

# Up and away: ontogenic transference as a pathway for aerial dispersal of microplastics

Article

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## Up and away: ontogenic transference as a pathway for aerial dispersal of microplastics

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1	Up and away: ontogenic transference as a pathway for aerial dispersal of
2	microplastics
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#### 28 Abstract

29 Microplastics (MPs) are ubiquitous pollutants found in marine, freshwater and terrestrial 30 ecosystems. With so many MPs in aquatic systems it is inevitable that they will be ingested 31 by aquatic organisms, and be transferred up through the food chain. However, to date, no 32 study has considered whether MPs can be transmitted by means of ontogenic transference i.e. 33 between life stages that utilise different habitats. Here, we determine whether fluorescent 34 polystyrene beads could transfer between *Culex* mosquito life stages and, particularly, could 35 move into the flying adult stage. We show for the first time that MPs can be transferred 36 ontogenically from a feeding (larva) into a non-feeding (pupa) life stage and subsequently 37 into the adult terrestrial life stage. However, transference is dependent on particle size, with 38 smaller 2µm MPs transferring readily into pupae and adult stages, whilst 15µm MPs 39 transferred at a significantly reduced rate. Microplastics appear to accumulate in the 40 Malpighian tubule renal excretion system. The transfer of MPs to the adults represents a 41 potential aerial pathway to contamination of new environments. Thus, any organism that 42 feeds on terrestrial life phases of freshwater insects could be impacted by MPs found in 43 aquatic ecosystems.

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45 Keywords	S
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46 Food chain: ontology; life stage; Malpighian tubules, microplastics; Culex pipiens
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#### 54 Introduction

55 Microplastics (MPs) are ubiquitous pollutants found in marine, freshwater and terrestrial 56 There is little doubt that plastic and MP pollution is a major ecosystems [1–3]. 57 environmental concern globally. Despite this, there is relatively little research into the impact 58 of MPs on freshwater ecosystems, with most research concentrating on marine systems and 59 organisms [2]. MPs have been defined as plastic particles smaller than 5mm in size [4,5]. 60 However, this simple description covers a wide range of types, including, among others, 61 polypropylene, polyethylene and polystyrene MPs entering the environment in different 62 shapes and sizes, including fibres, pellets and cosmetic beads [6,7]. MPs are categorised 63 based on their origin as primary or secondary types, depending on whether they were 64 released into the environment as MPs (primary) or have degraded to that size in the 65 environment (secondary) [8,9]. Microplastics pass through terrestrial environments in 66 household wastewater [2,10]. Rivers can subsequently deliver MPs into the sea and lakes, 67 where they can be found in high concentrations [11-13].

68

69 Microplastics are ingested by aquatic organisms, and can be transferred through the food 70 chain in both freshwater and marine environments [14-18]. However, to date no study has 71 considered whether MPs can be transmitted by means of ontogenic transference i.e. between 72 life stages that utilise different habitats. Freshwater environments are inhabited by insects that 73 spend their juvenile stages in water but their adult stages in the terrestrial environment. Such 74 insects include mayflies, dragonflies, midges and mosquitoes, most of which are eaten by 75 terrestrial vertebrates. This raises the potential for MPs to enter terrestrial ecosystems from 76 freshwater habitats aerially via transference to adult invertebrate life stages. Here, we thus 77 determine whether 2 and 15µm fluorescent polystyrene beads could transfer between insect life stages and, particularly, could move into the flying adult stage. Fluorescent beads were selected to enable MPs to be easily detected in the non-feeding stages and also to allow an investigation of location within the body during metamorphosis. The *Culex pipiens* mosquito complex was selected as a model for this study given their worldwide distribution and broad habitat preference [19]. Mosquitoes develop through four feeding larval instars and a nonfeeding pupal stage, and finally emerge into a flying adult.

