

Interspecific variation, habitat complexity and ovipositional responses modulate the efficacy of cyclopoid copepods in disease vector control

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1 **Interspecific variation, habitat complexity and ovipositional responses**
2 **modulate the efficacy of cyclopoid copepods in disease vector control**

3

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21 **ABSTRACT**

22 The use of predatory biological control agents can form an effective component in the
23 management of vectors of parasitic diseases and arboviruses. However, we require predictive
24 methods to assess the efficacies of potential biocontrol agents under relevant environmental
25 contexts. Here, we applied functional responses (FRs) and reproductive effort as a proxy of
26 numerical responses (NRs) to compare the Relative Control Potential (RCP) of three
27 cyclopoid copepods, *Macrocyclops albidus*, *M. fuscus* and *Megacyclops viridis* towards
28 larvae of the mosquito *Culex quinquefasciatus*. The effects of habitat complexity on such
29 predatory impacts were examined, as well as ovipositional responses of *C. quinquefasciatus*
30 to copepod cues in pairwise choice tests. All three copepod species demonstrated a
31 population destabilising Type II FR. *M. albidus* demonstrated the shortest handling time and
32 highest maximum feeding rate, whilst *M. fuscus* exhibited the highest attack rate. The
33 integration of reproductive effort estimations in the new RCP metric identifies *M. albidus* as
34 a very promising biocontrol agent. Habitat complexity did not impact the FR form or
35 maximum feeding rate of *M. albidus*, indicating that potentially population destabilising
36 effects are robust to habitat variations; however, attack rates of *M. albidus* were reduced in
37 the presence of such complexity. *C. quinquefasciatus* avoided ovipositing where *M. albidus*
38 was physically present, however it did not avoid chemical cues alone. *C. quinquefasciatus*
39 continued to avoid *M. albidus* during oviposition when both the treatment and control water
40 were dyed; however, when an undyed, predator-free control was paired with dyed, predator-
41 treated water, positive selectivity towards the treatment water was stimulated. We thus
42 demonstrate the marked predatory potential of cyclopoid copepods, utilising our new RCP
43 metric, and advocate their feasibility in biological control programmes targeting container-
44 style habitats. We also show that behavioural responses of target organisms and
45 environmental context should be considered to maximise agent efficacy.

46 Keywords: Functional response; Numerical response; Relative Control Potential; Copepod;
47 Mosquito; Oviposition

48

49 **1. Introduction**

50 Mosquitoes continue to endanger public health globally through the transmission of
51 vector-borne diseases, which account for hundreds of thousands of deaths annually (World
52 Health Organisation, 2017). Transmission rates of arboviruses and the ecology of their
53 vectors are shifting rapidly under environmental change (Medlock and Leach, 2015; Benelli
54 and Mehlhorn, 2016), and at different spatiotemporal scales (Townroe and Callaghan, 2014;
55 Cunze et al. 2016). Furthermore, incursions and proliferations of highly invasive mosquito
56 species are occurring as a direct result of anthropogenic activity (e.g. Yee, 2016; Medlock et
57 al. 2017), with implications for public health and persistence of native species (Juliano, 2010;
58 Schöffner et al. 2013). Thus, the development of effective strategies to assist or augment the
59 control of vectors of mosquito-borne disease is critical. These strategies in turn rely on rapid
60 and reliable assessments of the efficacy of biocontrol agents under relevant environmental
61 contexts.

62 *Culex quinquefasciatus* Say, 1823 is a member of the *C. pipiens* complex, which are
63 vectors of diseases such as West Nile virus, Sandbis virus, St Louis encephalitis, Rift Valley
64 fever virus and lymphatic filariasis (Turell, 2012; Manimegalai and Sukanya, 2014). This
65 mosquito breeds rapidly in artificial, urban environments, enabling a high potential for human
66 contact (Yee, 2008). Furthermore, the species is characterised as being invasive, exhibiting
67 the potential to negatively impact and replace native species (Juliano, 2010). Climate change
68 may additionally bolster the potential of *C. quinquefasciatus* to invade novel ecosystems
69 through range expansion (Samy et al. 2016). Cyclopoid copepods have had marked success in

70 the biological control of *Aedes aegypti* mosquito larvae (e.g. Kay and Nam, 2005), and
71 predatory impacts have been demonstrated towards *Culex* spp. (e.g. Calliari et al. 2003;
72 Tranchida et al. 2009). Furthermore, the global distribution, high fecundity, environmental
73 hardiness, voraciousness and ease of mass-production of copepods enable high levels of
74 potential impact on target populations, whilst negating the need for potentially ecologically
75 harmful species introductions (Marten and Reid, 2007). Yet, while interspecific variations in
76 control efficiencies between cyclopoids have been demonstrated (Marten, 1989), as well as
77 demographic variations across differing species strains (Marten, 1990b), many candidate
78 copepod species are yet to be examined in the context of mosquito control.

79 Functional responses (FRs), which discern the *per capita* intake rate of a resource as a
80 function of its availability (Holling, 1959; 1966), exhibit great utility in the quantification of
81 interspecific differences in consumer-resource interactions (Dick et al. 2017). The search,
82 capture and handling components of FRs can be highly informative in the contexts of
83 biological control (O'Neil, 1990; Van Driesche and Bellows, 2011) and invasive species
84 (Dick et al. 2014). Indeed, high impact invasive species are strongly associated with higher
85 maximum feeding rates driven by low handling times (Dick et al. 2017), and this associates
86 strongly with measurable changes in affected populations (i.e. ecological impact). Functional
87 response form and magnitude can also be significant with respect to prey population stability;
88 a Type II FR is deemed to have potentially population destabilising effects due to high levels
89 of proportional predation at low prey densities, whilst a Type III FR may impart stability to
90 prey by providing refugia at low prey densities (Holling, 1959). Moreover, coupling such *per*
91 *capita* effects with corresponding numerical responses at the consumer population level can
92 bolster the robustness of predictions of consumer impacts on resources (Dick et al. 2017).
93 These approaches thus yield high potential in the comparison of efficacies of potential
94 biocontrol agents. Further, context-dependencies, such as habitat complexity, can modify the

95 nature of interactions within predator-prey systems and can be quantified using a FR
96 approach (e.g. Barrios-O'Neill et al. 2015; South et al.2017). Habitat complexity may enable
97 physical refuge for prey, directly affecting interaction strengths, and, accordingly, the
98 structuring of ecological communities (e.g. Alexander et al. 2012). These interactions are
99 often specific to predator-prey pairings, with habitat complexity having been found to also
100 heighten predation (e.g. Alexander et al. 2015).

101 Besides direct consumptive effects, trait-mediated indirect interactions (TMIIIs),
102 including behavioural responses of target organisms to predators, can exert strong effects on
103 predator-prey dynamics (e.g. Alexander et al. 2013), and can be as strong or stronger in their
104 population effects (Peacor and Werner 2001; Trussell et al. 2004; 2008). Behavioural
105 responses of mosquitoes to predator cues can be marked, yet also species-specific (e.g.
106 Vonesh and Blaustein, 2010; Zuhurah and Lester, 2011). Thus, considering oviposition
107 selectivity among potential habitats by mosquitoes is of integral importance to biological
108 control, particularly for species which invest all of their eggs in one environment at one time,
109 such as *C. quinquefasciatus* (Wachira et al. 2010). Broadly, natural selection favours
110 individuals that are able to actively evade predation through avoidance of oviposition habitats
111 containing predators (Sih, 1986; Blaustein and Chase, 2007). Water colouration, serving as a
112 proxy for nutritional load, has also been demonstrated to significantly attract oviposition by
113 *Culex* mosquitoes (Ortiz Perea and Callaghan, 2017). The effects of the presence of copepods
114 on oviposition of mosquitoes has been seldom considered, however, Torres-Estrada et al.
115 (2001) found that *A. aegypti* preferentially oviposited in water treated with the copepod
116 *Mesocyclops longisetus* due to the emission of attractive compounds. This finding is an
117 oddity in the context of the general avoidance by mosquitoes of other predators (Vonesh and
118 Blaustein, 2010).

