

Metal bioaccumulation and cellular fractionation in an epigeic earthworm (Lumbricus rubellus): the interactive influences of population exposure histories, site-specific geochemistry and mitochondrial genotype

Article

**Accepted Version** 

Andre, J., Stürzenbaum, S. R., Kille, P., Morgan, A. J. and Hodson, M. E. (2010) Metal bioaccumulation and cellular fractionation in an epigeic earthworm (Lumbricus rubellus): the interactive influences of population exposure histories, sitespecific geochemistry and mitochondrial genotype. Soil Biology & Biochemistry, 42 (9). pp. 1566-1573. ISSN 0038-0717 doi: 10.1016/j.soilbio.2010.05.029 Available at https://centaur.reading.ac.uk/7686/

It is advisable to refer to the publisher's version if you intend to cite from the work. See <u>Guidance on citing</u>.

To link to this article DOI: http://dx.doi.org/10.1016/j.soilbio.2010.05.029

Publisher: Elsevier



All outputs in CentAUR are protected by Intellectual Property Rights law, including copyright law. Copyright and IPR is retained by the creators or other copyright holders. Terms and conditions for use of this material are defined in the <a href="End User Agreement">End User Agreement</a>.

# www.reading.ac.uk/centaur

# **CentAUR**

Central Archive at the University of Reading

Reading's research outputs online

earthworm (Lumbricus rubellus): the interactive influences of population exposure histories, site-specific geochemistry and mitochondrial genotype. Jane Andre<sup>ab\*</sup>, Stephen R. Stürzenbaum<sup>c</sup>, Peter Kille<sup>a</sup>, A. John Morgan<sup>a</sup> and Mark E. Hodson<sup>b</sup>. <sup>a</sup> Cardiff School of Biosciences, Cardiff University, Park Place, Cardiff, CF10 3US. <sup>b</sup> Department of Soil Science, School of Human and Environmental Sciences, University of Reading, Whiteknights, Reading, RG6 6DW. <sup>c</sup> King's College London, School of Biomedical & Health Sciences, Department of Biochemistry, Pharmaceutical Sciences Research Division, London, SE1 9NH \* Author for correspondence. Current address: Dr. Jane Andre, School of Health and Medicine, Division of Biomedical and Life Sciences, Lancaster University, Lancaster, LA1 4YQ 

Metal bioaccumulation and cellular fractionation in an epigeic

#### <u>Abstract</u>

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

Subcellular fractionation techniques were used to describe temporal changes (at intervals from T<sub>0</sub> to T<sub>70</sub> days) in the Pb, Zn and P partitioning profiles of Lumbricus rubellus populations from one calcareous (M<sub>DH</sub>) and one acidic (M<sub>CS</sub>) geographically isolated Pb/Zn-mine sites and one reference site (C<sub>PF</sub>). M<sub>DH</sub> and M<sub>CS</sub> individuals were laboratory maintained on their native field soils; CPF worms were exposed to both  $M_{DH}$  and  $M_{CS}$  soils. Site-specific differences in metal partitioning were found: notably, the putatively metal-adapted populations,  $M_{DH}$  and  $M_{CS}$ , preferentially partitioned higher proportions of their accumulated tissue metal burdens into insoluble CaPO<sub>4</sub>-rich organelles compared with naive counterparts, C<sub>PF</sub>. Thus, it is plausible that efficient metal immobilization is a phenotypic trait characterising metal tolerant ecotypes. Mitochondrial cytochrome oxidase II (COII) genotyping revealed that the populations indigenous to mine and reference soils belong to distinct genetic lineages, differentiated by ~13%, with 7 haplotypes within the reference site lineage but fewer (3 and 4, respectively) in the lineage common to the two mine sites. Collectively, these observations raise the possibility that site-related genotype differences could influence the toxico-availability of metals and, thus, represent a potential confounding variable in field-based eco-toxicological assessments.

41 42

43

Keywords: earthworms, Pb & Zn, subcellular fractionation, field & lab exposures,

44 genotyping

45

46 47

#### Introduction

49

50

51

52

53

54

55

56

57

58

59

60

61

62

63

64

65

66

67

68 69

70

71

72

73

74

Direct toxic effects arise in metal-exposed organisms not as a consequence of the total accumulated tissue metal burden per se but when the rate of uptake overtakes the combined rates of excretion and detoxification, such that the internal metal-specific concentration threshold of metabolically-available metal is exceeded (Rainbow, 2007; Pan and Wang, 2008). The threshold concentration denoting the transition from no adverse effect to an observable adverse effect for a given metal is referred to as the critical body residue (CBR) (McCarthy and Mackay, 1993; Péry, et al., 2005). Thus, only a fraction of the body burden is toxicologically (re)active or available (Rainbow, 2002; Vijver, et al., 2004). Organisms have evolved mechanisms to regulate the bioreactivities of essential and non-essential metals (Campbell, et al., 2006). In general these initially entail binding and trafficking by chaperone molecules. Essential cations may subsequently be delivered to physiologically labile intracellular storage sites, classically exemplified by Ca<sup>2+</sup>-storing endoplasmic reticulum regions, whilst excess essential and non-essential cations can also either be excreted directly or immobilized as insoluble products in specialized organelles often with long half-lives. These structures possess diverse morphologies and matrix compositions (Hopkin, 1989) that are generically referred to as 'metal-rich granules' or 'concretions' (Campbell, et al., 2006). Improved toxic effects prediction and ecological risk assessment would be likely outcomes of a better knowledge of the fate and speciation of metal within sentinel organisms (Vijver, et al., 2006; Huang, et al., 2009; Jones, et al., 2009). Although there is some evidence from studies on aquatic invertebrates that the toxico-available metals are associated with the cytosolic (soluble) fraction (Perceval, et al., 2006; Péry, et al., 2008), it is generally the case that the relationship between metal induced toxicity and accumulated burden is difficult to evaluate due to the cellular compartmentalization of metals (Campbell, et al., 2006; Vijver, et al., 2006). Techniques such as analytical electron microscopy and synchrotron-based X-ray absorption spectroscopy have been used to some extent to characterize the ligand-binding speciation of metals and metalloids in invertebrate tissues (Cotter-Howells, et al., 2005; Langdon, et al., 2005; Arnold, et al., 2008; Andre, et al., 2009). However, a much more widely used method for segregating invertebrate metal burdens into operationally defined detoxified- and non-detoxified subcellular metal compartments is to differentially centrifuge tissue homogenates. To date, such studies have mainly concentrated on aquatic animals (Honeycutt, et al., 1995; Wallace and Lopez, 1997; Conder, et al., 2002; Wallace, et al., 2003; Cain, et al., 2004; Vijver, et al., 2004), but there is a burgeoning body of publications on the assessment of metal partitioning in earthworms (Arnold et al., 2008; Andre et al., 2009; Huang, et al., 2009; Vijver, et al., 2006; Li, et al., 2008; Jones, et al., 2009).