84

#### 85 Materials and methods

86 For additional details of all methods and analyses, see the electronic supplementary material. 87 Two types of MPs were used: a 2µm fluorescent yellow-green carboxylate-modified polystyrene (density 1.050g/cm<sup>3</sup>, excitation 470nm; emission 505nm, Sigma-Aldrich, UK) 88 89 and a  $15.45+1.1\mu m$  fluorescent dragon green polystyrene (density 1.06 g/cm<sup>3</sup> (5x10<sup>6</sup>) 90 particles/ml, excitation 480nm; emission 520nm, Bangs Laboratories Inc., USA). Four treatments were used; a control with no microplastics, a treatment of  $8 \times 10^5$  2µm particles/ml, 91 a treatment of  $8 \times 10^2$  15µm particles/ml, and a 1:1 mixture of both treatments. Each replicate 92 (five per treatment) contained ten 3<sup>rd</sup> instar C. pipiens larvae in a 50ml glass beaker filled 93 94 with 50ml of tap water. The control and all treatments contained 100mg of pelleted guinea 95 pig food. Treatments were assigned randomly to a position on the laboratory bench to reduce 96 experimental error.

97 One random individual was removed from each beaker when every mosquito had moulted 98 into the 4<sup>th</sup> instar, and again when they pupated or emerged as adults. All samples were then 99 placed in separate 1.5ml Eppendorf tubes and stored at -20 °C prior to examination. 100 Microplastics were extracted from mosquitoes by homogenization and filtration. The filter 101 membrane was examined using an epi-fluorescent microscope (Zeiss Axioskop) under a 20x 102 lens to count the number of fluorescent MPs. Adults were further dissected under a binocular stereo microscope (0.7X-4.5X) to extract the gut and quantify the numbers of MPs under theepi-fluorescent microscope [20].

105 All data were analyzed using the statistical software R v3.4.2 [21]. Microplastic counts were

106 analysed using generalized linear models (GLMs) assuming a quasi-Poisson distribution.

107 Uptake of microplastics was examined with respect to 'particle size', 'treatment' and 'life

108 stage'. We performed model simplification via stepwise removal of non-significant effects.

109 Tukey tests were used post hoc for multiple comparisons.

110

#### 111 **Results**

No MPs were found in control groups of any mosquito life stage. Densities of MPs were significantly different between life stages ( $F_{2, 56}$ =160.42, P<0.001), with MP numbers significantly falling as mosquitoes moved between successive ontogenic levels (all P< 0.001) (Figure 1, Table S1, S2). Microplastic transference to adults was confirmed by fluorescent microscopy where the beads were detected in the adult abdomen, specifically inside the Malpighian tubules (Figure 2).

Significantly more  $2\mu$ m particles were found in mosquito life stages than 15 $\mu$ m particles overall ( $F_{1, 58}$ =303.98, P<0.001). Microplastics uptake was also significantly greater overall in mixed exposure treatments ( $F_{1, 55}$ =6.00, P=0.02). Although 2 $\mu$ m particles were transferred to adults in all instances, we found no transference of 15 $\mu$ m particles following single treatment exposures. However, in the mixed MPs treatment, transference to adults of both 2 $\mu$ m and 15 $\mu$ m particles was evidenced (Figure 1).

124

#### 125 Discussion

Here, we show for the first time that MPs can be transferred ontogenically from a feeding(larval) into a non-feeding (pupal) life stage and subsequently into the flying (adult) life

stage. Transference through to adults was found in both MP sizes, although the larger 15µm MPs were not ingested as readily as the 2µm MPs. Dissection of mosquito adults showed that 2µm MPs accumulated in the renal excretion system of Malpighian tubules which, unlike the gut, pass from larvae to adult stages without visible reorganization [22]. This has been demonstrated previously to provide a physical transport system between stages during metamorphosis for *Pseudomonas* bacteria and seems to be important for ontogenic transmission from larvae to adults [23].

135 Few 15µm MPs were transferred into adults suggesting that MP size is an important factor in 136 ontogenic transfer which could be related to the transfer and accumulation of MPs in the 137 Malpighian tubes. Although the translocation mechanism of MPs to the Malpighian tubules 138 is unclear in mosquitoes, analysis of fish, fiddler crab and marine mussels has demonstrated 139 that MPs can be translocated from gastrointestinal tracts into other tissues in a wide range of 140 phyla [24, 25,26]. Malpighian tubules have an entry point to the gut between the mid- and 141 hindgut of mosquitoes, but the flow of fluid is from the Malpighian tubules to the hindgut 142 [27]. Diptera are known to produce structures called concretions in the Malpighian tubules 143 which have been shown to sequester heavy metals [28]. However, it is unlikely that this 144 pathway would operate with a solid MP.

