

*Improvement in rice seed storage longevity from high-temperature drying is a consistent positive function of harvest moisture content above a critical value*

Article

Accepted Version

Whitehouse, K. J., Hay, F. R. and Ellis, R. H. ORCID:  
<https://orcid.org/0000-0002-3695-6894> (2018) Improvement in rice seed storage longevity from high-temperature drying is a consistent positive function of harvest moisture content above a critical value. *Seed Science Research*, 28 (4). pp. 332-339. ISSN 1475-2735 doi:  
<https://doi.org/10.1017/S0960258518000211> Available at  
<https://centaur.reading.ac.uk/76957/>

It is advisable to refer to the publisher's version if you intend to cite from the work. See [Guidance on citing](#).

To link to this article DOI: <http://dx.doi.org/10.1017/S0960258518000211>

Publisher: Cambridge University Press

All outputs in CentAUR are protected by Intellectual Property Rights law, including copyright law. Copyright and IPR is retained by the creators or other copyright holders. Terms and conditions for use of this material are defined in the [End User Agreement](#).

[www.reading.ac.uk/centaur](http://www.reading.ac.uk/centaur)

**CentAUR**

Central Archive at the University of Reading

Reading's research outputs online

**Improvement in rice seed storage longevity from high-temperature drying is a consistent positive function of harvest moisture content above a critical value**

**K. J. Whitehouse<sup>1,2††</sup>, F. R. Hay<sup>1,†,\*</sup> and R. H. Ellis<sup>2</sup>**

*<sup>1</sup>T.T. Chang Genetic Resources Center, International Rice Research Institute, Los Baños, Philippines, <sup>2</sup>School of Agriculture, Policy and Development, University of Reading, Earley Gate, PO Box 237, Reading, RG6 6AR, UK*

*<sup>†</sup>New address: Department of Agroecology, Aarhus University, Forsøgsvej 1, 4200 Slagelse, Denmark.*

*<sup>††</sup>New address: Australian Grains Genebank, Agriculture Victoria Research, Departments of Economic Development, Jobs, Transport and Resources, Private Bag 260, Horsham, Victoria, 3401, Australia*

*\*For correspondence. E-mail [fiona.hay@agro.au.dk](mailto:fiona.hay@agro.au.dk)*

Running title: High-temperature drying of moist rice seed

Keywords: genebanks; rice; seed drying; seed longevity; seed maturity; seed storage; temperature

**Financial support**

This research received no specific grant from any funding agency, commercial or non-for-profit sectors.

## Abstract

Drying reduces seed moisture content which improves subsequent seed survival periods. Diverse maximum temperatures have been recommended to limit or avoid damage to seeds, but some high-temperature drying regimes may improve subsequent seed quality. Seeds from 20 different accessions of five rice (*Oryza sativa* L.) variety groups (aromatic, Aus, Indica, temperate Japonica, tropical Japonica) were harvested over several seasons at different stages of maturation and either dried throughout at 15°C/15% RH or for different initial periods (continuous or intermittent) in different drying regimes at 45°C before final equilibrium drying at 15°C/15% RH. Subsequent seed longevity in hermetic storage at 45°C with 10.9% moisture content was determined. In no case did initial drying at 45°C provide poorer longevity than drying at 15°C/15% RH throughout. There was a split-line relation, which did not differ amongst investigations, between longevity after initial drying at 45°C relative to that at 15°C/15% RH throughout and harvest moisture content, with a break point at 16.5% (a seed moisture status of about -14 MPa). Below 16.5%, relative longevity did not differ with harvest moisture content with little or no advantage to longevity from drying at 45°C. Above 16.5%, relative longevity showed a positive relation with harvest moisture content, with substantial benefit from drying at 45°C to subsequent longevity of seeds harvested whilst still moist. Hence there are temporal (immediately *ex planta* cf. subsequent air-dry storage) and water status discontinuities (above cf. below 16.5%) in the effect of temperature on subsequent air-dry longevity.

## Introduction

Temperature affects crop seed production in numerous ways (e.g. Porter and Gawith, 1999; Wheeler *et al.*, 2000; Sanchez *et al.*, 2014). It also affects the quality of seeds produced, including post-harvest survival (Ellis *et al.*, 1993; Sanhewe *et al.*, 1996). It is becoming clear that the effect of temperature during the development of seed quality from fertilisation to maturity *in planta* and its effect on seed quality thereafter *ex planta* may well vary over time. There is an overall positive effect of higher mean temperature (within limits) throughout wheat (*Triticum aestivum* L.) seed development and maturation on the development of seed quality (Sanhewe *et al.*, 1996). However, the relation is initially negative during early seed development but then becomes positive during later development and throughout seed maturation *in planta* (Nasezadeh and Ellis, 2017). It then alters again with a negative relation between air-dry seed survival *ex planta* and storage temperature (Ellis and Hong, 2007).

High temperature (32/24°C) throughout seed production is damaging to seed quality in Japonica cultivars of rice (*Oryza sativa* L.) (Ellis *et al.*, 1993; Ellis and Hong, 1994), but not if exposure to higher temperatures is delayed to the seed maturation phase (Ellis, 2011). On the other hand, the development of Japonica rice seed quality is damaged by exposure to either cool (18/14°C) or hot temperature (38/34°C) during histodifferentiation and early seed filling (Martínez-Eixarch and Ellis, 2015). After harvest, the relationship between rice air-dry seed survival and storage temperature is negative and quantitatively similar to that in wheat (Ellis and Hong, 2007).

Post-harvest seed drying takes place following late seed maturation and before subsequent storage. Advice on suitable temperatures for the operation of heated-air seed dryers imply a negative relation between seed survival and temperature above a “safe” threshold value, with cooler temperatures recommended the greater the initial seed moisture content in crop seeds (Nellist, 1980), including rice (Lewis, 1950). Safe drying temperatures

were calibrated for different designs of heated-air seed dryers using standard procedures with mature, dry seeds rewetted to different “initial” higher moisture contents (Nellist, 1980).