Voets, et al., (2009) reviewed some of the literature demonstrating that the cellular metal distribution patterns in indigenous invertebrate and vertebrate populations often differ from the distribution patterns observed in naive counterpart organisms exposed to metals in laboratory or field-based transplant experiments. Evidently both exposure history and genetic differentiation are biotic variables that can lead to modifications of the efficiency of metal detoxification by invertebrates (Wallace, et al., 2003) as well as vertebrates (Knapen, et al., 2004). Morgan, et al. (2007) also noted that the genetic background of a population can confound biomarker assays, a further indication that the balance between the sensitive and detoxified metal pools can be altered by microevolutionary events. Given that comprehensive phylogenetic studies on earthworms

using mitochondrial and nuclear markers have recently revealed high intra-species genetic diversity (Velavan, et al., 2007; Novo, et al., 2008) and deeply divergent genetic lineages, possibly in some cases corresponding with cryptic species (King, et al., 2008; Shepeleva, et al., 2008; Pérez-Losada, et al., 2009), it is a major omission that, to the best of our knowledge, no studies hitherto have explicitly attempted to describe the cellular partitioning of metals in field populations of earthworms with respect to exposure history and genotype. A recent report (Langdon, et al., 2009) that populations of the species *Lumbricus rubellus* inhabiting abandoned arsenic mine sites have evolved resistance to the metalloid brings the omission into sharp focus.

The present study had two main aims. First, to investigate the interactive influences of population exposure history and site-specific geochemistry on subcellular metal (Pb, Zn) and P partitioning by comparing two putative adapted *L. rubellus* populations sampled from geochemically contrasting disused Pb/Zn mines (one acidic and one calcareous, respectively) and maintained on their native soils with each other and with reference earthworms transferred experimentally to both polluted soils. Phosphorus partitioning was monitored because phosphate is recognised as the predominant counter-ion in earthworm Pb- and Zn-sequestering cellular compartments (Cotter-Howells, *et al.*, 2005). The second study aim was to use mitochondrial cytochrome oxidase II (COII) to genotype the three field populations. Andre, *et al.* (2010) observed site-specific differences in the tissue and subcellular partitioning profiles of *L. rubellus* populations indigenous to calcareous and acidic sites, respectively. Moreover, the authors reported that the two identified genetically distinct *L. rubellus* lineages were differentially distributed across a heterogeneous polluted landscape, with lineage 'A' predominating within a calcareous Pb/Zn-polluted 'island' and

lineage 'B' predominating in an adjacent acidic polluted location. The present study extended these previous observations through the novel combination of cell fractionation and genotype analyses applied to geographically isolated populations.

128

129

130

131

132

133

134

135

136

137

138

139

140

141

142

143

144

145

146

147

148

125

126

127

## Materials and Methods

#### Soil and earthworm collection and preparation

Soil and earthworms (mature, L. rubellus) were collected from one control site, Pontcanna Fields (CPF) ST 165779 (GPS: 51:29.63122N 3:12.24983W) and two contaminated disused, metalliferous mine sites, Draethen Hollow ( $M_{DH}$ ) ST 217877 (GPS:  $51:34.96185N\ 3:7.88760W$ ) and Cwmystwyth Stream (M<sub>CS</sub>) SN 803748 (GPS: 52:21.48890N 3:45.54702W). At least ten soil samples (excluding the litter layer), taken from a 0-5cm depth, were randomly collected from the sampling areas, combined and mixed. The pH of all soils were measured in deionised H<sub>2</sub>O (Boisson, et al., 1998) prior to them being oven dried at 30°C overnight, sieved to <2mm, then digested in boiling 16N HNO<sub>3</sub> (Morgan and Morgan, 1990) and analysed for major inorganic constituents by inductively coupled plasma - optical emission spectroscopy (ICP-OES; Perkin-Elmer Opitma 3000). Analysis of an in house certified reference material (a sewage sludge amended soil) indicated that that the overall analytical error did not exceed 5.2%. In addition the calibration accuracy of the instrument was assessed through the analysis of an in-house matrix-matched standard and was within 10%. To provide an indication of the organic matter content, loss on ignition (LOI) was determined for each soil sample. 10g (dry weight) of each soil was weighed in a glass crucible and heated to 500°C overnight. The percentage weight reduction was then recorded.

Comment [MEH1]: Have to include this as otherwise the reviewer would expect a reference number to see which commercially available CRM was used. If you have the name or number of the CRM you used (normally something like SSXX I can add in the certified composition.

Comment [MEH2]: This reads correctly

#### Sub-cellular fractionation

150

151

152

153

154

155

156

157

158

159

160

161

162

163

164

165

166

167

168

169

170

171

172

173

174

Thirty boxes containing 300g of contaminated M<sub>CS</sub> soil and 30 boxes containing 300g of contaminated MDH soil were established. The soils were wetted to 70% of their water-holding capacity and reference site (C<sub>PF</sub>) earthworms were placed into 15 boxes (three individuals per box) of M<sub>CS</sub> and 15 boxes of M<sub>DH</sub> soil. Similarly, 3 native M<sub>DH</sub> earthworms were placed into each of the remaining 15 boxes containing M<sub>DH</sub> soil and 3 native M<sub>CS</sub> earthworms into each of the remaining 15 boxes containing M<sub>CS</sub> soil. At 1, 3, 14, 28 and 70 days of soil exposure, 3 boxes of each soil-earthworm combination (i.e. maximum 'n' per 'treatment' = nine) were selected at random and the earthworms depurated prior to freezing. They were depurated for an intial period of 48 hours on moistened filter paper (which was changed daily to prevent coprophagy), followed by 24 hours in a filter-paper free petri-dish (with de-ionised H<sub>2</sub>O) to allow exudation of any filter paper consumed (Arnold and Hodson, 2007). The total exposure period extended to 10 weeks in order to allow sufficient time for the toxicoavailable fraction to stabilize, as was shown to be the case in the lumbricid species Eisenia fetida with no previous history of metal exposure (Jones et al., 2009). When required the earthworms were defrosted, weighed, homogenized in 0.01M Tris-HCl, pH 7.5, and fractionated as described in Arnold, et al., (2008) into a soluble "C fraction" (cytosolic fraction including soluble proteins such as metallothionein and heat shock proteins) and separate insoluble "D" (metal-rich granules) and "E fractions" (tissue fragments, mitochondrial and gut contents) which for the purposes of this study were combined (see supplementary Figure 1). Individual fractions were digested in boiling 16N HNO<sub>3</sub> (Morgan and Morgan, 1990). Samples were made up to volume with ultra-pure water and analysed for major inorganic constituents by ICP-OES with resulting concentrations expressed as mg of metal per kg (wet weight) of earthworm. Blanks were included for each analyses and detection limits were calculated as  $800 \mu g L^{-1}$ ,  $200 \mu g L^{-1}$  and  $400 \mu g L^{-1}$  for P, Pb and Zn respectively. No certified reference materials exist for use with this fractionation method but previous analysis of standard additions were within range (10%), indicating good recovery in the matrix (Arnold et al., 2008) and precision, calculated from repeat analyses of samples was < 5%. Fresh, depurated, weights of the worms sacrificed at each time interval were recorded prior to fractionation; weight change over the exposure periods were taken as estimates of physiological condition.