145 Our results have important implications since any aquatic life stage that is able to consume 146 MPs and transfer them to their terrestrial life stage is a potential vector of MPs onto novel 147 aerial and terrestrial habitats. Ingestion of MP-contaminated organisms by terrestrial 148 organisms is not new [29]. Indeed, the widespread distribution of MPs in marine 149 environments has meant that animals such as fish and shellfish sold for human consumption 150 are contaminated with a range of plastics with a consequent transference of MPs between 151 trophic levels [24]. Unlike MP fibres, which are common in the air and atmosphere, there 152 has been no evidence for MPs being transported into the air [24]. We have demonstrated here

153	that species with aquatic and terrestrial life stages can harbour MPs through their life history.	
154	Adu	Its are predated on emergence by many animals including dipteran flies Empididae and
155	Doli	chopodididae, whilst resting predominantly by spiders and in flight they are the prey of
156	drag	onflies, damselflies, birds (such as swallows and swifts) and bats (31). Where many
157	insec	cts are emerging from a highly contaminated site, the possibility of contamination of
158	these	e predators could be high. Whilst mosquitoes were used here as a model organism, any
159	fresł	water insect that can ingest MPs will likely equally transmit plastics into a terrestrial
160	adul	t stage. This has implications for organisms that feed on adult mosquitoes with aerial and
161	terre	strial animals accordingly open to MP exposure and transference would appear to occur
162	at a 🛛	higher rate for smaller MPs.
163	Ref	erences
164	1.	Sighicelli M, Pietrelli L, Lecce F, Iannilli V, Falconieri M, Coscia L, Di Vito S, Nuglio
165		S, Zampetti G. 2018 Microplastic pollution in the surface waters of Italian Subalpine
166		Lakes. Environ. Pollut. 236, 645-651. (doi:10.1016/j.envpol.2018.02.008)
167	2.	Wagner M, Lambert S. 2018 Freshwater Microplastics. Cham: Springer International
168		Publishing. (doi:10.1007/978-3-319-61615-5)
169	3.	Mason SA, Welch V, Neratko J. 2018 Synthetic polymer contamination in bottled
170		water. Fredonia State Univ. New York, 1–17.
171	4.	Eriksen M, Lebreton LCM, Carson HS, Thiel M, Moore CJ, Borerro JC, Galgani F,
172		Ryan PG, Reisser J. 2014 Plastic Pollution in the World's Oceans: More than 5
173		Trillion Plastic Pieces Weighing over 250,000 Tons Afloat at Sea. PLoS One 9,
174		e111913. (doi:10.1371/journal.pone.0111913)
175	5.	Imhof HK, Ivleva NP, Schmid J, Niessner R, Laforsch C. 2013 Contamination of
176		beach sediments of a subalpine lake with microplastic particles. Curr. Biol. 23, 1-15.
177		(doi:10.1016/j.cub.2013.09.001)
178	6.	Andrady AL, Neal MA. 2009 Applications and societal benefits of plastics. Philos.