119 Here, we quantify the FRs of three locally-abundant and widespread cyclopoid
120 copepods, *Macrocylops albidus* (Jurine 1820), *Macrocylops fuscus* (Jurine, 1820) and
121 *Megacyclops viridis* (Jurine, 1820) towards larvae of the mosquito *C. quinquefasciatus*. We
122 then combine maximum feeding rate and attack rate estimations generated from the FRs with
123 reproductive effort data from Maier (1994) to compare the Relative Control Potential (RCP;
124 Cuthbert et al. 2018) among the three species. Benthic habitat complexity is integrated in a
125 separate experiment to assess potential context-dependencies relevant to the efficacy of
126 copepods in their predation of *C. quinquefasciatus*. Furthermore, we utilise pairwise choice
127 tests to assess the ovipositional responses of *C. quinquefasciatus* to the predators in several
128 bioassays consisting of visual and chemical cues from predatory copepods, and examine
129 whether the integration of pond dye reduces avoidance behaviour by the mosquito, hence
130 enhancing biocontrol efforts. Thus, we seek to decipher the most effective biocontrol agent
131 using the RCP metric, and consider the effects of habitat complexity and target prey
132 behavioural responses on agent efficacies.

133 2. Methods

134 2.1. Experimental organisms

135 The predators, *Macrocylops albidus*, *M. fuscus* and *Megacyclops viridis*, were
136 collected from Glastry Clay Pit Ponds, Northern Ireland (54°29'18.5"N; 5°28'19.9"W) during
137 January 2017 using a polypropylene dipper, transported to Queen's Marine Laboratory,
138 Portaferry in source water, and maintained at 25 ± 2 °C under a 16:8 light:dark regime.
139 Cultures were initiated separately for each species by placing a single ovigerous adult female
140 into 250 mL dechlorinated tap water, with the protists *Paramecium caudatum* and
141 *Chilomonas paramecium* supplied *ad libitum*. Emerging nauplii were transferred into larger

142 10 L tanks and fed on the same protist diet, while autoclaved wheat seeds were used to
143 sustain the protists.

144 The prey, *Culex quinquefasciatus*, were obtained from a laboratory colony maintained
145 at the same conditions as the predators. *Culex quinquefasciatus* were originally collected in
146 Cyprus in 2005 by Dr A. Callaghan and had been reared in laboratory conditions at the
147 University of Reading since then. Adults were maintained in 32.5 cm³ cages (Bugdorm,
148 Watkins and Doncaster, Leominster, England). Defibrinated horse blood (TCS Biosciences,
149 Buckingham, England) was fed to adults three times per week using an artificial membrane
150 feeding system (Hemotek Ltd., Accrington, England). Cotton pads soaked in 10% sucrose
151 solution were provided for other sustenance. A black oviposition cup containing 200 mL tap
152 water was kept in each cage, with egg rafts removed three times per week and placed into 3 L
153 larval bowls containing tap water and ground guinea pig pellets (Pets at Home,
154 Newtownabbey, Northern Ireland) provided *ad libitum* until mosquito pupation.

155 2.2. Experiment 1: Functional responses (FRs) and Relative Control Potential (RCP)

156 Adult, non-ovigerous female *M. albidus* (1.6 – 1.8 mm body length), *M. fuscus* (1.9 –
157 2.1 mm body length) and *M. viridis* (2.0 – 2.3 mm body length) were selected for
158 experiments to standardise predators as cyclopoids are sexually dimorphic (Laybourn-Parry
159 et al. 1988). Predators were starved for 24 hours to standardise hunger levels. Prey, first instar
160 *C. quinquefasciatus* (1.1- 1.3 mm), were provided to copepods at six densities (2, 4, 8, 15, 30,
161 60, $n = 3$ per density) in 20 mL arenas of 42 mm diameter, and allowed to settle for three
162 hours prior to the addition of individual predators. Predators were allowed to feed for 24
163 hours, after which they were removed and remaining live prey counted. Controls consisted of
164 three replicates at each prey density without a predator. We integrated reproductive effort

165 data derived from Maier (1994) with maximum feeding rates and attack rates to calculate
166 RCP among the three predators (see below).

167 2.3. Experiment 2: Habitat complexity

168 Adult female *M. albidus* (1.6 – 1.8 mm body length) were selected for experiments
169 and starved for 24 hours to standardise hunger levels. Prey, first instar *C. quinquefasciatus*
170 (1.1- 1.3 mm), were provided at the same densities ($n = 3$ per experimental group) and in the
171 same arenas as Experiment 1, and also allowed the same acclimation. However, arenas
172 exhibited either ‘low’ or ‘high’ complexity, with ‘low’ complexity treatment arenas being
173 vacant and ‘high’ complexity arenas containing five polypropylene caps (7 mm diameter, 10
174 mm height), arranged in a uniform array (Figure 1) and positioned using non-toxic mounting
175 putty. As in Experiment 1, predators were allowed to feed for 24 hours before being removed
176 and remaining live prey counted. Controls consisted of three replicates at each prey density
177 and complexity level without predators.

178 2.4. Experiment 3: Ovipositional responses

179 We assessed ovipositional preferences of *C. quinquefasciatus* with pairwise choice
180 tests. Recently emerged female *C. quinquefasciatus* were blood fed for the first time over a
181 24 hour period. Following this, each experimental replicate consisted of cages (32.5 cm³)
182 containing 20 blood-fed females and a 10% sucrose solution, refilled *ad libitum*. Mosquitoes
183 were maintained in these cages for 144 hours in bioassay 1, and 168 hours in following
184 bioassays, at 25 °C ± 2 °C and in a 16:8 light:dark regime. In all bioassays, control and
185 treatment cups were established 48 hours before being added to cages in 80 ml arenas of 6.5
186 cm diameter using water from a continuously aerated source. Adult female *M. albidus* were
187 used to produce all bioassays and were fed with mosquito larvae *ad libitum* prior to use. In
188 bioassay 1, three copepods were physically present in the treatment cups when added to the

189 cages ($n = 7$ per experimental group). In bioassay 2, three copepods were removed from the
 190 treatment cups immediately before being added to cages ($n = 13$ per experimental group). In
 191 bioassay three, ten copepods were ground up using a pestle and mortar and added to
 192 treatment cups ($n = 7$ per experimental group). In bioassays 4 and 5, three copepods were
 193 added to arenas containing black liquid pond dye (0.3 g L^{-1} , Dyofix, Leeds, United Kingdom,
 194 $n = 7$ per experimental group). Controls lacked predators, and those in bioassays 1-3 and 5
 195 consisted of dechlorinated tap water, whilst controls in bioassay 4 consisted of dyed
 196 dechlorinated tap water. Paired cups were added to opposing corners within cages in a
 197 randomised design and mosquitoes given 48 hours to oviposit. Following this, cups were
 198 removed and the numbers of egg rafts counted.

199 2.5. Statistical analyses

200 Statistical analyses were undertaken in R v3.4.2. (R Core Team, 2017). In
 201 Experiments 1 and 2, logistic regression was used to infer FR types with respect to
 202 proportions of prey eaten as functions of prey density. Categorically, a Type II FR is
 203 indicated by a significantly negative first order term. Rogers' random predator equation was
 204 fitted to the data as prey were not replaced as they were consumed (Juliano, 2001):

$$205 \quad N_e = N_0(1 - \exp(a(N_e h - T)))$$

206 (1)

207 where N_e is the number of prey eaten, N_0 is the initial density of prey, a is the attack constant,
 208 h is the handling time and T is the total experimental period. The Lambert W function was
 209 used to aid model fitting (Bolker, 2008). Estimates of attack rate, handling time and
 210 maximum feeding rate ($1/h$) were non-parametrically bootstrapped ($n = 30$) to provide means
 211 and standard errors (SEs). In Experiment 1, we calculated RCP (see Cuthbert et al. 2018)
 212 using maximum feeding and attack rates with reproductive effort data (clutch weight

213 produced per female body weight per day) from Maier (1994) as a proxy for the numerical
 214 response:

$$215 \quad RCP = \left(\frac{FR \text{ agent A}}{FR \text{ agent B}} \right) \times \left(\frac{RE \text{ agent A}}{RE \text{ agent B}} \right)$$

216 (2)

217 where FR represents maximum feeding or attack rates, and RE is reproductive effort. Where
 218 $RCP = 1$, no difference between biocontrol agents is predicted; where $RCP > 1$, agent A has a
 219 greater control potential than agent B, whilst where $RCP < 1$, agent A has a lesser control
 220 potential than agent B (see Dick et al. 2017; Cuthbert et al. 2018). Uncertainty around RCP
 221 values, in the form of confidence intervals (CIs) and p -values, were calculated as per Dick et
 222 al. (2017). RCP ‘biplots’ were used to visually illustrate the results (e.g. see Cuthbert et al.
 223 2018).