Comment [MEH3]: If you want to convert these into an equivalent mg per kg wet weight so that they are the same units as the results you report you need to assume a mass of earthworm digested and use this, together with the volume of digestate to calculate them

## Mitochondrial Cytochrome Oxidase II (COII) genotyping

*L. rubellus* earthworms from the M<sub>DH</sub> (n=22), M<sub>CS</sub> (n=32) and C<sub>PF</sub> (n=29) sites, and single *L. castaneus* and *L. eiseni* (from uncontaminated sites in Lancashire, England) were collected by digging and hand-sorting. The animals were transported back to the laboratory in their native soil and depurated (Arnold and Hodson, 2007). A short length (approximately 3cm) of posterior segments was clipped from all *L. rubellus* individuals, and genomic DNA was extracted using DNAzol reagent (Invitrogen Ltd., Paisley, UK). Forward (5'-TAGCTCACTTAGATGCCA) and reverse (5'-GTATGCGGATTTCTAATTGT) *L. rubellus*-specific primers were used to amplify the cytochrome oxidase II (COII) gene, prior to an Exo-SAP-IT PCR clean-up and sequencing using ABI PRISM® BigDye v3.1 Terminator Sequencing technology (Applied Biosystems, USA) as described by Andre, *et al.*, (2010). Raw sequence traces were confirmed using Finch TV before being imported into Mega v3.1 (Kumar, *et al.*, 2004) for alignment and tree construction. The distance-based neighbour joining (NJ) algorithm (Saitou and Nei, 1987), using p-distance, was used to estimate tree topology and calculate branch lengths.

#### Results

#### 202 Soil analysis

Tables 1 and 2 show the concentration of Pb, Zn and P in soil and earthworms sampled from the metalliferous  $M_{CS}$  and  $M_{DH}$  and reference  $C_{PF}$  sites, as well as the percentage body weight change over the full extent of the exposure period. The Pb and Zn soil concentrations were highest at the calcareous  $M_{DH}$  site; acidic  $M_{CS}$  soil was only mildly contaminated, but contained significantly higher Pb and Zn concentrations than the reference  $C_{PF}$  soil. Phosphorus concentration was significantly higher in  $C_{PF}$  reference soil than in the two metalliferous soils.  $C_{PF}$  earthworms maintained higher whole body P concentrations after 70 days of exposure to both metalliferous soils when compared with their  $M_{DH}$  and  $M_{CS}$  counterparts. Mean total earthworm tissue Pb and Zn levels to some extent reflected the corresponding soil Pb and Zn concentrations, although it is noteworthy that the worms indigenous to the acidic  $M_{CS}$  site had a Pb bioaccumulation factor of greater than 1 (based upon dryweight values, data not shown).

# Body mass changes

Mortality was evident across all treatment groups and the mean fresh weights of earthworms, including  $C_{PF}$  worms on their 'own' unpolluted reference soil, decreased considerably over the exposure period. These observations indicate that a degree of stress mediated by dietary restriction and/or metal toxicity was experienced by all earthworms in our experimental regime.

# Sub-cellular fractionation

Lead: Following a ten-week exposure period, the ex-situ partitioning profiles were similar for both indigenous and naive introduced earthworms exposed to the same soil (Figure 1). Significant increases in Pb concentration were seen in the soluble (C) and insoluble (D+E) fractions of all worms exposed to M<sub>DH</sub> soil, and C<sub>PF</sub> individuals exposed to M<sub>CS</sub> soil. M<sub>CS</sub> individuals only demonstrated a slight increase in insoluble Pb. Destroying the physical integrity of the field soils by indiscriminate sampling, drying, sieving, homogenisation, and re-hydrating appears to have released more metal for uptake into earthworm tissues above the corresponding equilibrated field levels. Pb was found to preferentially partition into the non-soluble or detoxified (D+E) fraction in all earthworm/soil combinations (Figure 1B). Plotting the time course partitioning data with the soluble fraction Pb values expressed as a percentage of the whole body Pb concentration values (Figure 2) revealed differences in the efficiencies of incorporating Pb into the detoxified fraction between indigenous worms and naive worms introduced into the metalliferous soils. Specifically, and consistently over the entire exposure period, the proportion of Pb within the sensitive soluble fraction of M<sub>CS</sub> earthworms was proportionately less than that in C<sub>PF</sub> earthworms maintained on M<sub>CS</sub> soil (Figure 2A). A similar efficiency difference was found between M<sub>DH</sub> and C<sub>PF</sub> earthworms, but only after 10-weeks of exposure (Figure 2B); at earlier intervals no difference was apparent in Pb partitioning between these two populations. Naïve earthworms accumulated Pb linearly in all three fractions over the duration of the exposure period. In contrast, after 28 days M<sub>DH</sub> earthworms preferentially partitioned the majority of accumulated metal into the insoluble (detoxified) fraction. As the concentrations of Cu, Ni, and Sr did not change appreciably over the 10 week exposure period (data not shown), this implies that the

225

226

227

228

229

230

231

232

233

234

235

236

237

238

239

240

241

242

243

244

245

246

247

temporal changes in the concentrations of Pb and Zn in the subcellular fractions were not directly linked to the loss of whole-worm weight over this period.