179 Trans. R. Soc. Lond. B. Biol. Sci. 364, 1977–1984. (doi:10.1098/rstb.2008.0304)

- 180 7. Rocha-Santos T, Duarte AC. 2014 A critical overview of the analytical approaches to
- the occurrence, the fate and the behavior of microplastics in the environment. *Trends Anal. Chem.* 65, 47–53. (doi:10.1016/j.trac.2014.10.011)
- 183 8. Moore CJ. 2008 Synthetic polymers in the marine environment: A rapidly increasing,
- 184 long-term threat. *Environ. Res.* **108**, 131–139. (doi:10.1016/j.envres.2008.07.025)
- Barnes DKA, Galgani F, Thompson RC, Barlaz M. 2009 Accumulation and
   fragmentation of plastic debris in global environments. *Philos. Trans. R. Soc. Lond. B. Biol. Sci.* 364, 1985–98. (doi:10.1098/rstb.2008.0205)
- Mason SA, Garneau D, Sutton R, Chu Y, Ehmann K, Barnes J, Fink P, Papazissimos
  D, Rogers DL. 2016 Microplastic pollution is widely detected in US municipal
  wastewater treatment plant effluent. *Environ. Pollut.* 218, 1045–1054.
  (doi:10.1016/j.envpol.2016.08.056)
- Eriksen M, Mason S, Wilson S, Box C, Zellers A, Edwards W, Farley H, Amato S.
   2013 Microplastic pollution in the surface waters of the Laurentian Great Lakes. *Mar. Pollut. Bull.* 77, 177–182. (doi:10.1016/j.marpolbul.2013.10.007)
- 195 12. Fischer EK, Paglialonga L, Czech E, Tamminga M. 2016 Microplastic pollution in
  196 lakes and lake shoreline sediments A case study on Lake Bolsena and Lake Chiusi
  197 (central Italy). *Environ. Pollut.* 213, 648–657. (doi:10.1016/j.envpol.2016.03.012)
- 13. Su L, Xue Y, Li L, Yang D, Kolandhasamy P, Li D, Shi H. 2016 Microplastics in
  Taihu Lake, China. *Environ. Pollut.* 216, 711–719. (doi:10.1016/j.envpol.2016.06.036)
- Aljaibachi R, Callaghan A. 2018 Impact of polystyrene microplastics on *Daphnia magna* mortality and reproduction in relation to food availability. *PeerJ* 6, e4601.
  (doi:10.7717/peerj.4601)
- 203 15. Cole M, Lindeque P, Fileman E, Halsband C, Goodhead R, Moger J, Galloway TS.

#### Submitted to Biology Letters

204	2013 Microplastic ingestion by zooplankton. Environ. Sci. Technol. 47, 6646-6655
205	(doi:10.1021/es400663f)

- 206 16. Scherer C, Brennholt N, Reifferscheid G, Wagner M. 2017 Feeding type and
  207 development drive the ingestion of microplastics by freshwater invertebrates. *Sci. Rep.*
- 208 7, 17006. (doi:10.1038/s41598-017-17191-7)
- 209 17. Sussarellu R *et al.* 2016 Oyster reproduction is affected by exposure to polystyrene
  210 microplastics. *Proc. Natl. Acad. Sci.* 113, 2430–2435. (doi:10.1073/pnas.1519019113)
- 18. Messinetti S, Mercurio S, Parolini M, Sugni M, Pennati R. 2018 Effects of polystyrene
  microplastics on early stages of two marine invertebrates with different feeding
  strategies. *Environ. Pollut.* 237, 1080–1087. (doi:10.1016/j.envpol.2017.11.030)
- Dow JA, Maddrell SH, Görtz A, Skaer NJ, Brogan S, Kaiser K. 1994 The malpighian
  tubules of *Drosophila melanogaster*: a novel phenotype for studies of fluid secretion
  and its control. *J. Exp. Biol.* 197, 421–8.
- 217 20. Coleman J, Juhn J, James AAA. 2007 Dissection of Midgut and Salivary Glands from
  218 *Ae. aegypti* Mosquitoes. *J. Vis. Exp.*, 2007. (doi:10.3791/228)
- 219 21. R Development Core Team. 2017 R: A Language and Environment for Statistical
  220 Computing. *R Found. Stat. Comput. Vienna Austria* (doi:10.1038/sj.hdy.6800737)
- 221 22. Clements AN. 1992 *The biology of mosquitoes. Volume 1: Development, nutrition and reproduction.* London: Chapman and Hall.
- 223 23. Chavshin AR, Oshaghi MA, Vatandoost H, Yakhchali B, Zarenejad F, Terenius O.
  224 2015 Malpighian tubules are important determinants of *Pseudomonas* transstadial
- transmission and longtime persistence in *Anopheles stephensi*. Parasites and Vectors
- 226 **8**, 1–7. (doi:10.1186/s13071-015-0635-6)
- 227 24. Von Moos N, Burkhardt-Holm P, Köhler A. 2012 Uptake and Effects of Microplastics
  228 on Cells and Tissue of the Blue Mussel *Mytilus edulis* L. after an Experimental