224 In Experiment 3, we used Shapiro-Wilk and Bartlett tests to assess normality and
 225 homoscedasticity for each treatment pair, respectively. Where data were normal and
 226 homoscedastic, we employed paired t -tests for analysis of oviposition data. Wilcoxon tests
 227 were used where data did not meet the assumptions of a parametric test. We calculated
 228 oviposition activity index (OAI) values (Kramer and Mulla, 1979), which enable data to be
 229 standardised as proportions for graphical visualisation:

$$230 \quad OAI = \frac{(NT - NC)}{(NT + NC)}$$

231 (3)

232 where the oviposition activity index (OAI) is a function of the number of egg rafts laid in
 233 treated water (NT) relative to controls (NC). The OAI ranges from -1 to 1, with 0 indicating
 234 no preference between options. Positive values indicate that greater preference is

235 demonstrated for the treatment rather than control cups, whilst negative values show
236 preference for controls.

237 **3. Results**

238 *3.1. Experiment 1: Functional responses (FRs) and Relative Control Potential (RCP)*

239 Prey survival in controls exceeded 99% and thus most prey deaths were attributed to
240 cyclopoid copepod predation. Type II FRs were found for each predator (Table 1; Figure 2).
241 *M. albidus* demonstrated the shortest handling times and, inversely, highest maximum
242 feeding rates, followed by *M. viridis*, which exhibited shorter handling times and higher
243 maximum feeding rates than *M. fuscus* (Table 2; Figure 3a, b). Attack rates varied among the
244 predators; *M. fuscus* exhibited greater attack rates than *M. albidus*, which in turn exhibited
245 greater attack rates than *M. viridis* (Table 2; Figure 3c).

246 Relative Control Potential (RCP) calculations and probabilities are illustrated in Table
247 2. When maximum feeding rates are used in the RCP metric (Figure 4a), *M. albidus* exhibits
248 greater efficacy in controlling *C. quinquefasciatus* than both *M. fuscus* (RCP = 2.73) and *M.*
249 *viridis* (RCP = 1.33). *M. viridis* exhibits a greater efficacy compared to *M. fuscus* (RCP =
250 2.13). With attack rates incorporated into the RCP metric (Figure 4b), *M. albidus*
251 demonstrates similar efficacy predictions to *M. fuscus* (RCP = 0.96), and greater efficacy than
252 *M. viridis* (RCP = 1.42). *M. viridis* exhibits a lower efficacy than *M. fuscus* here (RCP =
253 0.71). These calculations are bolstered graphically in Figure 4 using RCP biplots, with *M.*
254 *albidus* showing greatest efficacy overall when maximum feeding rates are integrated, and
255 similarities in efficacies projected between these predators when attack rates are applied.

256 *3.2. Experiment 2: Habitat complexity*

257 Prey survival in controls was 100%, and thus all prey deaths in treatment groups were
258 attributed to predation by cyclopoid copepods. Type II FRs were observed under both levels

259 of habitat complexity (Table 3; Figure 5). Handling times and maximum feeding rates were
260 similar between both levels of habitat complexity (Figure 6a, b). Attack rates were higher in
261 low compared to high complexity treatments (Figure 6c).

262 3.3. Experiment 3: Ovipositional responses

263 In bioassay 1, significantly fewer egg rafts were laid in cups containing copepods in
264 comparison to controls ($W = 45, p = 0.01$). However, in bioassays 2 and 3, no significant
265 differences in oviposition levels were detected between the chemical treatments with
266 predatory cyclopoids removed ($t = 1.41, df = 12, p = 0.18$), or with ground cyclopoids ($W =$
267 $29.5, p = 0.55$). In bioassay 4, significantly fewer egg rafts were oviposited in dyed water
268 containing predatory copepods compared to dyed water alone ($t = 5.35, df = 6, p = 0.002$). In
269 bioassay 5, significantly more egg rafts were laid in cups treated with both dye and copepods
270 compared to undyed, predator-free controls ($t = 5.60, df = 6, p = 0.001$). Figure 7 illustrates
271 OAI scores for each of the five treatments graphically.

272 4. Discussion

273 The development and application of control strategies to combat mosquito-borne
274 disease is of immense public health importance (Benelli and Mehlhorn, 2016), particularly in
275 light of changes to population dynamics being driven by globalisation, urbanisation and
276 climate change (Townroe and Callaghan, 2014; Medlock and Leach, 2015), as well as
277 emergent context-dependencies (e.g. Fischer et al. 2013). Biological control of mosquitoes
278 using predatory agents can be an effective tool to assist or augment control strategies
279 targeting important disease vector species (e.g. Nam et al. 2012). In this context, the coupled
280 use of functional and numerical responses (or their proxies) provides a robust comparative
281 framework to decipher and project the potential efficacy in controlling target organisms by
282 agents (see Dick et al. 2017; Cuthbert et al. 2018). Consumer-resource interactions are,

283 however, subject to a range of context-dependencies which may modulate interaction
284 strengths, such as habitat complexity (Barrios-O'Neill et al. 2015; Alexander et al. 2015),
285 temperature (Wasserman et al. 2016; Cuthbert et al. 2018) and parasitism (Lavery et al.
286 2017a). Such effects may reduce or nullify the potential of agents in the field, and thus are
287 critical to consider during biocontrol agent evaluations. Furthermore, elucidating behavioural
288 responses, such as those relating to ovipositional choice, of target organisms is imperative to
289 derive a holistic view of agent efficacy, particularly in ecological systems where organisms
290 are not confined to a single habitat patch, such as is often the case with container-breeding
291 mosquitoes in urban and peri-urban environments (Silberbush and Blaustein, 2011; Townroe
292 and Callaghan, 2014).

293 Here, we applied functional responses (FRs) to assess the potential predatory impact
294 of three native cyclopoid copepods towards *C. quinquefasciatus*, a container-breeding
295 mosquito and capable vector of diseases such as West Nile virus and lymphatic filariasis
296 (Turell, 2012; Manimegalai and Sukanya, 2014). Further, we examined the impact of habitat
297 complexity on interaction strengths in this predator-prey system and discerned the
298 ovipositional responses of *C. quinquefasciatus* to predatory cues. The focal predators, *M.*
299 *albidus*, *M. fuscus* and *M. viridis* all exhibited potentially population destabilising Type II
300 FRs towards first instar *C. quinquefasciatus*. This characteristic encompasses high levels of
301 killing at low prey densities, limiting conditions for prey refugia (Holling, 1959) and,
302 concurrently, avoids the compensatory effect of reduced competitive interactions in surviving
303 mosquito prey (Juliano, 2007). This destabilising FR may thus reduce the likelihood of size
304 refuge attainment in prey, given that cyclopoid copepods only effectively consume early
305 instar mosquito larvae (Marten and Reid, 2007). *Macrocyclops albidus* exhibited the highest
306 magnitude FR of the predators, characterised by comparatively low handling times and thus
307 high maximum feeding rates. *M. fuscus* demonstrated the greatest attack rates, indicating

308 strong *per capita* efficacies at lower prey densities. *M. albidus* has already shown promise
309 during previous trials for the control of arbovirus vectors (e.g. Marten, 1990a; Veronesi et al.
310 2015), however the predatory potential of the congeneric *M. fuscus* had yet to be examined
311 prior to this study. The use of RCP illustrates that *M. albidus* exhibits the highest efficacy for
312 the control of *C. quinquefasciatus* when maximum feeding rates are integrated, particularly
313 given the comparatively low reproductive effort of *M. fuscus* (Maier, 1994). When attack
314 rates are used in the RCP metric, similarities between *M. albidus* and *M. fuscus* are deduced.
315 However, in a meta-analysis of the impacts of invasive species, the maximum feeding rate
316 combined with numerical response proxies gave 100% association with degree of ecological
317 impact. That is, the ‘offtake rate’ of prey, sometimes known as the ‘total response’, was the
318 best predictor of consumer impact on resources (Dick et al. 2017; Lavery et al. 2017b). Also,
319 the attack rate in experiments where prey are not replaced following consumption may be
320 artificially constrained (Alexander et al. 2012). Thus, we propose that, while attack rates may
321 give insight into predation at low prey densities, it is the maximum feeding rate combined
322 with numerical response proxies that gives best RCP predictions.