Genebanks are recommended to dry seeds before storage to moisture contents lower than commercial seed practice and, currently, to use low humidity drying environments combined with cool temperatures rather than heated-air seed dryers. These recommendations are neither species-specific nor dependent upon initial seed moisture content (FAO, 2014). This advice to genebanks has altered since the International Board for Plant Genetic Resources (now Bioversity International) was created in 1974. No singular recommendation was initially made for seed drying procedures in genebanks: two-stage procedures were suggested with advice to avoid using heated-air seed dryers above 40°C “until moisture content falls below 11% in which case 60°C is safe for most species” as the first stage, with 15% RH as the second (IBPGR, 1976). The latter report cautioned that genebanks should investigate to confirm whether their drying procedures are reliable and safe. Subsequent advice evolved from the “suggestion” of thin-layer drying at about 15°C with 10-15% RH with good air recirculation (Cromarty *et al.*, 1982), to “acceptable” warm-air drying of certain crops, but “preferred” conditions of 15°C with 10-15% RH (IBPGR, 1985). Then, there were “recommendations” of 10-25°C with 10-15% RH (FAO/IPGRI, 1994); and, more recently, the cooler more moist range of 5-20°C with 10-25% RH (FAO, 2014).

The International Rice Research Institute’s (IRRI) genebank (T.T. Chang Genetic Resources Center [GRC]) opened in the 1960s and now conserves > 127,000 accessions of cultivated rice and wild relatives. Initially, rice seeds were dried at high temperatures of 45-50°C. That environment was compatible with recommended safe drying temperatures for rice of 48.9°C at 20% moisture content or above, 51.7°C at 15–20%, and 60°C at <15% (Lewis, 1950). However, in accordance with the publication of the preferred or recommended

conditions for genebanks (IBPGR, 1985; FAO/IPGRI, 1994), a drying room (DR) was built in the early 1990s to dry the seeds at  $15\pm 2^{\circ}\text{C}$  with  $15\pm 5\%$  RH.

Critical re-examination of procedures at IRRI's GRC, however, has now provided evidence that in many cases initial high-temperature ( $45^{\circ}\text{C}$ ) seed drying immediately after harvest, before further drying at  $15^{\circ}\text{C}$  with 15% RH in the drying room, improves seed quality (Crisostomo *et al.*, 2011) and seed storage longevity considerably (Whitehouse *et al.*, 2015, 2017). The greater longevity from initial high-temperature drying was associated with harvest moisture contents above a critical value close to 16% (Whitehouse *et al.*, 2015, 2017). This benefit to air-dry seed survival may result from protection mechanisms being stimulated by high-temperature stress in high moisture content, and so still metabolically active, seeds (Whitehouse *et al.*, 2017).

We test here the hypotheses that: (i) there is a critical harvest moisture content below which there is little or no benefit to subsequent rice seed longevity from high-temperature drying but above which the benefit is a function of harvest moisture content; and (ii) that this broken-stick relation does not differ amongst genetically-diverse accessions of rice from different variety groups. Data from Whitehouse *et al.* (2015, 2017) is included within the research reported here to provide a wide test of the second hypothesis.

## **Materials and methods**

### *Plant material*

Twenty *O. sativa* accessions (IRGC 117264 - 117283) from five variety groups (McNally *et al.*, 2009) were selected for different experiments carried out between 2013 and 2015 (Table 1). The accession(s) planted in the dry season (DS) and/or wet season (WS) each year differed amongst experiments, but the standard rice growing protocol at IRRI including production practices and plant protection measures (Reaño *et al.*, 2008) was followed in each

case. Prior to sowing, seeds were sampled from the active collection of the GRC and held at 50°C for 5 d to break dormancy. With the exception of the 2015 Batch dryer (BD) experiment, in which seedlings were transplanted in a screen house (Table 1), all seedlings were raised in a seed bed and then transplanted in the field at IRRI's Ziegler Experiment Station (14° 9' N, 121° 16' W). Seeds were harvested by hand between March and May in the DS, or October and November in the WS. The number of seed lots harvested per accession and their stage of maturity at harvest differed between experiments.

Immediately after harvest, seed lots were threshed and blown to remove debris and half-filled seeds, then cleaned by hand to remove empty, damaged and/or diseased seeds. Seed moisture content (% , fresh weight basis) at harvest was determined using three 5g samples and the high-constant-temperature-oven method (ISTA, 2013).

### *Seed drying*

Seeds from each lot were divided into samples of equal weight (total number varying with number of different drying treatments, Table 1) and placed into 0.2 × 0.33m (length × width) nylon mesh bags (1 mm-diameter holes). One sample from each seed lot was immediately placed in the drying room (DR; 15°C/15% RH) for 14 days. The DR environment complies with that recommended for genebanks (FAO, 2014) and so provided a control treatment, the “baseline” against which the effects of different drying treatments were compared. The remaining seeds within each lot were dried for an initial period in the experimental drying regimes at 45°C provided by different equipment (Table 1), as follows.

1. A locally-fabricated batch dryer (BD) that dried seeds in an open environment with a kerosene gas burner providing hot air at about 45°C on average. The RH could not be controlled or reduced below about 35% (estimate based on prevailing ambient conditions and air-water relations) because there was no additional dehumidification.



2. Saturated salt solutions of  $MgCl_2$  provided approximately 30% RH with seeds placed above the solution in a hermetically-sealed electrical enclosure box (ENSTO Finland Oy, Finland). There was no airflow with passive drying (relying on a limited seed bulk and exposure of all seeds to the atmosphere) at 45°C.
3. A hermetically-sealed climate chamber (VC3 0034-M, Vötsch Industrietechnik, Germany) with in-built dehumidifier, heater, and cooler which provides a controlled, stable environment at the programmed temperature (45°C) and humidity.

After the initial high-temperature drying treatment (Table 1), all seeds were further dried towards equilibrium in the DR. Once seed samples had equilibrated in the DR (which required a total drying period of 14 days), they were sealed inside  $0.17 \times 0.12$  m (length  $\times$  width) laminated-aluminium-foil packets (Moore and Buckle, Saint Helens, UK) and stored at 2-4°C prior to experimental storage.