Zinc: The temporal partitioning profiles of Zn resemble those of Pb, with indigenous worms with multi-generational histories of metal exposure ( $M_{DH}$  and  $M_{CS}$ ) and naive worms with no previous field history of exposure ( $C_{PF}$ ) each sequestering Zn primarily in the insoluble (D+E) fraction, and restricting the cytosolic soluble Zn fraction within relatively narrow limits (Figure 3). Zn uptake in both the soluble and insoluble fractions by naive  $C_{PF}$  earthworms exposed to  $M_{DH}$  soil occurred in a linear fashion during the entire exposure period, whereas after 28 days of exposure  $M_{DH}$  earthworms appeared to preferentially partition Zn into the insoluble (detoxified) fraction. The similarities between Pb and Zn partitioning extended to the comparative efficiency of restricting the metals to the detoxified compartment in indigenous versus introduced populations (Figure 4): the proportion of Zn present in the soluble fraction was appreciably lower in earthworms from the heavily polluted  $M_{DH}$  site at all time points compared with that in  $C_{PF}$  worms introduced to the  $M_{DH}$  soil; the proportion of soluble fraction Zn in  $M_{CS}$  worms was appreciably lower than in  $C_{PF}$  worms maintained on  $M_{CS}$  soil at three time points (3, 14, and 70 days).

*Phosphorus*: In both indigenous mine-site populations maintained on their 'own' soils, and in naive worms introduced to the metalliferous soils, a fairly steady redistribution of P from the soluble cytosolic phase to the insoluble compartment occurred during the ten-week exposure period (cf. Figures 5A and 5B). A considerably higher insoluble P concentration was measured in indigenous and

introduced earthworms exposed to  $M_{DH}$  soil compared with the two treatment groups exposed to the significantly less polluted  $M_{CS}$  soil (Figure 5).

#### Cytochrome oxidase II (COII) genotyping

The phylogenetic structure of the study populations was assessed using the mitochondrial cytochrome oxidase II (mtDNA COII) gene sequence data of individuals sampled from the three sites. Good quality COII nucleotide sequences (304bp) were aligned from 85 *L. rubellus* earthworms and from individuals of *L. castaneus* and *L. eiseni* to rule out the possibility of misidentification. Only functional COII sequences, with no stop or nonsense codons in the reading frame, were used. Intra- and inter-site evolutionary relationships were phylogenetically analysed (Figure 6), with the tree constructed using the distance-based neighbour joining (NJ) algorithm based upon p-distance. Only one representative of each site and haplotype are shown and the resulting tree topology was well supported by bootstrap analyses. The sampled *L. rubellus* individuals could be resolved into two distinct genetic lineages (lineage A and B, respectively), with a mean inter-lineage mtDNA sequence divergence of 13%.

 $C_{PF}$  earthworms grouped exclusively within lineage A, and comprised 7 distinct haplotypes, with a between-haplotype diversity of 1 to 4% (Figure 6). In comparison, the  $M_{DH}$  and  $M_{CS}$  populations derived from mine-associated soils belonged predominantly to the lineage B genotype, and comprised 3 ( $M_{DH}$ ) and 4 haplotypes ( $M_{CS}$ ) exhibiting between-haplotype diversity of 1 to 2%, respectively. Only one individual from each mine site had lineage A genotype signatures. The genetic distance between L rubellus and two other L umbricus species (L castaneus and L

*eiseni*) was calculated as 18.1%, thus indicating that the *L. rubellus* field populations were correctly assigned.

300

301

302

303

304

305

306

307

308

309

310

311

312

313

314

315

316

317

318

319

320

298

299

#### Discussion

Abandoned metal mine soils in the UK and elsewhere harbour locally adapted earthworm populations with innate abilities to tolerate phenomenally high internal body loads of certain metals. For example, earthworms evidently thrive in field soils contaminated to degrees exceeding by an order of magnitude the exposure level that severely compromises reproduction in spiked laboratory soils (Spurgeon, et al., 1994). That these are residents and not immigrants from less-contaminated surrounding soil is one way of interpreting the 'patchy' pattern of genotype distributions observed in L. rubellus across geochemically heterogeneous metalliferous landscapes (Andre, et al., 2010). A number of published studies provide mechanistic insights concerning the modes of metal detoxification within discrete subcellular compartments in these chronically exposed natural populations (Morgan and Morris, 1982; Morris and Morgan, 1986; Morgan and Morgan, 1989a; 1998; Sturzenbaum, et al., 2001). However, evidence of phenotypic differences at the behavioural, physiological and molecular levels between populations that have undergone multiple generations of exposure and their counterparts with no comparable metal exposure history in their native habitat remain sparse. Therefore, by comparing the subcellular partitioning profiles amongst earthworm populations native to contaminated and clean sites, further inferences into the metal management strategies of putatively adapted ecotypes may be gained.

A Cd-resistant ecotype of the freshwater oligochaete Lumbriculus hoffmeisteri has been shown to possess enhanced Cd accumulation efficiency (Klerks and Bartholomew, 1991) and a concomitant reduction in the amount of trophicallyavailable Cd (Wallace and Lopez, 1997). Such integrated duality is also expressed for Pb in at least one of the earthworm populations, M<sub>DH</sub>, examined in the present study. Specifically, Pb partitioning profiles for M<sub>DH</sub> individuals showed a much lower absolute level of soluble Pb (approximately 57% after 70 days exposure), when compared to naive  $C_{\text{PF}}$  earthworms exposed to the same  $M_{\text{DH}}$  polluted soil. This population appears to have evolved a capability to limit Pb toxico-availability possibly through modifications of components of Ca<sup>2+</sup> transporting pathway, such as the sarcoplasmic/endoplasmic reticulum calcium ATPase (SERCA) (Andre, et al., 2010). The notion is supported by plotting the soluble Pb fraction of M<sub>DH</sub> worms as a percentage of the total body load; although the total body burden increases over the 10 week exposure period on their native soil, there is a significantly lower proportion of the Pb burden distributed in the soluble fraction compared with that found in C<sub>PF</sub> reference site worms maintained in the laboratory for the same period on M<sub>DH</sub> metalliferous soil. A similar differential was also recorded in the proportional Pb content of the soluble fractions of M<sub>CS</sub> and C<sub>PF</sub> worms at all time intervals of laboratory exposure to M<sub>CS</sub> soil. This phenomenon of increasing metal concentrations in earthworms from metal-contaminated soils maintained on their 'own' soils in the laboratory has been reported by others (e.g. Corp and Morgan, 1991). It is not easily explained in the cases of metal that are not especially redox active other than as a consequence of the destruction of the physical structure of soil, with a release of previously stabilized metal into the 'bioavailable' pool, i.e. a partial reversal of the 'ageing' process. The indication that Pb bioreactivity is reduced in earthworms

322

323

324

325

326

327

328

329

330

331

332

333

334

335

336

337

338

339

340

341

342

343

344

345

indigenous to Pb-contaminated soils was reinforced by the finding that the weight loss experienced by reference  $C_{PF}$  earthworms maintained on the two studied metalliferous soils exceeded that experienced by earthworms native to the field soils. These data highlight the crucial role that intracellular components and machinery play in facilitating the efficient delivery of metals to intracellular compartments where they are sequestered in insoluble states.