323 Habitat complexity was found to modulate interactions between *M. albidus* and *C.*
324 *quinquefasciatus* through a reduction in attack rates where complexity was present. However,
325 the Type II FR exhibited was robust to variations in habitat complexity, and thus population
326 destabilising effects towards disease vectors such as *C. quinquefasciatus* may be sustained
327 under such conditions. Indeed, habitat structure has previously been demonstrated to affect
328 survivability of mosquito prey to predatory copepods (e.g. Rey et al. 2004), however, density-
329 dependent effects (i.e. FRs) had yet to be considered. Our results contrast to other studies
330 whereby variations to benthic habitat structure fundamentally change the nature or magnitude
331 of the observed FR (e.g. Alexander et al. 2012; but see Alexander et al. 2015). Furthermore,
332 handling times and thus maximum feeding rates were largely unaffected by habitat

333 complexity, and hence feeding magnitudes of *M. albidus* are maintained under these contexts.
334 These traits favour the application of *M. albidus* as a biocontrol agent given the dynamic
335 nature of container-style habitats within which *C. quinquefasciatus* colonises (Bohart and
336 Washino, 1978; Meyer and Durso, 1993).

337 Predator avoidance by culicids during oviposition is common (see Vonesh and
338 Blaustein, 2010), yet interspecific variations with respect to coexistence patterns are
339 commonplace (e.g. van Dam and Walton, 2008). Furthermore, interacting environmental
340 factors and density-dependent effects may further modulate oviposition responses (Silberbush
341 and Blaustein, 2011; Silberbush et al. 2014). Pairwise choice tests have been deemed more
342 powerful and advocated over multiple-choice comparisons when examining responses to
343 predatory cues (Silberbush and Blaustein, 2011). Avoidance behaviours have been deemed
344 stronger for mosquito species which oviposit directly onto waterbodies, such as *Culex* spp.
345 (Vonesh and Blaustein, 2010). Our results are indicative of predator avoidance behaviour of
346 *C. quinquefasciatus* towards *M. albidus* when the copepod is physically present. Yet, these
347 effects were dependent upon the nature of the aquatic environment. Generally, this contrasts
348 with limited results which demonstrate positive selection by *A. aegypti* for sites treated with
349 cyclopoid copepods (Torres-Estrada et al. 2001). We found no evidence for avoidance
350 behaviour in the presence of chemical cues of *M. albidus*. Mosquitoes within the *C. pipiens*
351 complex have been shown to prefer black dyed water when ovipositing, likely as a result of
352 greater perceived depth or a higher nutritional load for their progeny (Ortiz Perea and
353 Callaghan, 2017). Here, when both predator and control treatments were dyed, *C.*
354 *quinquefasciatus* continued to avoid *M. albidus* in pairwise tests. However, when the control
355 treatment was undyed and predator-free, *C. quinquefasciatus* demonstrated clear preference
356 for dyed, predator-treated water. These results indicate interactive complexity, whereby the
357 preference towards dyed habitats overrides the avoidance stimulus relating to physical

358 predator presence. Thus, the use of dye in synergy with predator inoculations may foster
359 more effective biocontrol potential in certain environments.

360 In conclusion, the use of FR demonstrates strong predatory impact and potential for
361 the application of cyclopoid copepods in biocontrol strategies, particularly in light of Type II
362 forms conducive to population destabilising effects. We illustrate that this overarching FR
363 form is robust to interspecific variations and habitat complexities, however, variations in
364 underlying FR parameters are apparent among cyclopoid species and habitat complexities.
365 The integration of reproductive effort estimations allows the better resolution of differential
366 RCP between the species, favouring the use of *M. albidus* over other comparators overall.
367 These results corroborate generally with the success of cyclopoid copepods in mosquito
368 control during field trials, whereby large-scale eradication has been achieved (Kay and Nam,
369 2005; Nam et al. 2012), and the particular interest in the use of *M. albidus* in field-based trials
370 which has been demonstrated (e.g. Marten, 1990a; Veronesi et al. 2015). Understanding the
371 non-consumptive effects of predators is central to biocontrol applications and broader
372 concerns relating to predator-prey interactions (Preisser and Bolnick, 2008; Vonesh and
373 Blaustein, 2010). Here, we demonstrate context-dependent avoidance behaviour of *C.*
374 *quinquefasciatus* in response to copepod predators, yet highlight that the synergistic use of
375 dye may bolster the efficacy of biocontrol releases due to favoured oviposition in these
376 conditions. Additional field-based research to elucidate the effects of dye on community
377 structure and predatory impact is thus warranted.

378

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384

385 **References**

386 Alexander, M.E., Dick, J.T.A., O'Connor, N.E., Haddaway, N.R., Farnsworth, K.D.,
387 2012. Functional responses of the intertidal amphipod *Echinogammarus marinus*: effects of
388 prey supply, model selection and habitat complexity. *Mar. Ecol. Prog. Ser.* 468, 191-202.
389 <http://dx.doi.org/10.3354/meps09978>.

390

391 Alexander, M.E., Dick, J.T.A., O'Connor, N.E., 2013. Trait-mediated indirect
392 interactions in a marine intertidal system as quantified by functional responses. *Oikos*
393 122(11), 1521–1531. <http://dx.doi.org/10.1111/j.1600-0706.2013.00472.x>.

394

395 Alexander, M.E., Kaiser, H., Weyl, O.L.F., Dick, J.T.A., 2015. Habitat simplification
396 increases the impact of a freshwater invasive fish. *Environ. Biol. Fish.* 98(2), 477–486.
397 <http://dx.doi.org/10.1007/s10641-014-0278-z>.

398

399 Benelli, G., Mehlhorn, H., 2016. Declining malaria, rising of dengue and Zika virus:
400 insights for mosquito vector control. *Parasitol. Res.* 115(5), 1747-1754.
401 <http://dx.doi.org/10.1007/s00436-016-4971-z>.

402

403 Barrios-O'Neill, D., Dick, J.T.A., Emmerson, M.C., Ricciardi, A., MacIsaac, H.J.,
404 2015. Predator-free space, functional responses and biological invasions. *Funct. Ecol.* 29(3),
405 377–384. <http://dx.doi.org/10.1111/1365-2435.12347>.

406

- 407 Bohart, R.M., Washino, R.K., 1978. Mosquitoes of California. University of
408 California Press, Berkeley.
- 409
- 410 Bolker, B.M., 2008. Ecological Models and Data in R. Princeton University Press,
411 Princeton.
- 412
- 413 Blaustein, L., Chase, J.M., 2007. Interactions between mosquito larvae and species
414 that share the same trophic level. *Annu. Rev. Entomol.* 52, 489–507.
415 <https://doi.org/10.1146/annurev.ento.52.110405.091431>.
- 416
- 417 Calliari, D., Sanz, K., Martínez, M., Cervetto, G., Gómez, M., Basso, C., 2003.
418 Comparison of the predation rate of freshwater cyclopoid copepod species on larvae of the
419 mosquito *Culex pipiens*. *Med. Vet. Entomol.* 17, 339–342.
- 420
- 421 Cunze, S., Koch, L.K., Kochmann, J., Klimpel, S., 2016. *Aedes albopictus* and *Aedes*
422 *japonicus* - two invasive mosquito species with different temperature niches in Europe.
423 *Parasit. Vectors* 9(1), 573. <http://dx.doi.org/10.1186/s13071-016-1853-2>.
- 424
- 425 Cuthbert, R.N., Dick, J.T.A., Callaghan, A., Dickey, J.W.E., 2018. Biological control
426 agent selection under environmental change using functional responses, abundances and
427 fecundities; the Relative Control Potential (RCP) metric. *Biol. Control* 121, 50–57.
428 <https://doi.org/10.1016/j.biocontrol.2018.02.008>.
- 429
- 430 Dick, J.T.A., Alexander, M.E., Jeschke, J.M., Ricciardi, A., MacIsaac, H.J.,
431 Robinson, T.B., Kumschick, S., Weyl, O.L.F., Dunn, A.M., Hatcher, M.J., Paterson, R.A.,

432 Farnsworth, K.D., Richardson, D.M., 2014. Advancing impact prediction and hypothesis
433 testing in invasion ecology using a comparative functional response approach. *Biol. Invasions*
434 16(4), 735–753. <http://dx.doi.org/10.1007/s10530-013-0550-8>.