#### *Seed storage*

Seed samples from each experiment were removed from storage and left at room temperature for approximately 1h to equilibrate before opening. They were divided into 5g subsamples and rehydrated to 10.9% moisture content in 30 mm-diameter open Petri dishes either over a non-saturated LiCl solution (60% RH), in a hermetically-sealed box, for 7 days at room temperature; or, in the climate chamber set at 60% RH and 21.5°C for 4-5 days. The moisture content of each sample was checked (using three subsamples) before estimating the initial ability to germinate (using one subsample). The remaining subsamples were sealed within individual  $0.12 \times 0.09$ m (length  $\times$  width) laminated-aluminium-foil packets and stored hermetically in an incubator at 45°C (experimental storage). One packet from each seed sample was removed after different periods of storage at 1-3d intervals and tested for ability to germinate.

### *Seed germination*

The seeds from each packet were divided between four 90mm diameter Petri dishes with two layers of Whatman No. 1 filter paper wetted with 7.5ml distilled water (30 seeds in each dish). Seeds were incubated at 30°C (12h light/ 12h dark) and then assessed at regular intervals. Seeds were scored as germinated once the radicle had emerged by at least 2mm. After 14d in test, non-germinated fresh seeds were dehulled and tested for an additional 7d before final scoring.

### *Statistical analysis*

The improved seed viability equation

$$v = K_i - p/10^{K_E - C_W \log_{10} m - C_H t - C_Q t^2}$$

(1)

where  $v$  is probit percentage viability (NED) of a seed lot after  $p$  days in hermetic storage at constant temperature  $t$  (°C) and moisture content  $m$  (% , fresh weight basis),  $K_E$ ,  $C_W$ ,  $C_H$  and  $C_Q$  are species-dependent constants, and  $K_i$  is the seed lot constant (Ellis and Roberts, 1980) provided the basis of our approach to the analyses. Probit analysis was carried out using GenStat for Windows, Versions 15-17 (VSN International Ltd, Hemel Hempstead, UK) in accordance with the equation

$$v = K_i - p/\sigma$$

(2)

derived from Eqn (1) for a single storage environment where  $\sigma$  is the standard deviation of the frequency distribution of seed deaths in time (days) (Ellis and Roberts, 1980) or, for seed

lots showing loss in dormancy during early storage, the combined loss in dormancy and loss in viability model

$$g = (K_d + \beta_1 p) \times (K_i - (p/\sigma))$$

(3)

where  $g$  is ability to germinate (NED),  $p$ ,  $K_i$  and  $\sigma$  are as in Eqn (2),  $K_d$  is the initial proportion of non-dormant seeds (NED) and  $\beta_1$  is the probit rate of loss in dormancy (Kebreab and Murdoch, 1999). For those seed lots showing reduced initial viability when fitting Eqn (2), the ‘control mortality’ parameter was included to estimate the proportion of ‘non-responding’ seeds (dead or empty) within the population (Mead and Gray, 1999; Hay *et al.*, 2014). For each experiment, probit analysis was carried out for all seed lots within an accession simultaneously, first fitting full models (all parameters fitted to seed lots independently) and then reduced models (one or more parameters constrained to the same value for different seed lots within an accession). An approximate F-test was used to select the best model.

The period in days to 50% viability ( $p_{50}$ , the product of  $K_i$  and  $\sigma$ ) is the most accurately estimated period of longevity and it was used as the measure of longevity to compare the effects of different drying treatments on seed storage longevity. For each seed lot within each experiment, the difference in longevity between drying in the alternative drying regime (AR  $p_{50}$ ) and the control regime (DR  $p_{50}$ ) was calculated as a proportion of DR  $p_{50}$  as follows

$$\text{Relative improvement in longevity (\%)} = ((\text{AR } p_{50} - \text{DR } p_{50}) / \text{DR } p_{50}) \times 100$$

(4)

In all comparisons, the estimate of  $p_{50}$  in alternative drying regimes was never less than that in the control regime (DR  $p_{50}$ ); and so all values derived from Eqn (4) were zero or (more often) positive.

## Results

Seed lots varied considerably in the relative benefit from initial drying at high temperature to subsequent seed longevity (Fig. 1). The majority of initial treatments at 45°C provided greater longevity compared with drying in the DR throughout. Beneficial treatments were detected in each variety type and in every experiment.

The greatest benefit was more than a three-fold increase in longevity. This was detected in a temperate Japonica cultivar harvested at 28.9% moisture content and initially dried intermittently (8 h d<sup>-1</sup>) for 1-6 days (i.e. all six treatment durations provided no difference in longevity,  $P > 0.05$ ) in the BD until transfer to the DR. The lowest harvest moisture content at which benefit from initial drying at 45°C was detected was 13.0%, the driest sample at harvest in these studies (but an outlier – see later).

A minority of treatments at 45°C provided similar longevity to treatment in the DR throughout. These were largely confined to seeds harvested at moisture contents below 16.5% (Fig. 1). The greatest harvest moisture content at which no benefit from initial drying at 45°C was detected was 17.7%.

The magnitude of the improvement to subsequent seed longevity from drying at 45°C was associated positively with harvest moisture content (Fig. 1). The split-line regression shown in Figure 1 accounted for 81.5% of the variance in relative longevity (once an outlier at 13.0% moisture content was omitted from analysis). Moreover, the common split-line

regression did not differ amongst experiments: constraining all the observations to this relation did not increase residual deviance significantly compared with fitting split-line regressions to each experiment separately ( $F_{(5,75)} = 0.96$ ;  $P = 0.45$ ). Seed longevity relative to the DR control was increased by initial drying at 45°C the greater the harvest moisture content exceeded a critical value of 16.5%. Seeds harvested at moisture contents below 16.5% showed limited or no improvement in relative longevity from initial high-temperature drying compared with seeds dried throughout in the cooler environment of the DR.

The separate estimates of  $\sigma$  (Eqn 2) for storage at 45°C with 10.9% moisture content varied considerably amongst and within both accessions and experiments (Fig. 2). For example, estimates of  $\sigma$  for Indica accessions dried solely in the DR ranged between 6.3 and 12.5 days and between 4 and 7.7 days in the 2013 BD and chamber 1 experiments, respectively. Seeds of the aromatic variety showed a  $\sigma$  value of 6.7 days in the 2013 BD experiment and ranged between 8.3 and 14.3 days in the chamber 1 experiment. This variation in  $\sigma$  was slightly greater after initial drying at 45°C (Fig. 2b) than for the DR alone (Fig. 2a), whilst the 2015 BD investigation with one aromatic accession provided the widest range of estimates of  $\sigma$  (5.3-20 in DR only, compared with 11.1 - 25 days when initially dried at 45°C) and the greatest mean value. The observed values, most notably when seeds were dried in the DR, were not consistent with independent estimates, with the observed values being considerably lower (Fig. 2).