Several publications have shown that lysosome-like chloragosomes within the chlorogogenous tissue (possessing some functional similarities to vertebrate hepatocytes) are the main metal-sequestering organelles. Chloragosomes represent phosphate-rich storage compartments for group A, O-seeking, metals (Morgan and Morris, 1982; Morgan and Morgan, 1989a; b; 1998). Andre, *et al.*, (2009) investigated the ligand speciation of Pb within whole earthworms using synchrotron- based XAS analysis, and obtained XANES spectra that unambiguously revealed that *L. rubellus* with a protracted population history of Pb exposure preferentially sequester the metal as insoluble pyromorphite [Pb<sub>5</sub>(PO<sub>4</sub>)<sub>3</sub>Cl] and Pb<sub>3</sub>(PO<sub>4</sub>)<sub>2</sub>. Given this fact, the observation in the present study that intracellular P speciation shifts appreciably in all exposures over the entire 10 week period from a relatively soluble to less soluble state presumably to associate with intruding Pb is functionally logical.

Due to its biological essentiality it is predictable that invertebrates are able to regulate intracellular Zn levels to a considerable degree. Chromatographic observations demonstrate that this may be achieved through Zn binding to a variety of low- and high-molecular weight molecules (Susuki, *et al.*, 1988; Cain and Luoma, 1998; Lock

and Janssen, 2001). Homeostatic systems operate to not only sequester and detoxify excess Zn but, when needed, to release Zn in order to meet the cells physiological requirements. This system is undoubtedly at work in earthworms from  $M_{DH}$  as, despite considerable increases in total body load, they demonstrate the ability to maintain their intracellular soluble Zn content within relatively narrow limits. Again, the phosphate-rich chloragosomes are implicated in Zn storage and detoxification alongside a less well characterised sulphur-rich organelle, the cadmosome (Sturzenbaum, *et al.*, 1998). The involvement of chloragosomes and cadmosomes in excess Zn sequestration has been corroborated by XAS analyses, with XANES spectra indicating that Zn binds to both O- and S-donating ligands (Andre, *et al.*, 2009).

Cryptic or sibling species are typically found in taxa that thrive in complex, heterogeneous, environments and have been discovered by genotyping fauna inhabiting diverse marine, freshwater, and terrestrial habitats (Sturmbauer, *et al.*, 1999; Pinceel, *et al.*, 2004; Mathews, 2006; Pfenninger and Schwenk, 2007). The *L. rubellus* population indigenous to the unpolluted field site, C<sub>PF</sub>, belongs exclusively to lineage A and can be resolved into 7 haplotypes, whilst the two geographically isolated mine-site populations both belong to the genotypically distinct lineage B comprised of 3 and 4, respectively, distinct haplotypes. The number of *L. rubellus* populations examined was too restricted to draw firm conclusions regarding microevolutionary genealogies, but the higher intra-lineage diversity of the C<sub>PF</sub> sample is indicative of a relatively stationary population that has undergone multiple introductions and bottleneck episodes during its evolutionary history (Harpending, 1994). It is tempting to interpret the comparatively narrow genetic diversity within the

lineage B inhabitants of the mine sites as a hallmark of stress-driven genetic erosion processes (natural selection, genetic drift, inbreeding) having acted upon these populations. Genetic erosion can certainly accompany small fragmented populations (Buza et al., 2000) such as those found inhabiting the 'islands of toxicity' that typify abandoned metal mine sites. However, the genetic erosion notion as an explanation of the genetic structure of mine-associated earthworm populations should be tempered with the knowledge that calculations from genetic parameters lead to the conclusion that lineage A (with an inter-stadial expansion time of ~250 000 years BP) is appreciably 'older' than lineage B (expansion time of ~17000 years BP) (Andre, et al., 2010) and may have had the opportunity to evolve more genetic richness. Peles, et al. (2003) suggested that certain alleles and genotypes in L. rubellus may be more sensitive to the effects of heavy metals because the frequency of both differed significantly at polymorphic loci between populations inhabiting sewage contaminated and reference soils. Conversely, Haimi, et al. (2007) reported that metal contamination did not significantly impact upon clonal diversity in the earthworm Dendrobaena octaedra. Analogous inter- or intra-lineage conclusions cannot firmly be drawn from the present study on L. rubellus. Whether or not the two deeply divergent L. rubellus lineages warrant the status of (cryptic) species must await further genetic and breeding evidence. Nevertheless, it is noteworthy that Lentzsch and Golldack (2006) observed that species richness not ambient soil conditions was the overriding factor affecting intraspecific diversity and genotype abundance in the earthworm Aporrectodea caliginosa, thus ostensibly supporting the hypothesis that ecological niches are colonised at a species level prior to local population-level adaption. The shallow soils often associated with abandoned metal mines usually

396

397

398

399

400

401

402

403

404

405

406

407

408

409

410

411

412

413

414

415

416

417

418

harbour impoverished earthworm communities, in many instances no more than two taxonomically accepted representatives of the epigeic ecophysiological group.

422

423

424

425

426

427

428

429

430

431

432

433

434

435

436

437

438

439

440

420

421

In conclusion, the ability of an adapted population to tolerate the prevailing stressevoking conditions in severely polluted habitats most probably involve heritable and integrated combinations of physiological, morphological and behavioural modifications. Thus, it is plausible to hypothesise that a metal tolerant earthworm population has evolved efficient mechanisms of detoxification that feature an enhanced immobilisation capacity coupled to a relative reduction in the metal sensitive fraction as an important component of their holistic adaptive arsenal. However, inferences about population-specific adaptation based on subcellular metal partitioning profiles should be drawn with a measure of caution because of the possibility that they could be attributable to lineage-specific traits that are independent of chronic metal exposure. This is illustrated by the findings of (Heethoff, et al., 2004) that the parthenogenic earthworm Octolasion tyrtaeum is differentiated into two lineages differing significantly in body size. Such findings, together with those arising from the present study, raise the spectre that field-based eco-toxicological assessments that utilise earthworms, particularly those emanating from discriminating 'omics' measurements, might benefit from the elimination of a potential confounding biotic variable through prior genotyping of all individuals to establish that they possess some genetic background equivalence.

441

442

443

444

<u>Acknowledgments</u>. This work was performed during the tenure of a Natural Environment Research Council studentship (JA) registered at Reading University, and a research grant (F0 0407/AI) awarded by The Leverhulme Trust to AJM. We thank

- 445 Dr Trevor Piearce of Lancaster University for providing us with specimens of
- 446 Lumbricus eiseni for genotyping.