435
436 Dick, J.T.A., Lavery, C., Lennon, J.J., Barrios-O'Neill, D., Mensink, P.J., Britton,
437 R., Médoc, V., Boets, P., Alexander, M.E., Taylor, N.G., Dunn, A.M., Hatcher, M.J.,
438 Rosewarne, P.J., Crookes, S., MacIsaac, H.J., Xu, M., Ricciardi, A., Wasserman, R.J.,
439 Ellender, B.R., Weyl, O.L.F., Lucy, F.E., Banks, P.B., Dodd, J.A., MacNeil, C., Penk, M.R.,
440 Aldridge, D.C., Caffrey, J.M., 2017. Invader Relative Impact Potential: a new metric to
441 understand and predict the ecological impacts of existing, emerging and future invasive alien
442 species. *J. Appl. Ecol.* 54(4), 1259–1267. <http://dx.doi.org/10.1111/1365-2664.12849>.

443
444 Fischer, S., Zanotti, G., Castro, A., Quiroga, L., Vargas, D.V., 2011. Effect of habitat
445 complexity on the predation of *Buenoa fuscipennis* (Heteroptera: Notonectidae) on mosquito
446 immature stages and alternative prey. *J. Vector Ecol.* 38(2), 215–223.
447 <https://doi.org/10.1111/j.1948-7134.2013.12033.x>.

448
449 Holling, C.S., 1959. Some characteristics of simple types of predation and parasitism.
450 *Can. Entomol.* 91(7), 385–398. <http://dx.doi.org/10.4039/Ent91385-7>.

451
452 Holling, C. S., 1966. The functional response of invertebrate predators to prey
453 density. *Mem. Entomol. Soc. Can.* 48: 1–86. <https://doi.org/10.4039/entm9848fv>.

454
455 Juliano, S.A., 2001. Non-linear curve fitting: predation and functional response
456 curves, in: Scheiner, S.M., Gurevitch, J. (Eds.), *Design and Analysis of Ecological*

457 Experiments. Oxford University Press, Oxford, pp. 178–196.

458 <http://dx.doi.org/10.4319/lo.1995.40.8.1533>.

459

460 Juliano, S.A., 2007. Population dynamics, in: T.G. Floore (Ed.). Biorational Control
461 of Mosquitoes. J. Am. Mosq. Contr. Assoc. 23 (Suppl. 2), pp. 93–109.

462

463 Juliano, S.A., 2010. Coexistence, exclusion, or neutrality? A meta-analysis of
464 competition between *Aedes albopictus* and resident mosquitoes. Israel J. Ecol. Evol. 56(3–4),
465 325–351. <https://doi.org/10.1560/ijee.55.3-4.325>.

466

467 Kay, B.H., Nam, V.S., 2005. New strategy against *Aedes aegypti* in Vietnam. Lancet
468 365(9459), 613–617. [http://dx.doi.org/10.1016/S0140-6736\(05\)17913-6](http://dx.doi.org/10.1016/S0140-6736(05)17913-6).

469

470 Kramer, W.L., Mulla, M.I.R.S., 1979. Oviposition attractants and repellents of
471 mosquitoes: oviposition responses of *Culex* mosquitoes to organic infusions. Environ.
472 Entomol. 8(6), 1111–1117. <https://doi.org/10.1093/ee/8.6.1111>.

473

474 Lavery, C., Brenner, D., McIlwaine, C., Lennon, J.J., Dick, J.T.A., Lucy, F.E.,
475 Christian, K.A., 2017a. Temperature rise and parasitic infection interact to increase the
476 impact of an invasive species. Int. J. Parasitol. 47(5), 291–296.
477 <http://dx.doi.org/10.1016/j.ijpara.2016.12.004>.

478

479 Lavery, C., Green, K.D., Dick, J.T.A., Barrios-O'Neill, D., Mensink, P.J., Médoc, V.,
480 Spataro, T., Caffrey, J.M., Lucy, F.E., Boets, P., Britton, J.R., Pegg, J., Callagher, C.,

481 2017b. Assessing the ecological impacts of invasive species based on their functional
482 responses and abundances. *Biol. Invasions* 19(5), 1653–1665.
483 <http://dx.doi.org/10.1007/s10530-017-1378-4>.

484
485 Laybourn-Parry, J., Abdullahi, B.A., Tinson, S.V., 1988. Temperature-dependent energy
486 partitioning in the benthic copepods *Acanthocyclops viridis* and *Macrocyclus albidus*. *Can.*
487 *J. Zool.* 66(12), 2709–2713. <https://doi.org/10.1139/z88-397>.

488
489 Maier, G., 1994. Patterns of life history among cyclopoid copepods of central Europe.
490 *Freshw. Biol.* 31(1), 77–86. <https://doi.org/10.1111/j.1365-2427.1994.tb00840.x>.

491
492 Manimegalai, K., Sukanya, S., 2014. Biology of the filarial vector, *Culex*
493 *quinquefasciatus* (Diptera:Culicidae). *Int. J. Curr. Microbiol. Appl. Sci.* 3(4), 718–24.

494
495 Marten, G.G., 1989. A survey of cyclopoid copepods for control of *Aedes albopictus*
496 larvae. *Bull. Soc. Vector Ecol.* 14(2), 232–236.

497
498 Marten, G.G., 1990a. Elimination of *Aedes albopictus* from tire piles by introducing
499 *Macrocyclus albidus* (Copepoda, Cyclopidae). *J. Am. Mosq. Control Assoc.* 6(4), 689–693.

500
501 Marten G.G., 1990b. Issues in the development of cyclops for mosquito control.
502 *Arbovirus Res. Aust.* 5, 159–164.

503
504 Marten, G.G., Reid, J.W., 2007. Cyclopoid copepods. *J. Am. Mosq. Control Assoc.*
505 23(2), 65–92. [http://dx.doi.org/10.2987/8756-971X\(2007\)23%5B65:CC%5D2.0.CO;2](http://dx.doi.org/10.2987/8756-971X(2007)23%5B65:CC%5D2.0.CO;2).

506

507 Medlock, J.M., Leach, S.A., 2015. Effect of climate change on vector-borne disease
508 risk in the UK. *Lancet Infect. Dis.* 15(6), 721–730. [http://dx.doi.org/10.1016/S1473-](http://dx.doi.org/10.1016/S1473-3099(15)70091-5)
509 [3099\(15\)70091-5](http://dx.doi.org/10.1016/S1473-3099(15)70091-5).

510

511 Medlock, J.M., Vaux, A.G.C., Cull, B., Schäffner, F., Gillingham, E., Pfluger, V.,
512 Leach, S.A., 2017. Detection of the invasive mosquito species *Aedes albopictus* in southern
513 England. *Lancet Infect. Dis.* 17(2), 140. [http://dx.doi.org/10.1016/S1473-3099\(17\)30024-5](http://dx.doi.org/10.1016/S1473-3099(17)30024-5).

514

515 Meyer, R.P., Durso, S.L. 1993. Identification of the mosquitoes of California. Mosq.
516 Vector Contr. Assoc. Cali., Sacramento, USA.

517

518 Nam, V.S., Yen, N.T., Duc, H.M., Tu, T.C., Thang, V.T., Le, N.H., Le Loan, L.,
519 Huong, V.T.Q., Khanh, L.H.K., Trang, H.T.T., Lam, L.Z.Y., Kutcher, S.C., Aaskov, J.G.,
520 Jeffery, J.A.L., Ryan, P.A., Kay, B.H., 2012. Community-based control of *Aedes aegypti* by
521 using *Mesocyclops* in Southern Vietnam. *Am. J. Trop. Med. Hyg.* 86(5), 850–859.
522 <http://dx.doi.org/10.4269/ajtmh.2012.11-0466>.

523

524 O’Neil, R.J., 1990. Functional response of arthropod predators and its role in the
525 biological control of insect pests in agricultural systems, in: Dunn, P.E., Baker, R.R. (Eds.)
526 New directions in biological control: alternatives for suppressing agricultural pests and
527 diseases. Alan R. Liss, Inc., New York, pp. 83–96.