## **Discussion**

This analysis of results from several separate investigations builds upon recent research (Whitehouse *et al.*, 2015, 2017) to confirm that the subsequent longevity of rice seeds harvested while still moist is improved by initial, high-temperature drying (Fig. 1). The

critical seed moisture content at harvest of 16.5% below which little or no benefit from high-temperature drying was detected (Fig. 1) was similar to the previous estimate of 16.2% (Whitehouse *et al.*, 2015). Moreover, there was not one case of damage from drying moist seeds at 45°C (Fig. 1), although damage was detected previously in some treatment combinations at 60°C (Whitehouse *et al.*, 2017). It is also worth remembering that seeds losing moisture at 45°C, or at 60°C, will be cooler than the air temperature due to evaporative cooling (Nellist, 1980). There was an outlying observation at 13.0% moisture content (Fig. 1) where relative longevity was 65% greater than the DR control and so well above the fitted line. The cause is not known. Nevertheless, this is a “right-side error” which does not contradict the conclusion that initial, high-temperature drying of rice seeds harvested at a high moisture content benefits longevity.

The sequence of contrasting responses of rice seed quality post-harvest to temperature (approximately 15 to 45°C) from anthesis to post-harvest air-dry storage is therefore: negative early in seed development during histo-differentiation and early seed filling (Martínez-Eixarch and Ellis, 2015); then positive for brief durations late in the maturation phase whilst the seeds approach maturity but remain moist (16.5 – 31.0% moisture content at harvest, Fig. 1); then neutral for brief periods as seed moisture content transitions to the air-dry range (13.4 – 16.5%, Fig. 1); and then, finally, negative in subsequent air-dry storage (Ellis and Hong, 2007).

There is a similarity between the critical harvest moisture content of 16.5% (Fig. 1) and discontinuities in the post-harvest relations between seed survival and moisture content. The latter occurs at a water potential of around -14 MPa above which, provided oxygen is available, there is a positive relation between seed longevity and moisture content in contrast to the negative relation in conditions drier than -14 MPa (Roberts and Ellis, 1989). In rice, the critical value of 16.5% moisture content (Fig. 1) is close to 90% equilibrium relative

humidity at 25°C (Breese, 1955; Whitehouse et al., 2015). This provides a water potential close to -14 MPa (Roberts and Ellis, 1989).

The improving quality of immature seeds may result from continued development *ex planta* mimicking maturation drying *in planta* (Hong et al., 2000), because such seeds remain metabolically active (Angelovici et al., 2010). The main driver of the improvement in quality detected here, however, may be a stress response caused by high-temperature exposure during this brief phase immediately *ex planta*, allowing enhanced metabolism of protectants and other metabolic pathways which aid the stabilisation of the seed during desiccation and survival in air-dry storage (Whitehouse et al., 2017). For example, the accumulation of sugars and heat-stable proteins during development is associated with desiccation tolerance and potential longevity, but the latter are more likely to account for differences in longevity between seed lots as they accumulate comparatively late in seed development (Sinniah et al., 1998). Moreover, and in accord with the critical value estimated here, the results of proteomic studies suggest two stages of desiccation (Chatelain et al., 2012): one with increasing seed longevity (equivalent to >16.5% moisture content in the current study); and a later one of no improvement because seeds are no longer metabolically active ( $\leq 16.5\%$  moisture content).

Our results clearly show that two-stage drying of moist rice seeds at a high temperature (45°C, and sometimes warmer) before the cool (5-20°C) and dry (10-25% RH) conditions improves subsequent longevity greatly compared with immediate cool and dry conditions. As no high-temperature drying treatment reduced subsequent seed longevity compared with drying in the DR, and to avoid measuring the harvest MC of all seed lots, FAO might consider updating their advice so that all rice seeds intended for genebank storage are dried using this two-stage procedure. Rice grown in paddy conditions in the tropics is under conditions where ambient RH is rarely less than 80% and plant senescence and seed

desiccation is delayed (if the paddy field is not drained), such that the maturing seeds may remain moist. Whilst many crop seeds mature under much drier conditions, we nonetheless suggest that they should be investigated for improvement in seed longevity if moist seeds are initially dried at high-temperature.

The seed viability equation assumes that  $\sigma$  is the same for all seed lots of a species in a constant, defined storage environment, with only  $K_i$  varying in value amongst lots (Ellis and Roberts, 1980). Considerable variation in  $\sigma$  in one environment (45°C with 10.9% moisture content) was found in these experiments (Fig. 2), however, with most observations shorter than the independent estimate provided by the seed viability constants for rice (Ellis and Hong, 2007). Estimates of  $\sigma$  for Indica and temperate and tropical Japonica cultivars from the Seed Information Database (SID; Royal Botanic Gardens Kew, 2018) in this regime were closer to the experimental observations, but with temperate Japonica predicted to lose viability sooner than Indica cultivars (Fig. 2). Tejakhod and Ellis (2018) found that the seed viability constants from Ellis and Hong (2007) overestimated longevity in a Japonica cultivar, but provided reasonable agreement for Indica cultivars. Chang (1991) and Ellis *et al.* (1992) also concluded that seeds of Japonica cultivars are the shorter lived. In contrast, in the current studies with less mature seeds, the variation within a variety group was greater than that between groups and the Indica cultivars tended to be the shorter-lived (Fig. 2). The viability equations provide estimates of seed longevity in air-dry storage, but they are just estimates and each with their own significant error (see e.g. Hay *et al.*, 2003). Once an indication of the seed longevity of a species in genebank storage has been established, whether from the viability equations or, for example, from historical routine viability monitoring data (Hay *et al.*, 2013; van Treuren *et al.*, 2013; Ellis *et al.*, 2018), simple initial storage experiments may be useful. These can determine the relative longevity of different, new seed lots of a species prior to genebank storage in order that accession retest intervals can be customized from



these rankings (Hay and Whitehouse, 2017). The variation in  $\sigma$  observed here also emphasizes the plasticity of seed longevity as a trait (Leprince *et al.*, 2017) and the possibility of developing greater understanding of the temporal effects of low moisture content for seed lots of the same species that differ in longevity (Walters, 2015).

