratory Society (ERS) and the European Academy of Allergy and Clinical Immunology (EAACI) in cooperation 474 with GA2LEN. Allergy 63(4), 387-403. 475 2. Weiler JM, Anderson SD, Randolph C et al. (2010) Pathogenesis, prevalence, diagnosis, and management of 476 exercise-induced bronchoconstriction: a practice parameter. Ann Allergy Asthma Immunol 105(6), S1-S47. 477 3. Hallstrand TS (2012) New insights into pathogenesis of exercise-induced bronchoconstriction. Curr Opin Allergy 478 Clin Immunol 12(1), 42-48. 479 4. Hallstrand TS, Altemeier WA, Aitken ML et al. (2013) Role of cells and mediators in exercise-induced 480 bronchoconstriction. Immunology Allergy Clinics North Am 33(3), 313-328. 481 5. Barnes PJ (2010) New therapies for asthma: is there any progress? Trends Pharmacol Sci 31(7), 335-343.
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FIGURE 1 – Participant flow diagram. HIB = hyperpnoea-induced bronchoconstriction; CTRL = controls; PUFA = polyunsaturated fatty acid; GI = gastrointestinal.

266x355mm (96 x 96 DPI)



FIGURE 2 – Peak falls in forced expiratory volume in 1 s (FEV1) before and after placebo (A), 6.2 g/d n-3 PUFA (B), and 3.1 g/d n-3 PUFA (C) in the HIB group (n = 8). Bars represent mean (SD) and identical symbols represent the same HIB participant. \*Day 0 v. day 21 (P = 0.001).

98x202mm (300 x 300 DPI)



FIGURE 3 – The percent change in forced expiratory volume in 1 second (FEV1) after eucapnic voluntary hyperpnoea in participants with hyperpnoea-induced bronchoconstriction (HIB). A 10% fall in FEV1 (shown by the dashed line) is diagnostic of HIB. Closed circles, day 0 of placebo treatment; open circles, day 21 of placebo treatment; closed squares, day 0 of 6.2 g/d n-3 PUFA treatment; open squares, day 21 of 6.2 g/d n-3 PUFA; closed triangles, day 0 of 3.1 g/d n-3 PUFA; open triangles, day 21 of 3.1 g/d n-3 PUFA. Significant differences indicated by letters: a, 6.2 g/d n-3 PUFA vs. placebo; b, 3.1 g/d n-3 PUFA vs. placebo; c, day 0 vs. day 21 of 6.2 g/d n-3 PUFA; d, day 0 vs. day 21 of 3.1 g/d n-3 PUFA. Single letters, P < 0.05; \*P < 0.01; \*\*P < 0.001.

104x72mm (300 x 300 DPI)



FIGURE 4 – Baseline fraction of exhaled nitric oxide (FENO) before and after placebo (A), 6.2 g/d n-3 PUFA (B), and 3.1 g/d n-3 PUFA (C) in the HIB group (n = 8). Bars represent mean (SD) and identical symbols represent the same HIB participant. \*Day 0 v. day 21 (P < 0.05).

100x202mm (300 x 300 DPI)



FIGURE 5 - Peak increase in urinary 9a, 11 $\beta$  PGF2 after EVH in the HIB group (n = 8). Filled and open bars represent day 0 and day 21, respectively, of the treatment. Data are mean (SD). \*Day 0 v. day 21 (P < 0.05

145x104mm (300 x 300 DPI)