528

529 Ortiz Perea, N., Callaghan, A., 2017. Pond dyes are *Culex* mosquito oviposition
530 attractants. *PeerJ* 5, e3361. <http://dx.doi.org/10.7717/peerj.3361>.

531

532 Peacor, S.D., Werner, E.E., 2001. The contribution of trait-mediated indirect effects to
533 the net effects of a predator. *Proc. Natl. Acad. Sci. USA* 98, 3904–3908.

534 <https://doi.org/10.1073/pnas.071061998>.

535

536 Preisser, E.L., Bolnick, D.I., 2008. The many faces of fear: comparing the pathways
537 and impacts of nonconsumptive predator effects on prey populations. *PLoS ONE* 3(6), e2465.

538 <https://doi.org/10.1371/journal.pone.0002465>.

539

540 R Core Team, 2017. *R: A language and environment for statistical computing*. R
541 Foundation for Statistical Computing, Vienna.

542

543 Rey, J.R., O’Connell, S., Suárez, S., Menéndez, Z., Lounibos, L.P., Byer, G. 2004.
544 Laboratory and field studies of *Macrocyclops albidus* (Crustacea: Copepoda) for biological
545 control of mosquitoes in artificial containers in a subtropical environment. *J. Vector Ecol.* 29
546 (1), 124–134.

547

548 Samy, A.M., Elaagip, A.H., Kenawy, M.A., Ayres, C.F.J., Townsend Peterson, A.,
549 Soliman, D.A., 2016. Climate change influences on the global potential distribution of the
550 mosquito *Culex quinquefasciatus*, vector of West Nile virus and lymphatic filariasis. *PLOS*
551 *One* 11(10), e0163863. <https://doi.org/10.1371/journal.pone.0163863>.

552

553 Schäffner, F., Medlock, J.M., Van Bortel W., 2013. Public health significance of
554 invasive mosquitoes in Europe. *Clin. Microbiol. Infect.* 19(8), 685–692.
555 <https://doi.org/10.1111/1469-0691.12189>.

556

557 Sih, A., 1986. Antipredator responses and the perception of danger by mosquito
558 larvae. *Ecol.* 67(2), 434. <https://doi.org/10.2307/1938587>.

559

560 Silberbush, A., Blaustein, L., 2011. Mosquito females quantify risk of predation to
561 their progeny when selecting an oviposition site. *Funct. Ecol.* 25(5), 1091–1095.
562 <https://doi.org/10.1111/j.1365-2435.2011.01873.x>.

563

564 Silberbush, A., Tsurim, I., Margalith, Y., Blaustein, L., 2014. Interactive effects of
565 salinity and a predator on mosquito oviposition and larval performance. *Popul. Ecol.* 175(2),
566 565–575. <https://doi.org/10.1007/s00442-014-2930-x>.

567

568 South, J., Dick, J.T.A., McCard, M., Barrios-O'Neill, D., Anton, A., 2017. Predicting
569 predatory impact of juvenile invasive lionfish (*Pterois volitans*) on a crustacean prey using
570 functional response analysis: effects of temperature, habitat complexity and light regimes.
571 *Environ. Biol. Fish.* 100(10), 1155–1165. <http://dx.doi.org/10.1007/s10641-017-0633-y>.

572

573 Townroe, S., Callaghan, A., 2014. British container breeding mosquitoes: The impact
574 of urbanisation and climate change on community composition and phenology. *PLoS ONE*,
575 9(4), e95325. <https://doi.org/10.1371/journal.pone.0095325>.

576

577 Torres-Estrada, J., Rodriguez, M.H., Cruz-Lopez, L., Arredondo- Jimenez, J.I., 2001.
578 Selective oviposition by *Aedes aegypti* (Diptera: Culicidae) in response to *Mesocyclops*
579 *longisetus* (Copepoda: Cyclopoida) under laboratory and field conditions. J. Med. Entomol.
580 38(2), 188–192. <https://doi.org/10.1603/0022-2585-38.2.188>.

581

582 Tranchida, M.C., Micieli, M.V., Maciá, A., García, J.J., 2009. Native Argentinean
583 cyclopoids (Crustacea: Copepoda) as predators of *Aedes aegypti* and *Culex pipiens* (Diptera:
584 Culicidae) mosquitoes. Rev. Biol. Trop. 57(4), 1059–1068.

585

586 Trussell, G.C., Ewanchuk, P.J., Bertness, M.D., Silliman, B.R., 2004. Trophic
587 cascades in rocky shore tide pools: distinguishing lethal and nonlethal effects. Oecologia
588 139(3), 427–432. <https://doi.org/10.1007/s00442-004-1512-8>.

589

590 Trussell, G.C., Ewanchuk, P.J., Matassa, C.M., 2008. Resource identity modifies the
591 influence of predation risk on ecosystem function. Ecol. 89(10), 2798–2807.
592 <https://doi.org/10.1890/08-0250.1>.

593

594 Turell, M.J., 2012. Members of the *Culex pipiens* complex as vectors of viruses. J.
595 Am. Mosq. Control Assoc. 28(4), 123–6. <https://doi.org/10.2987/8756-971x-28.4.123>.

596

597 Van Dam, A.R., Walton, W.E., 2008. The effect of predatory fish exudates on the
598 ovipositional behaviour of three mosquito species: *Culex quinquefasciatus*, *Aedes aegypti* and
599 *Culex tarsalis*. Med. Vet. Entomol. 22(4), 399–404. [https://doi.org/10.1111/j.1365-
600 2915.2008.00764.x](https://doi.org/10.1111/j.1365-2915.2008.00764.x).

601

- 602 Van Driesche, R., Bellows, T.S., 2011. Biological Control. Springer, Berlin.
603 <https://doi.org/10.1007/978-1-4613-1157-7>.
604
- 605 Veronesi, R., Carrieri, M., Maccagnani, B., Maini, S., Bellini, R., 2015. *Macrocyclus*
606 *albidus* (Copepoda: Cyclopidae) for the biocontrol of *Aedes albopictus* and *Culex pipiens* in
607 Italy. J. Am. Mosq. Control Assoc. 31(1), 32–43. <https://doi.org/10.2987/13-6381.1>.
608
- 609 Vonesh, J.R., Blaustein, L., 2010. Implications of predator-induced shifts in mosquito
610 oviposition site selection for vector control: a meta-analysis. Israel J. Ecol. Evol. 56(3–4),
611 263–279. <https://doi.org/10.1560/ijee.56.3-4.263>.
612
- 613 Wachira, S.W., Ndung'u, M., Njagi, P.G., Hassanali, A., 2010. Comparative
614 responses of ovipositing *Anopheles gambiae* and *Culex quinquefasciatus* females to the
615 presence of *Culex* egg rafts and larvae. Med. Vet. Entomol. 24(4), 369–374.
616
- 617 Wasserman, R.J., Alexander, M.E., Weyl, O.L.F., Barrios-O'Neill, D., Froneman,
618 P.W., Dalu, T., 2016. Emergent effects of habitat complexity and temperature on predator-
619 prey interactions. Ecosphere, 7(2), e01239. <http://dx.doi.org/10.1002/ecs2.1239>.
620
- 621 World Health Organisation, 2017. Vector-borne diseases fact sheet.
622 <http://www.who.int/mediacentre/factsheets/fs387/en/> (accessed 01 December 2017).
623
- 624 Yee, D.A., 2008. Tires as habitats for mosquitoes: A review of studies within the
625 eastern United States. J. Med. Entomol. 45(4), 581–593S. [https://doi.org/10.1603/0022-
626 2585\(2008\)45\[581:tahfma\]2.0.co;2](https://doi.org/10.1603/0022-2585(2008)45[581:tahfma]2.0.co;2).

627

628 Yee, D.A., 2016. Thirty years of *Aedes albopictus* (Diptera: Culicidae) in America:
629 An introduction to current perspectives and future challenges. J. Med. Entomol. 53(5), 989-
630 991. <https://doi.org/10.1093/jme/tjw063>.

631

632 Zuharah, W.F., Lester, P.J., 2010. Can adults of the New Zealand mosquito *Culex*
633 *pervigilans* (Bergorth) detect the presence of a key predator in larval habitats? J. Vector Ecol.
634 35, 100–105. <https://doi.org/10.1111/j.1948-7134.2010.00065.x>.