In conclusion, this summary analysis of results with diverse genotypes over five experiments in different seed production environments confirms that seed quality (subsequent seed longevity) in rice is increased substantially by high-temperature drying of seeds harvested before maturation drying ends. The temporal (immediately *ex planta* cf. subsequent storage) and water status discontinuities (above cf. below 16.5%) in the effect of temperature on subsequent air-dry longevity, suggest moist rice seeds exposed to 45°C in late maturation are stimulated to synthesise protectants which stabilise seed during desiccation and aid survival in air-dry storage. The earlier during late maturation that rice seeds are harvested the greater the relative benefit of high-temperature drying to longevity, with no benefit detected once seeds have dried *in planta* to 16.5% moisture content.

### **Acknowledgements**

We thank the staff of the T.T. Chang Genetic Resources Center, especially R. Reanõ, F. de Guzman and Marlina Velasco-Punzalan, for technical assistance. This research was supported by funding from the German Federal Ministry for Economic Cooperation and Development (BMZ) through the Deutsche Gesellschaft für Internationale Zusammenarbeit (GIZ) GmbH.

## References

**Angelovici R, Galili G, Fernie AR and Fait A** (2010) Seed desiccation: a bridge between maturation and germination. *Trends in Plant Science* **15**, 211-218.

**Breese, MH** (1955) Hysteresis in the hygroscopic equilibria of rough rice at 25°C. *Cereal Chemistry* **32**, 481-487.

**Chang TT** (1991) Findings from a 28-year seed viability experiment. *International Rice Research Newsletter* **16**, 5-6.

**Chatelain E, Hundertmark M, Leprince O, Le Gall S, Satour P, Deligny-Pennick S, Rogniaux H and Buitink J** (2012) Temporal profiling of the heat-stable proteome during late maturation of *Medicago trunculata* seeds identifies a restricted subset of late embryogenesis abundant proteins associated with longevity. *Plant Cell and Environment* **35**, 1440-1455.

**Crisostomo S, Hay FR, Reano R and Borromeo T** (2011) Are the standard conditions for genebank drying optimal for rice seed quality? *Seed Science and Technology* **39**, 666-672.

**Cromarty AS, Ellis RH and Roberts EH** (1982) *The Design of Seed Storage Facilities for Genetic Conservation*. Rome, International Board for Plant Genetic Resources.

**Ellis RH** (2011) Rice seed quality development and temperature during late development and maturation. *Seed Science Research* **21**, 95-101.

**Ellis RH and Hong TD** (1994) Desiccation tolerance and potential longevity of developing seeds of rice (*Oryza sativa* L.). *Annals of Botany* **73**, 501-506.

**Ellis RH and Hong TD** (2007) Quantitative response of the longevity of seed of twelve crops to temperature and moisture in hermetic storage. *Seed Science and Technology* **35**, 432-444.

**Ellis RH and Roberts EH** (1980) Improved equations for the prediction of seed longevity. *Annals of Botany* **45**, 13-30.

**Ellis RH, Hong TD and Roberts EH** (1988) A low moisture content limit to logarithmic relations between seed moisture content and longevity. *Annals of Botany* **61**, 405-408.

**Ellis RH, Hong TD and Roberts EH** (1992) The low moisture content limit to the negative logarithmic relation between seed longevity and moisture content in three subspecies of rice. *Annals of Botany* **69**, 53-58.

**Ellis RH, Hong T and Jackson MT** (1993) Seed production environment, time of harvest and the potential longevity of seeds of three cultivars of rice (*Oryza sativa* L.). *Annals of Botany* **72**, 583-590.

**Ellis RH, Nasehzadeh M, Hanson J and Woldemariam Y.** (2018) Medium-term seed storage of 50 genera of forage legumes and evidence-based genebank monitoring intervals. *Genetic Resources and Crop Evolution* **65**, 607-623.

**FAO** (2014) *Genebank Standards for Plant Genetic Resources for Food and Agriculture*. Rome, Food and Agriculture Organisation of the United Nations.

**FAO/IPGRI** (1994) *Genebank Standards*. Rome, Food and Agriculture Organisation of the United Nations/International Plant Genetic Resources Institute.

**Hay FR and Whitehouse KJ** (2017). Rethinking the approach to viability monitoring in genebanks. *Conservation Physiology* 5 (1): cox009;doi:10.1093/conphys/cox009

**Hay FR, de Guzman F, Ellis D, Makahiya H, Borrromeo T and Sackville Hamilton NR** (2013) Viability of *Oryza sativa* (L.) seeds stored under genebank conditions for up to 30 years. *Genetic Resources and Crop Evolution* **60**, 275–296.

**Hay FR, Mead A, Manger K, Wilson FJ.** 2003. One-step analysis of seed storage data and the longevity of *Arabidopsis thaliana* seeds. *Journal of Experimental Botany* **54**, 993-1011.

**Hay FR, Mead A and Bloomberg M** (2014) Modelling seed germination in response to continuous variables: use and limitations of probit analysis and alternative approaches. *Seed Science Research* **24**, 165-186.

**Hong TD, Gedebo A and Ellis RH** (2000) Accumulation of sugars during the onset and development of desiccation tolerance in immature seeds of Norway maple (*Acer platanoides* L.) stored moist. *Seed Science Research* **10**, 147-152.

**IBPGR** (1976) *Report of IBPGR Working Group on Engineering, Design and Cost Aspects of Long-Term Seed Storage Facilities*. Rome, International Board for Plant Genetic Resources.

**IBPGR** (1985) *IBPGR Advisory Committee on Seed Storage: Report of the Third Meeting*. Rome, International Board for Plant Genetic Resources.