- 448 References
- 449 Andre, J., Charnock, J., Stürzenbaum, S.R., Kille, P., Morgan, A.J., Hodson, M.E.,
- 450 2009. Accumulated Metal Speciation in Earthworm Populations with
- 451 Multigenerational Exposure to Metalliferous Soils: Cell Fractionation and High-
- 452 Energy Synchrotron Analyses. Environmental Science and Technology 43, 6822-
- 453 6829.
- 454 Andre, J., King, R.A., Stürzenbaum, S.R., Kille, P., Hodson, M.E., Morgan, A.J.,
- 455 2010. Molecular genetic differentiation in earthworms inhabiting a heterogeneous Pb-
- 456 polluted landscape. Environmental Pollution 158, 883-890.
- 457 Arnold, R.E., Hodson, M.E., 2007. Effect of time and mode of depuration on tissue
- 458 copper concentrations of the earthworms Eisenia andrei, Lumbricus rubellus and
- 459 Lumbricus terrestris. Environmental Pollution 148, 21-30.
- 460 Arnold, R.E., Hodson, M.E., Charnock, J.M., Peijnenburg, W.J.G.M., 2008.
- 461 Comparison of the subcellular partitioning, distribution and internal speciation of Cu
- between Cu-tolerant and naive populations of Dendrodrilus rubidus Savigny.
- Environmental Science and Technology 42, 3900-3905.
- 464 Boisson, J., Mench, M., Sappin-Didier, V., Sold, P., Vangronsveld, J., 1998. Short-
- 465 term in situ immobilization of Cd and Ni by beringite and steel shots application to
- long-term sludged plots. Agronomie 18, 347-359.

- 467 Buza, L., Young, A., Thrall, P. (2000). Genetic erosion, inbreeding and reduced
- 468 fitness in fragmented populations of the endangered tetraploid pea Swainsona recta.
- 469 Biological Conservation 93, 177-186.
- 470 Cain, D.J., Luoma, S.N., 1998. Metal exposures to native populations of the caddisfly
- 471 Hydropsyche (Trichoptera: Hydropsychidae) determined from cytosolic and whole
- body metal concentrations. Hydrobiologia 386, 103-117.
- 473 Cain, D.J., Luoma, S.N., Wallace, W.G., 2004. Linking metal bioaccumulation of
- 474 aquatic insects to their distribution patterns in a mining- impacted river.
- Environmental Toxicology and Chemistry 23, 1463-1473.
- 476 Campbell, P.G.C., Chapman, P.M., Hale, B.A., 2006. Risk assessment of metals in the
- 477 environment. Issues in Environmental Science and Technology 22, 101-131.
- 478 Conder, J.M., Seals, L.D., Lanno, R.P., 2002. Method for determining toxicologically
- relevant cadmium residues in the earthworm *Eisenia fetida*. Chemosphere 49, 1-7.
- 480 Corp, N., Morgan, AJ., 1991. Accumulation of heavy metals from polluted soils by
- the earthworm Lumbricus rubellus: Can laboratory exposure of 'control' worms reduce
- 482 biomonitoring problems? Environmental Pollution 74, 39-52
- Cotter-Howells, J., Charnock, J.M., Winters, C., Kille, P., Fry, J.C., Morgan, A.J.,
- 484 2005. Metal Compartmentation and Speciation in a Soil Sentinel: The Earthworm,
- 485 Dendrodrilus rubidus. Environmental Science and Technology 39, 7731-7740.
- 486 Haimi, J., Knott, K.E., Yliniemi, I., 2007. Does metal contamination affect clonal
- diversity of the parthenogenetic earthworm *Dendrobaena octaedra*? European Journal
- 488 of Soil Biology 43, 268-272.

- 489 Harpending, H.C., 1994. Signature of Ancient Population-Growth in a Low-
- 490 Resolution Mitochondrial-DNA Mismatch Distribution. Human Biology 66, 591-600.
- 491 Heethoff, M., Etzold, K., Scheu, S., 2004. Mitochondrial COII sequences indicate that
- 492 the parthenogenetic earthworm Octolasion tyrtaeum (Savigny 1826) constitutes of
- 493 two lineages differing in body size and genotype. Pedobiologia 48, 9-13.
- 494 Honeycutt, M.E., Roberts, B.L., Roane, D.S., 1995. Cadmium deposition in the
- earthworm *Eisenia fetida*. Ecotoxicology and Environmental Safety 30, 143-150.
- 496 Hopkin, S.P., 1989. Ecophysiology of metals in terrestrial invertebrates. Elsevier
- 497 Applied Science, Barking, UK.
- 498 Huang, R., Wen, B., Pei, Z., Shan, X.Q., Zhang, S., Williams, P.N., 2009.
- 499 Accumulation, Subcellular Distribution and Toxicity of Copper in Earthworm
- 500 (Eisenia fetida) in the Presence of Ciprofloxacin. Environmental Science and
- 501 Technology 43, 3688-3693.
- Jones, R.P., Bednar, A.J., Inouye, L.S., 2009. Subcellular compartmentalization of
- 503 lead in the earthworm, Eisenia fetida: Relationship to survival and reproduction.
- 504 Ecotoxicology and Environmental Safety 72, 1045-1052.
- 505 King, R.A., Tibble, A., Symondson, W.O.C., 2008. Opening a can of worms:
- 506 unprecedented sympatric cryptic diversity within British lumbricid earthworms.
- 507 Molecular Ecology 17, 4684-4698.
- Klerks, P.L., Bartholomew, P.R., 1991. Cadmium accumulation and detoxification in
- 509 a Cd-resistant population of the oligochaete Limnodrilus hoffmeisteri. Aquatic
- 510 Toxicology 19, 97-112