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648 **Tables**

649 Table 1

Predator	First order term	<i>p</i>	<i>h</i>	<i>p</i>	<i>a</i>	<i>p</i>
<i>M. albidus</i>	-0.03	< 0.001	0.03	< 0.001	2.11	< 0.001
<i>M. fuscus</i>	-0.05	< 0.001	0.04	< 0.001	3.85	< 0.001
<i>M. viridis</i>	-0.02	< 0.001	0.04	< 0.001	1.36	< 0.001

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661 Table 2

Agent A, agent B comparison	FR (\pm SD)	Parameter	Reproductive effort	RCP	CI _s (60%), $p_{\text{RIP}} > 1$ (%)
<i>M. albidus</i> , <i>M.</i> <i>fuscus</i>	35.72 (\pm 7.15), 24.06 (\pm 5.41)	1/h	0.35, 0.20	2.73	2.03 – 3.36, 99.94***
<i>M. albidus</i> , <i>M.</i> <i>viridis</i>	35.72 (\pm 7.15), 26.30 (\pm 4.82)	1/h	0.35, 0.37	1.33	1.02 – 1.61, 82.13
<i>M. viridis</i> , <i>M.</i> <i>fuscus</i>	26.30 (\pm 4.82), 24.06 (\pm 5.41)	1/h	0.37, 0.20	2.13	1.60 – 2.60, 99.35**
<i>M. albidus</i> , <i>M.</i> <i>fuscus</i>	2.04 (\pm 0.36), 4.03 (\pm 1.20)	<i>a</i>	0.35, 0.20	0.96	0.68 – 1.21, 39.10
<i>M. albidus</i> , <i>M.</i> <i>viridis</i>	2.04 (\pm 0.36), 1.42 (\pm 0.29)	<i>a</i>	0.35, 0.37	1.42	1.09 – 1.71, 87.82
<i>M. viridis</i> , <i>M.</i> <i>fuscus</i>	1.42 (\pm 0.29), 4.03 (\pm 1.20)	<i>a</i>	0.37, 0.20	0.71	0.49 – 0.90, 12.63

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665 Table 3

Habitat complexity	First order term	p	h	p	a	p
Low	-0.03	< 0.001	0.04	< 0.001	1.68	< 0.001
High	-0.02	< 0.001	0.04	< 0.001	0.90	< 0.001

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680 Captions

681 Table 1. First order terms resulting from logistic regression (proportion of prey killed versus
682 prey density) for each predator species in Experiment 1, alongside functional response
683 parameter estimates (h , handling time; a , attack rate) resulting from Rogers' random predator
684 equation.

685 Table 2. Relative Control Potential (RCP; means) calculations between species using
686 bootstrapped ($n = 30$ per parameter) maximum feeding rates ($1/h$) and attack rates (a) with
687 reproductive effort data (Maier, 1994). Uncertainties are reflected through CIs. Asterisks
688 denote significant levels of certainty that the RCP score is greater than 1 (* > 95%, ** > 99%,
689 *** > 99.9%).

690 Table 3. First order terms resulting from logistic regression (proportion of prey killed versus
691 prey density) for each level of habitat complexity in Experiment 2, alongside functional
692 response parameter estimates (h , handling time; a , attack rate) resulting from Rogers' random
693 predator equation.

694 Figure 1. Illustration of low and high habitat complexity treatment arenas used in Experiment
695 2.

696 Figure 2. Functional responses of *M. albidus* (a), *M. fuscus* (b) and *M. viridis* (c) towards *C.*
697 *quinquefasciatus* prey over a 24 hour experimental period. Means are \pm SE ($n = 3$ per
698 density).

699 Figure 3. Bootstrapped functional response parameters (handling time, a; maximum feeding
700 rate, b; attack rate, c) of *M. albidus*, *M. fuscus* and *M. viridis* towards *C. quinquefasciatus*.
701 Means are \pm SE ($n = 30$ per parameter).

702 Figure 4. RCP biplot integrating mean bootstrapped maximum feeding rate (a) and attack rate
703 (b) and reproductive effort (Maier, 1994) for each predator. Increasing efficacy is projected
704 from bottom left to top right. Means are \pm SE ($n = 30$ per FR parameter).

705 Figure 5. Functional responses of *M. albidus* towards *C. quinquefasciatus* prey under low and
706 high habitat complexity over a 24 hour experimental period. Means are \pm SE ($n = 3$ per
707 density).

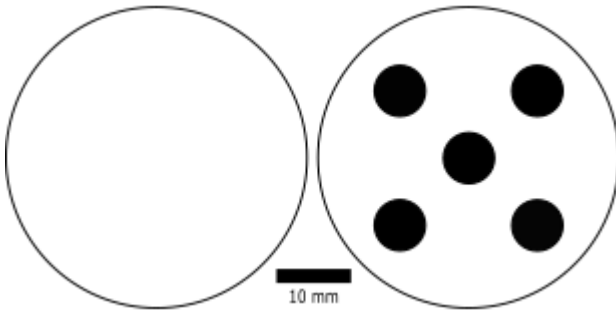
708 Figure 6. Bootstrapped functional response parameters (handling time, a; maximum feeding
709 rate, b; attack rate, c) of *M. albidus* towards *C. quinquefasciatus* under low and high habitat
710 complexity. Means are \pm SE ($n = 30$ per parameter).

711 Figure 7. Oviposition activity index values (OAI, \pm SE) shown by *C. quinquefasciatus*
712 resulting from pairwise tests with predator-free controls and treatments cups inoculated with:
713 (a) three physically-present *M. albidus* ($n = 7$); (b) the cues emitted by three removed *M.*
714 *albidus* ($n = 13$); (c) ten ground *M. albidus* ($n = 7$); (d) three physically-present *M. albidus* in
715 dyed water with a dyed control ($n = 7$); (e) three physically present *M. albidus* in dyed water
716 with an undyed control ($n = 7$). Controls in bioassays (a) – (c) comprised undyed arenas. OAI
717 values of 0 indicate no preference between predator-control treatments, with values closer to -
718 1 indicating an increasing preference for controls (i.e. predator avoidance), and those closer
719 to 1 indicating increasing selection for predator-treated water. Labels above indicate
720 significance levels resulting from statistical tests using raw oviposition data (p : * < 0.05 , ** $<$
721 0.01 , *** < 0.001 , NS ≥ 0.05).