**ISTA** (2013) *International Rules of Seed Testing*. Bassersdorf, Switzerland, International Seed Testing Association.

**Kebreab E and Murdoch AJ** (1999) A quantitative model for the loss of primary dormancy and induction of secondary dormancy in imbibed seeds of *Orobanch* spp. *Journal of Experimental Botany* **50**, 211-219.

**Leprince O, Pellizzaro A, Berriri S and Buitink J** (2017) Late seed maturation: drying without dying. *Journal of Experimental Botany* **68**, 827-841.

**Lewis RD** (1950) *Agricultural Research in Texas, 1947-49*. Texas, Texas Agricultural Experiment Station.

**Martínez-Eixarch M and Ellis RH** (2015) Temporal sensitivity of rice seed development from spikelet fertility to viable mature seed to extreme-temperature. *Crop Science* **55**, 354-364.

**Mead A and Gray D** (1999) Prediction of seed longevity: a modification of the shape of the Ellis and Roberts seed survival curves. *Seed Science Research* **9**, 63-73.

**McNally KL, Childs KL, Bohnert R, Davidson RM, Zhao K, Ulat VJ, Zeller G, Clark RM, Hoen DR, Bureau TE, Stokowski R, Ballinger DG, Frazer KA, Cox DR, Padhukasahasram B, Bustamante CD, Weigel D, Mackill DJ, Bruskiewich RM, Rötter G, Buell CR, Leung H and Leach JE** (2009) Genomewide SNP variation reveals relationships among landraces and modern varieties of rice. *Proceedings of the National Academy of Sciences of the USA* **106**, 12273-12278.

**Nasehzadeh M and Ellis RH** (2017) Wheat seed weight and quality differ temporally in sensitivity to warm or cool conditions during seed development and maturation. *Annals of Botany* **120**, 479-493.

**Nellist ME** (1980) Safe drying temperatures for seed grain, pp. 371-388 in Hebblethwaite PD (ed), *Seed Production*. London: Butterworths.

**Porter JR and Gawith M** (1999) Temperatures and the growth and development of wheat: a review. *European Journal of Agronomy* **10**, 23-36.

**Reaño RR, Sackville-Hamilton R and Romero G** (2008) Regeneration guidelines: rice, pp. 1-11 in Dulloo ME, Thormann I, Jorge MA and Hanson J (eds), *Crop Specific Regeneration Guidelines*. Rome, Italy, CGIAR System-wide Genetic Resource Programme.

**Roberts EH and Ellis RH** (1989) Water and seed survival. *Annals of Botany* **63**, 39-52.

**Royal Botanic Gardens Kew** (2018) Seed Information Database (SID). Version 7.1. Available from: <http://data.kew.org/sid/> (January 2018)

**Sanchez B, Rasmussen A and Porter JR** (2014) Temperatures and the growth and development of maize and rice: a review. *Global Change Biology* **20**, 408-417.

**Sanhewe AJ, Ellis RH, Hong TD, Wheeler TR, Batts GR, Hadley P and Morison JIL**

(1996) The effect of temperature and CO<sub>2</sub> on seed quality development in wheat (*Triticum aestivum* L.). *Journal of Experimental Botany* **47**, 631-637.

**Sinniah UR, Ellis RH and John P** (1998) Irrigation and seed quality development in rapid-cycling Brassica: Soluble carbohydrates and heat-stable proteins. *Annals of Botany* **82**, 647-655.

**Tejakhod S and Ellis RH** (2018) Effect of simulated flooding during rice seed development and maturation on subsequent seed quality. *Seed Science Research* (in press).

**Van Treuren R, de Groot EC, van Hintum ThJL. 2013.** Preservation of seed viability during 25 years of storage under standard genebank conditions. *Genetic Resources and Crop Evolution* **60**, 1407-1421.

**Walters C** (2015) Orthodoxy, recalcitrance and in-between: describing variation in seed storage characteristics using threshold responses to water loss. *Planta* **242**, 397-406.

**Wheeler TR, Craufurd PQ, Ellis RH, Porter JR and Vara Prasad PV** (2000)

Temperature variability and the yield of annual crops. *Agriculture, Ecosystems and Environment* **82**, 159-167.

**Whitehouse KJ, Hay FR and Ellis RH** (2015) Increases in the longevity of desiccation-phase developing rice seeds: response to high-temperature drying depends on harvest moisture content. *Annals of Botany* **116**, 245-259.

**Whitehouse KJ, Hay FR and Ellis RH** (2017) High-temperature stress during drying improves subsequent rice (*Oryza sativa* L.) seed longevity. *Seed Science Research* **27**, 281-291.