- Knapen, D., Bervoets, L., Verheyen, E., Blust, R., 2004. Resistance to water pollution
- 512 in natural gudgeon (Gobio gobio) populations may be due to genetic adaptation.
- 513 Aquatic Toxicology 67, 155-165.
- Kumar, S., Tamura, K., Nei, M., 2004. MEGA3: Integrated software for Molecular
- 515 Evolutionary Genetics Analysis and sequence alignment. Briefings in Bioinformatics
- 516 5, 150-163.
- 517 Langdon, C.J., Hodson, M.E., Arnold, R.E., Black, S., 2005. Survival, Pb-uptake and
- 518 behaviour of three species of earthworm in Pb treated soils determined using an
- 519 OECD-style toxicity test and a soil avoidance test. Environmental Pollution 138.
- 520 Langdon, C.J., Morgan, A.J., Charnock, J.M., Semple, K.T., Lowe, C.N., 2009. As-
- resistance in laboratory-reared F1, F2 and F3 generation offspring of the earthworm
- 522 Lumbricus rubellus inhabiting an As-contaminated mine soil. Environmental
- 523 Pollution 157, 3114-3119.
- 524 Lentzsch, P., Golldack, J., 2006. Genetic diversity of Aporrectodea caliginosa from
- 525 agricultural sites in Northeast Brandenburg, Germany. Pedobiologia 50, 369-376.
- 526 Li, L.Z., Zhou, D.M., Wang, P., Luo, X.S., 2008. Subcellular distribution of Cd and
- 527 Pb in earthworm *Eisenia fetida* as affected by Ca<sup>2+</sup> ions and Cd–Pb interaction.
- 528 Ecotoxicology and Environmental Safety 71, 632-637.
- 529 Lock, K., Janssen, C.R., 2001. Zinc and cadmium body burdens in terrestrial
- 530 oligochaetes: Use and sigificance in environmental risk assessment. Environmental
- Toxicology and Chemistry 20, 2067-2072.

- 532 Mathews, L.M., 2006. Cryptic biodiversity and phylogeographical patterns in a
- snapping shrimp species complex. Molecular Ecology 15, 4049-4063.
- 534 McCarthy, L.S., Mackay, D., 1993. Enhancing ecotoxicological moedeling and
- assessment. Environmental Science and Technology 27, 1719-1728.
- Morgan, A.J., Kille, P., Sturzenbaum, S.R., 2007. Microevolution and ecotoxicology
- of metals in invertebrates. Environmental Science and Technology 41, 1085-1096.
- 538 Morgan, A.J., Morris, B., 1982. The accumulation and intracellular compartmentation
- of cadmium, lead, zinc and calcium in two earthworm species (Dendrobaena rubida
- and Lumbricus rubellus) living in highly contaminated soil. Histochemistry and Cell
- 541 Biology 75, 269-285.
- Morgan, J.E., Morgan, A.J., 1989a. The effect of lead incorporation on the elemental
- 543 composition of earthworm (Annelida, Oligochaeta) chloragosome granules.
- 544 Histochemistry 92, 237-241.
- Morgan, J.E., Morgan, A.J., 1989b. Zinc sequestration by earthworm (Annelida:
- Oligochaeta) chloragocytes. Histochemistry and Cell Biology 90, 405-411.
- 547 Morgan, J.E., Morgan, A.J., 1990. The distribution of cadmium, copper, lead, zinc
- 548 and calcium in the tissues of the earthworm Lumbricus rubellus sampled from one
- uncontaminated and four polluted soils. Oecologia 84, 559-566.
- 550 Morgan, J.E., Morgan, A.J., 1998. The distribution and intracellular
- 551 compartmentalisation of metals in the endogeic earthworm Aporrectodea caliginosa
- sampled from an unpolluted and a metal contaminated site. Environmental Pollution
- 553 99, 167-175.

- Morris, B., Morgan, A.J., 1986. Calcium-lead interactions in earthworms:
- 555 Observations on Lumbricus terrestris sampled from a calcareous abandoned lead
- 556 mine site. Bulletin of Environmental Contamination and Toxicology 37, 226-233.
- 557 Novo, M., Velavan, T.P., AlmodOvar, A., Schulenburg, H., Diaz-CosIn, D.J.,
- 558 Michiels, N.K., 2008. Microsatellite markers for the drought-resistant earthworm
- 559 Hormogaster elisae. Molecular Ecology Resources 8, 901-903.
- Pan, K., Wang, W.X., 2008. The subcellular fate of cadmium and zinc in the scallop
- 561 Chlamys nobilis during waterborne and dietary metal exposure. Aquatic Toxicology
- 562 90, 253-260.
- Peles, J.D., Towler, W.I., Guttman, S.I., 2003. Population genetic structure of
- 564 earthworms (Lumbricus rubellus) in soils contaminated by heavy metals.
- 565 Ecotoxicology 12, 379-386.
- 566 Perceval, O., Couillard, Y., Pinel-Alloul, B., Campbell, P.G.C., 2006. Linking
- 567 changes in subcellular cadmium distribution to growth and mortality rates in
- transplanted freshwater bivalves (*Pyganodon grandis*). Aquatic Toxicology 79, 87-98.
- 569 Pérez-Losada, M., Ricoy, M., Marshall, J.C., Domínguez, J., 2009. Phylogenetic
- 570 assessment of the earthworm *Aporrectodea caliginosa* species complex (Oligochaeta:
- 571 Lumbricidae) based on mitochondrial and nuclear DNA sequences. Molecular
- 572 Phylogenetics and Evolution 52, 293-302.
- 573 Péry, A.R.R., Béthune, A., Gahou, J., Mons, R., Garric, J., 2005. Body residues: a key
- 574 variable to analyze toxicity tests with Chironomus riparius exposed to copper-spiked
- sediments. Ecotoxicology and Environmental Safety 61, 160-167.

- 576 Péry, A.R.R., Geffard, A., Conrad, A., Mons, R., Garric, J., 2008. Assessing the risk
- 577 of metal mixtures in contaminated sediments on Chironomus riparius based on
- 578 cytosolic accumulation. Ecotoxicology and Environmental Safety 71, 869-873.
- 579 Pfenninger, M., Schwenk, K., 2007. Cryptic animal species are homogeneously
- 580 distributed among taxa and biogeographical regions. BMC Evolutionary Biology 7,
- 581 121.
- Pinceel, J., Jordaens, K., Van Houtte, N., De Winter, A.J., Backeljau, T., 2004.
- 583 Molecular and morphological data reveal cryptic taxonomic diversity in the terrestrial
- 584 slug complex Arion subfuscus/fuscus (Mollusca, Pulmonata, Arionidae) in continental
- north-west Europe. Biological Journal of the Linnean Society 83, 23-38.
- Rainbow, P.S., 2002. Trace metal concentrations in aquatic invertebrates: why and so
- 587 what? Environmental Pollution 120, 497-507.
- 588 Rainbow, P.S., 2007. Trace metal bioaccumulation: Models, metabolic availability
- and toxicity. Environment International 33, 576-582.
- 590 Saitou, N., Nei, M., 1987. The Neighbor-Joining Method a New Method for
- Reconstructing Phylogenetic Trees. Molecular Biology and Evolution 4, 406-425.
- 592 Shepeleva, O.A., Kodolova, O.P., Zhukovskaya, E.A., Striganova, B.R., 2008.
- 593 Genetic diversity of populations of the earthworm Lumbricus rubellus
- 594 (Hoffm.)(Oligochaeta, Lumbricidae). Biology Bulletin 35, 170-177.
- 595 Spurgeon, D.J., Hopkin, S.P., Jones, D.T., 1994. Effects of cadmium, copper, lead and
- zinc on growth, reproduction and survival of the earthworm *Eisenia fetida* (Savigny):