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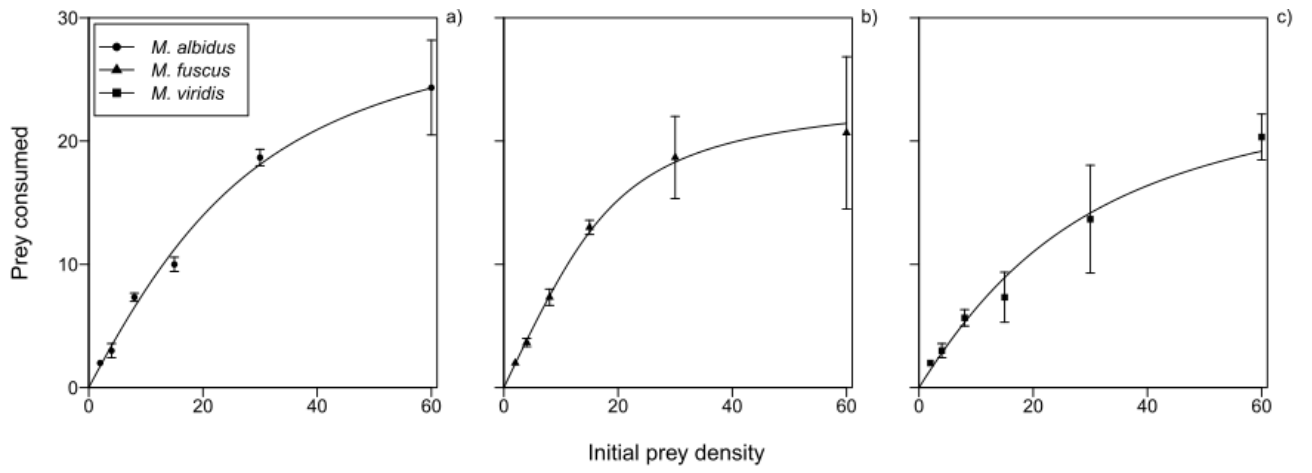
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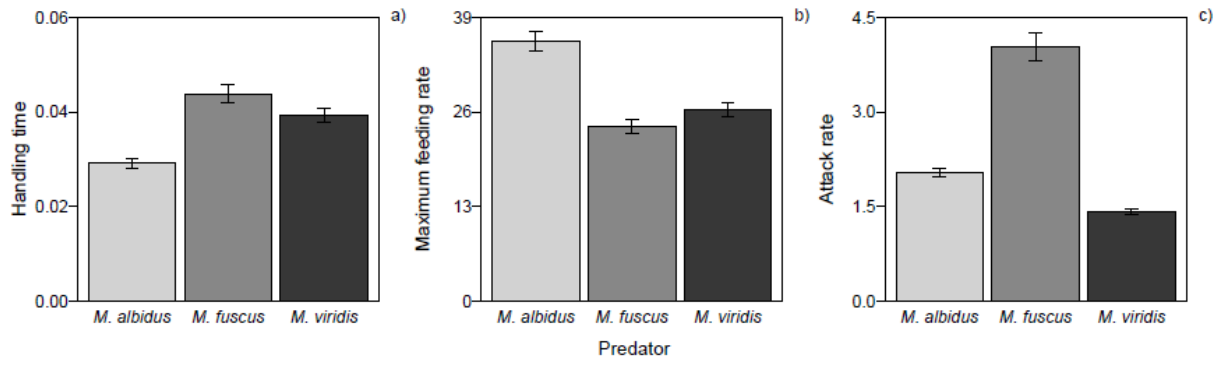
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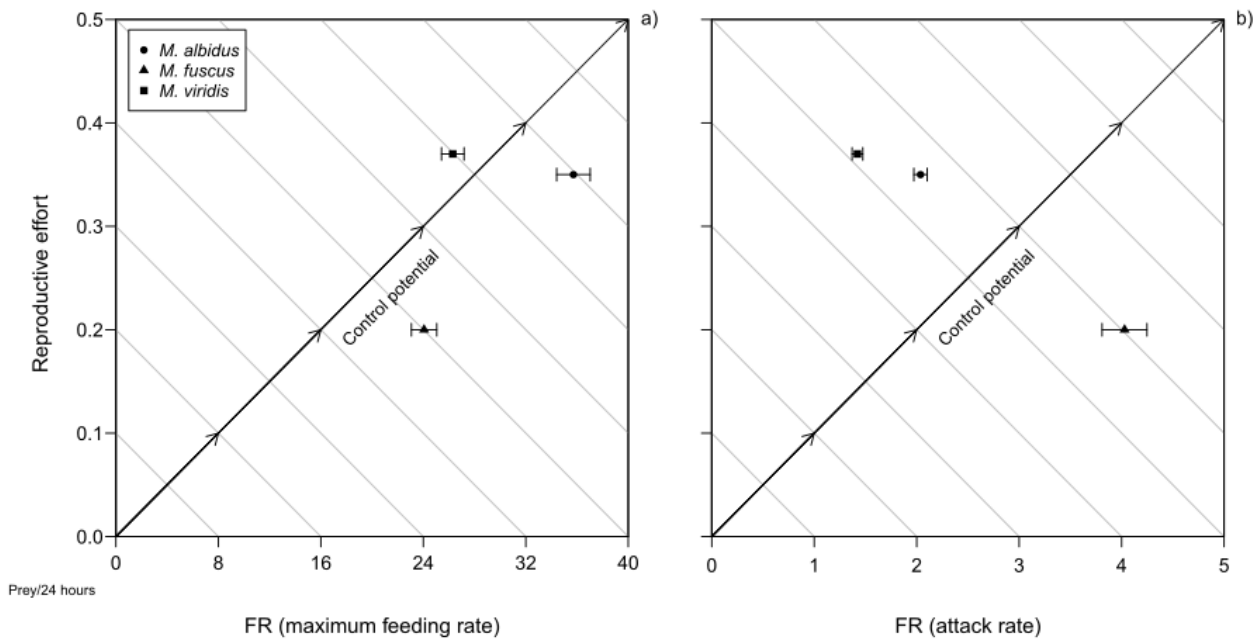
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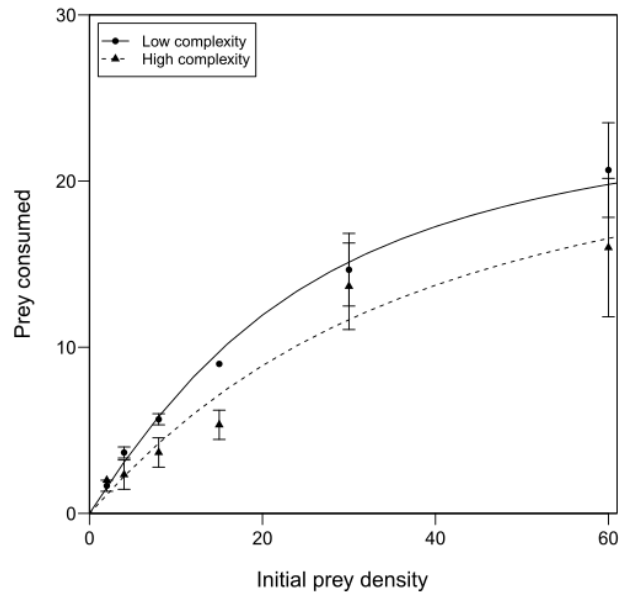
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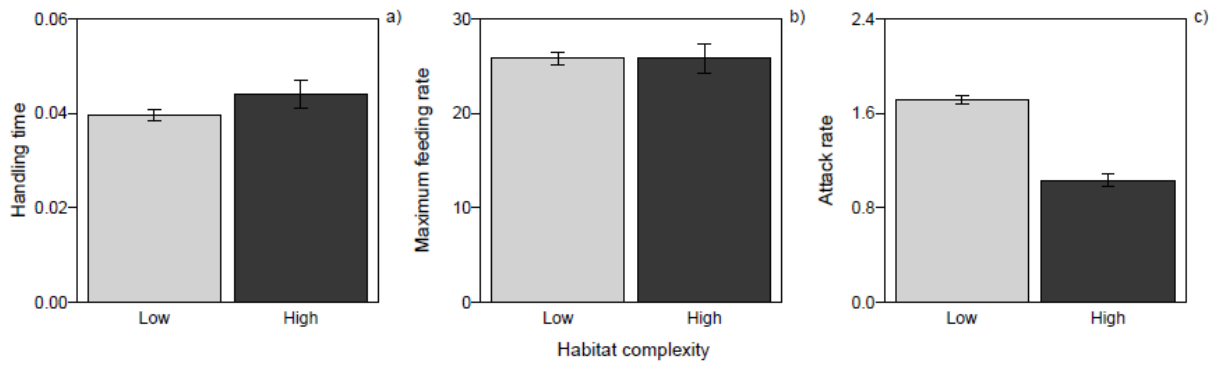
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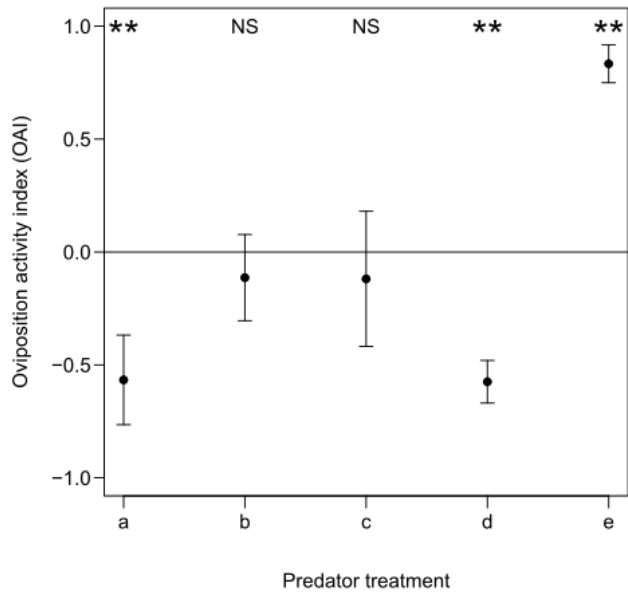
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