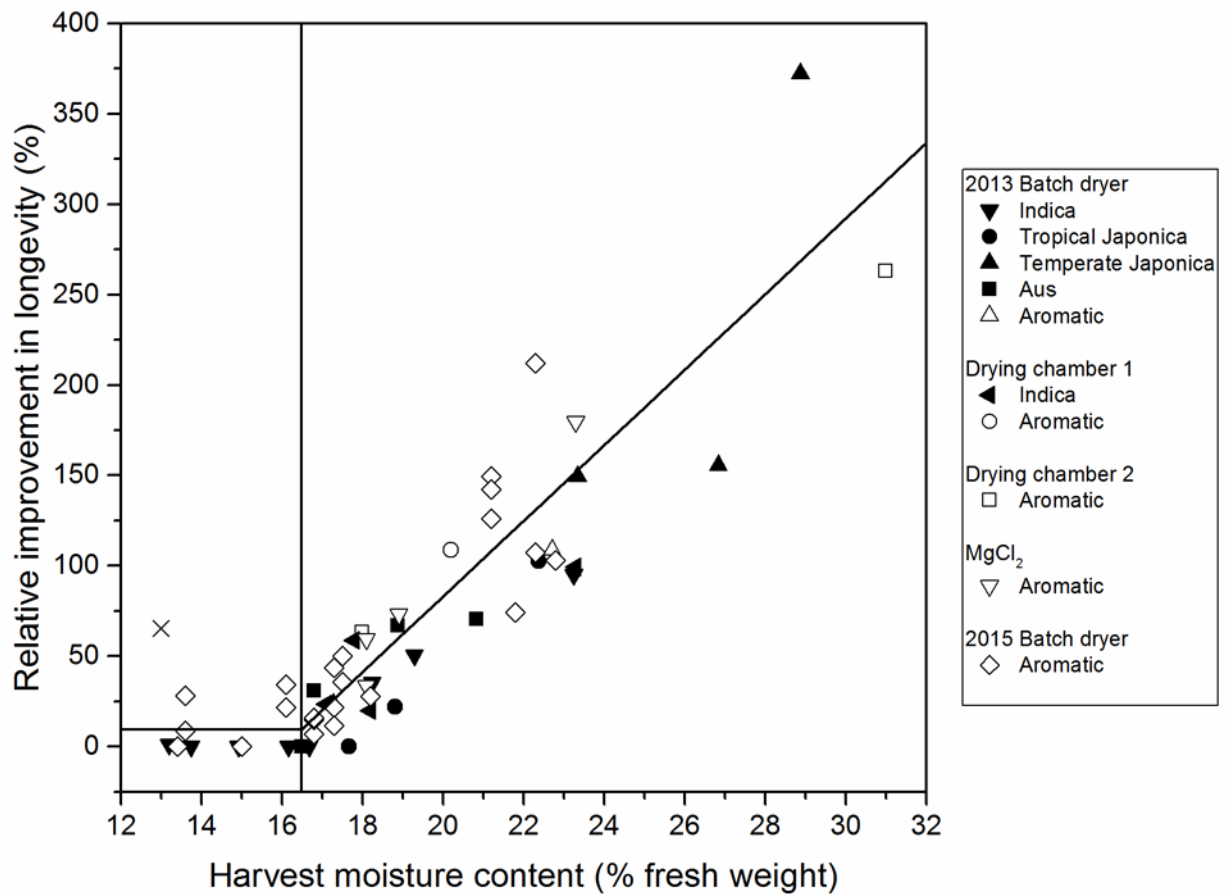
## Legends to Figures

**Figure 1.** Relationship between the improvement in longevity ( $p_{50}$ ) of rice seeds dried initially at 45°C under different conditions (Table 1) relative to that ( $p_{50}$ ) when dried in the dry room (DR) throughout (% , Eqn 4) and harvest moisture content (% , fresh weight). The results are for 20 accessions of different variety types (Indica, tropical Japonica, temperate Japonica, Aus and aromatic; McNally *et al.*, 2009) in 2013, 2014, and/or 2015 between 24 and 60 days after 50% anthesis (Table 1). A relative improvement of 100% is equivalent to a doubling in longevity compared with the control (DR). Some symbols represent multiple treatments (where longevity within an accession did not differ after different durations of high-temperature drying). The solid line is a split-line regression with a breakpoint at 16.5% harvest moisture content for all drying treatments; it accounted for 81.5% of the variance and provided no significant ( $P < 0.05$ ) increase in residual deviance over different regressions for each separate experiment. One outlying observation at 13% moisture content (x) was excluded from analysis (see text).

**Figure 2.** The standard deviation of the frequency of distribution of seed deaths in time,  $\sigma$  (days, logarithmic scale) for seed lots of different variety groups (Indica, tropical Japonica, temperate Japonica, Aus and aromatic; McNally *et al.*, 2009) from different experiments (Table 1) stored hermetically at 45°C with 10.9% moisture content either when first dried to equilibrium to 15% RH (a) in the dry room (DR) only, or (b) initially at 45°C and then in the DR. The symbols in b represent the longest-lived ( $p_{50}$ ) seed lots of all treatments within each experiment. The open symbols represent the aromatic variety, accession IRGC 117265, common to all experiments. The horizontal lines represent independent estimates of  $\sigma$  at 45°C

with 10.9% moisture content: from SID (Royal Botanic Gardens Kew, 2018) provided by extrapolations from observations for Indica (dashed line), tropical Japonica (dotted line), and temperate Japonica varieties of rice (dashed and dotted line) in different environments reported by Ellis *et al.* (1988) or Ellis *et al.* (1992); or from the seed viability constants for rice (solid line) calculated from observations over a wide range of temperatures and moisture contents (Ellis and Hong, 2007) in accordance with Eqn (1).