- 597 Assessing the environmental impact of point-source metal contamination in terrestrial
- 598 ecosystems. Environmental Pollution 84, 123-130.
- 599 Sturmbauer, C., Opadiya, G.B., Niederstatter, H., Riedmann, A., Dallinger, R., 1999.
- 600 Mitochondrial DNA reveals cryptic oligochaete species differing in cadmium
- resistance. Molecular Biology and Evolution 16, 967-974.
- 602 Sturzenbaum, S.R., Kille, P., Morgan, A.J., 1998. The identification, cloning and
- characterisation of earthworm metallothionein. FEBS Letters 431, 437-442.
- 604 Sturzenbaum, S.R., Winters, C., Galay, M., Morgan, A.J., Kille, P., 2001. Metal ion
- trafficking in earthworms. Identification of a cadmiun-specific metallothionein. The
- Journal of Biological Chemistry 276, 34013-34018.
- 607 Susuki, K.T., Sunaga, H., Aoki, Y., Hatakeyama, S., Sugaya, Y., Sumi, Y., Suzuki,
- T., 1988. Binding of cadmium and copper in the matfly Baetis Thermicus that inhabit
- 609 a river polluted with heavy metals. Comparative Biochemistry and Physiology C
- 610 Comparative Pharmacology 91C, 487-492.
- 611 Velavan, T.P., Schulenburg, H., Michiels, N., 2007. Development and
- 612 characterization of novel microsatellite markers for the common earthworm
- 613 (Lumbricus terrestris L.). Host–Parasite Interactions 7, 24.
- Vijver, M.G., Van Gestel, C.A.M., Lanno, R.P., Van Straalen, N.M., Peijnenburg,
- W.J.G.M., 2004. Internal metal sequestration and its ecotoxicological relevance: a
- 616 review. Environmental Science and Technology. 38, 4705-4712.
- Vijver, M.G., van Gestel, C.A.M., van Straalen, N.M., Lanno, R.P., Peijnenburg,
- W.J.G.M., 2006. Biological significance of metals partitioned to subcellular fractions

619 within earthworms (Aporrectodea Caliginosa). Environmental Toxicology and 620 Chemistry 25, 807-814. 621 Voets, J., Redeker, E.S., Blust, R., Bervoets, L., 2009. Differences in metal 622 sequestration between zebra mussels from clean and polluted field locations. Aquatic 623 Toxicology 93, 53-60. 624 Wallace, W.G., Lee, B.G., Luoma, S.N., 2003. Subcellular compartmentalistion of Cd 625 and Zn in two bivalves. 1. Significance of metal-sensitive fractions (MSF) and biologically detoxified metal (BDM). Marine Ecology Progress Series 249, 183-197. 626 627 Wallace, W.G., Lopez, G.R., 1997. Bioavailability of biologically sequestered 628 cadmium and the implications of metal detoxification. Marine Ecology Progress Series 1997, 149-157. 629 630 631 Figure Legends 632 Figure 1 633 Time course of Pb partitioning into soluble (A) and non-soluble (B) cellular fractions 634 in Lumbricus rubellus sampled from three populations and maintained on two mine-635 associated metal contaminated soils. Pb concentrations (per unit wet weight of 636 earthworm) are presented as mean ±S.E. (maximum 'n' =9). CPF refers to the 637 reference site at Pontcanna Fields, M<sub>CS</sub> the metalliferous acidic site Cwmystwyth 638 Stream and  $M_{DH}$  the metalliferous calcareous site, Draethen Hollow. 639 640 Figure 2 641 Time course of subcellular Pb distribution in Lumbricus rubellus sampled from three 642 populations and maintained on two mine-associated metal contaminated soils, with

the soluble metal concentration expressed as a percentage of the total body concentration. The Pb levels shown represent fractions extracted from M<sub>CS</sub> and C<sub>PF</sub> earthworms in M<sub>CS</sub> soil (A) and M<sub>DH</sub> and C<sub>PF</sub> earthworms in M<sub>DH</sub> soil (B). [See Fig. 1 for dataset error bars and Fig. 1 legend for site identifiers.] Figure 3 Time course of Zn partitioning into soluble (A) and non-soluble (B) cellular fractions in Lumbricus rubellus sampled from three populations and maintained on two mineassociated metal contaminated soils. Zn concentrations (per unit wet weight of earthworm) are presented as mean ±S.E. (maximum 'n' =9). [See Fig. 1 legend for site identifiers.] Figure 4 Time course of subcellular Zn distribution in Lumbricus rubellus sampled from three populations and maintained on two field mine-associated metal contaminated soils, with the soluble metal concentration expressed as a percentage of the total body concentration. The Zn levels shown represent fractions extracted from  $M_{CS}$  and  $C_{PF}$ earthworms in M<sub>CS</sub> soil (A) and M<sub>DH</sub> and C<sub>PF</sub> earthworms in M<sub>DH</sub> soil (B). [See Fig. 1 for dataset error bars and Fig. 1 legend for site identifiers.] Figure 5 Time course of P partitioning into soluble (A) and non-soluble (B) cellular fractions in Lumbricus rubellus sampled from three populations and maintained on two field-

derived metal contaminated soils. P concentrations (per unit wet weight of

643

644

645

646

647

648

649

650

651

652

653

654

655

656

657

658

659

660

661

662

663

664

665

667 earthworm) are presented as mean ±S.E. (maximum 'n' = 9). [See Fig. 1 legend for 668 site identifiers.] 669 670 Figure 6 671 Phylogenetic tree based on p-distance of the cytochrome oxidase II mitochondrial 672 gene of 87 Lumbricus rubellus individuals from the contaminated sites MDH 673 (triangles) and  $M_{CS}$  (circles) and the reference site  $C_{PF}$  (squares). L. castaneus and L. 674 eiseni individuals (open squares) are included. Two L. rubellus lineages are apparent, 675 termed lineage 'A' (light grey) and 'B' (dark grey), respectively, with a mean inter-676 lineage mtDNA sequence divergence of approximately 13%. Tree topology was well 677 supported by bootstrap analyses. Only one representative of each haplotype at the 678 particular sites and are shown; numbers in parentheses indicate the numbers of 679 individuals of a given haplotype.