Figure 1





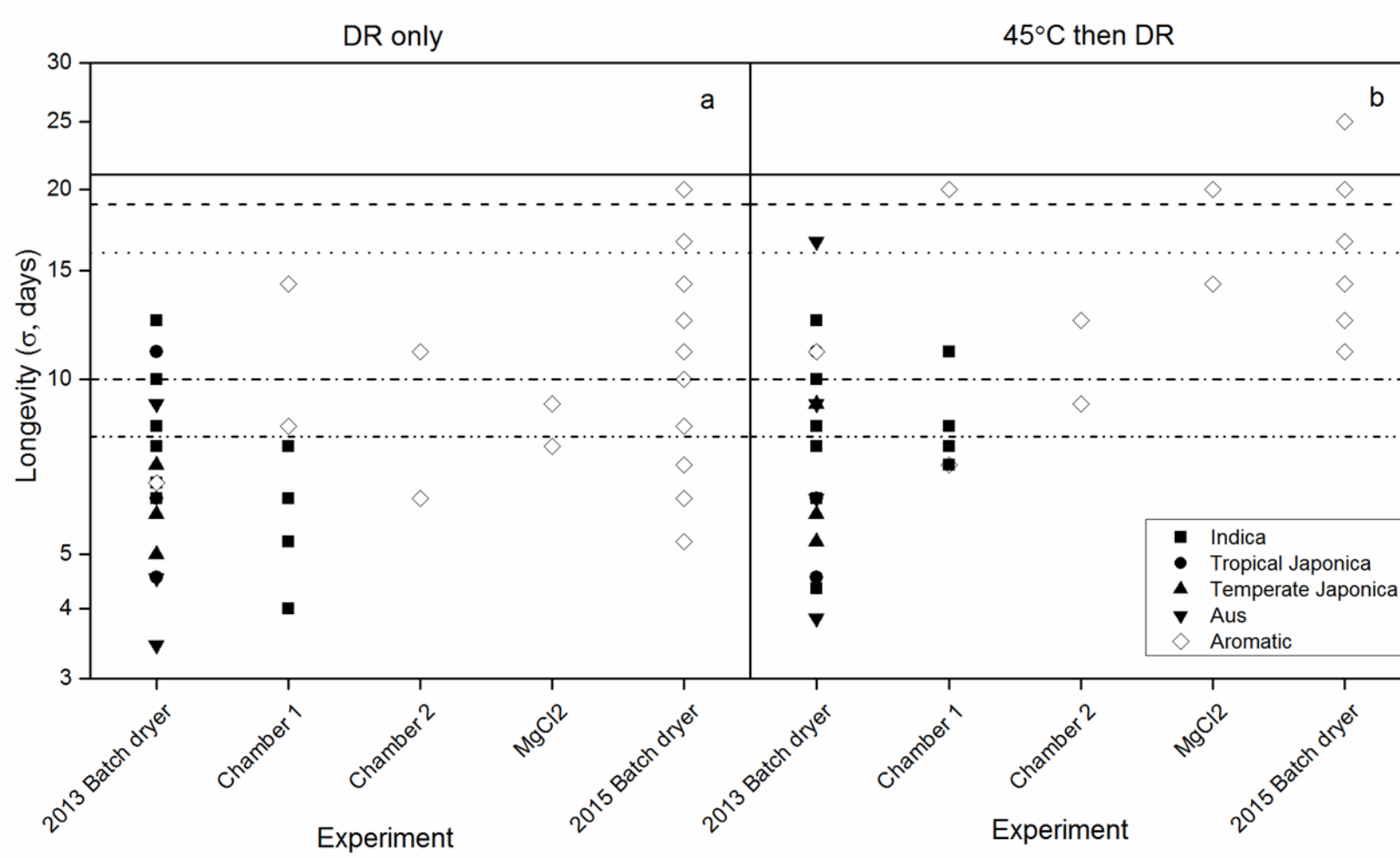


Figure 2

**Table 1.** Number of accessions and respective variety group (with number of accessions in each group) harvested at each maturity stage (DAA; days after 50% anthesis) for each experiment in the crop year and season (dry [DS] or wet [WS]) shown. Seeds were dried following the standard drying protocol (Control; 14 d in the drying room [DR; 15°C/15% RH]), or for an initial period in the drying regime shown before final drying in the DR.

Experiment	Season	Accession Number <sup>¶</sup>	Variety group *	Maturity (DAA)	No. harvests	Drying treatments
<b>2013 Batch dryer</b>	2013DS	IRGC 117265	Aromatic (1)	24-48	1 per accession	Batch dryer (45°C; 8 h d <sup>-1</sup> ) for a maximum of 6 cycles → final drying in DR  + Control (DR)
		IRGC 117266	Aus (4)			
		IRGC 117267				
		IRGC 117273				
		IRGC 117283				
		IRGC 117268	Indica (8)			
		IRGC 117271				
		IRGC 117275				
		IRGC 117276				
		IRGC 117277				
		IRGC 117278				
		IRGC 117280				
		IRGC 117281				
		IRGC 117269	Temperate Japonica (4)			
		IRGC 117270				
		IRGC 117274				
IRGC 117279						
IRGC 117264	Tropical Japonica (3)					
IRGC 117272						
IRGC 117282						

<b>Drying chamber 1</b> (45°C/23% RH)	2014DS	IRGC 117265	Aromatic (1)	35	2 per accession	Continuous drying (24 h d <sup>-1</sup> ) at 45°C/23% RH for a maximum of 5 days → final drying in DR  + Control (DR)
		IRGC 117276 IRGC 117280	Indica (2)			
<b>Drying chamber 2</b> (45°C/75% RH)	2014WS	IRGC 117265	Aromatic	35	2 per accession	Stepped drying over 6 days day 0-1: 45°C/75% RH day 1-3: 30°C/45% RH day 3-5: 20°C/25% RH day 5-6: 15°C/15% RH → final drying in DR  + Control (DR)
<b>MgCl<sub>2</sub></b> (45°C/30% RH)	2015DS	IRGC117265	Aromatic	25, 35, 45	2 per maturity stage	3 days of continuous (24 h d <sup>-1</sup> ) or intermittent (8 h d <sup>-1</sup> ) at 45°C/30% RH → final drying in DR  + Control (DR)
<b>2015 Batch dryer</b> (no misting)	2015DS	IRGC 117265	Aromatic	25, 35, 45	2 per maturity stage	3 days of continuous (24 h d <sup>-1</sup> ) or intermittent (8 h d <sup>-1</sup> ) drying in the batch dryer (45°C)→ final drying in DR  + Control (DR)
<b>2015 Batch dryer</b> (in-field misting)	2015DS	IRGC 117265	Aromatic	25, 35, 45, 50, 55, 60	2 per maturity stage	3 days of continuous (24 h d <sup>-1</sup> ) or intermittent (8 h d <sup>-1</sup> ) drying in the batch dryer (45°C)→ final drying in DR  + Control (DR)

\*McNally *et al.* (2009)

<sup>†</sup>IRGC 117265 (<https://doi.org/10.18730/1PG6J>); -66 (<https://doi.org/10.18730/4WZGR>); -67 (<https://doi.org/10.18730/4WZHS>); -68 (<https://doi.org/10.18730/4WZJT>); -69 (<https://doi.org/10.18730/4WZKV>); -70 (<https://doi.org/10.18730/4WZMW>); -71 (<https://doi.org/10.18730/4WZNX>); -72 (<https://doi.org/10.18730/4WZPY>); -73 (<https://doi.org/10.18730/4WZQZ>); -74 ([https://doi.org/10.18730/4WZR\\*](https://doi.org/10.18730/4WZR*)); -75 (<https://doi.org/10.18730/4WZS~>); -76 ([https://doi.org/10.18730/4WZT\\$](https://doi.org/10.18730/4WZT$)); -77 (<https://doi.org/10.18730/4WZV=>); -78 (<https://doi.org/10.18730/4WZWU>); -79 (<https://doi.org/10.18730/4WZX0>); -80 (<https://doi.org/10.18730/4WZY1>); -81 (<https://doi.org/10.18730/5CMYC>); -82 (<https://doi.org/10.18730/5CMZD>); -83 (<https://doi.org/10.18730/5CN0E>)