229 Exposure. *Environ. Sci. Technol.* **46**, 11327–11335.

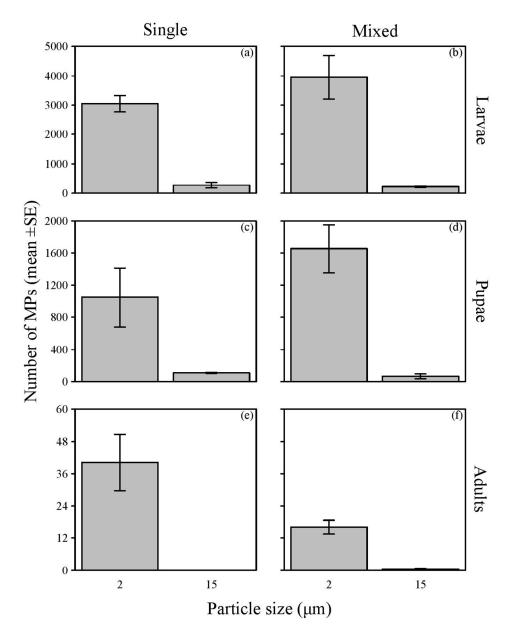
- 230 25. Brennecke D, Ferreira EC, Costa TMM, Appel D, da Gama BAP, Lenz M. 2015
- Ingested microplastics (>100μm) are translocated to organs of the tropical fiddler crab
   *Uca rapax. Mar. Pollut. Bull.* 96, 491–495. (doi:10.1016/j.marpolbul.2015.05.001)
- 233 26. Avio CG, Gorbi S, Regoli F. 2015 Experimental development of a new protocol for
  234 extraction and characterization of microplastics in fish tissues: First observations in
  235 commercial species from Adriatic Sea. *Mar. Environ. Res.* 111, 18–26.
  236 (doi:10.1016/j.marenvres.2015.06.014)
- 237 27. Piermarini PM. 2016 *Renal Excretory Processes in Mosquitoes*. 1st edn. Elsevier Ltd.
  238 (doi:10.1016/bs.aiip.2016.04.003)
- 239 28. Leonard EM, Pierce LM, Gillis PL, Wood CM, O'Donnell MJ. 2009 Cadmium
  240 transport by the gut and Malpighian tubules of *Chironomus riparius*. *Aquat. Toxicol.*241 92, 179–186. (doi:10.1016/j.aquatox.2009.01.011)
- 242 29. Huerta Lwanga E, Gertsen H, Gooren H, Peters P, Salánki T, Van Der Ploeg M,
  243 Besseling E, Koelmans AA, Geissen V. 2016 Microplastics in the Terrestrial
  244 Ecosystem: Implications for *Lumbricus terrestris* (Oligochaeta, Lumbricidae).
  245 *Environ. Sci. Technol.* (doi:10.1021/acs.est.5b05478)
- 30. Dris R, Gasperi J, Mirande C, Mandin C, Guerrouache M, Langlois V, Tassin B. 2017
  A first overview of textile fibers, including microplastics, in indoor and outdoor
  environments. *Environ. Pollut.* (doi:10.1016/j.envpol.2016.12.013)
- 249 31. Medlock JM, Snow, KR. 2008. Natural predators and parasites of British mosquitoes:
- a review. European Mosquito Bulletin, 25 (2008), 1-11.
- 251

#### 252 Figure legends

253 Figure 1. Uptake counts of microplastics (MP) across larval (a, b), pupal (c, d) and adult (e,

- f) Culex mosquito stages following single (a, c, e) and mixed (b, d, f) exposures to  $2\mu m$  and
- $15\mu$ m beads. Means are  $\pm$ SE (*n*=5 per experimental group).
- 256
- 257 Figure 2. Epi-fluorescent microscope images showing fluorescent microplastic particles
- 258 within (A) the abdomen of an adult mosquito before dissection, and (B) the abdominal
- 259 Malpighian tubules following dissection.
- 260
- 261 Ethics
- 262 Ethics committee approval was not required.
- 263 Data accessibility
- 264 Data files are available in online supplementary material.
- 265 Author contribution
- All authors provided substantial contributions to conception and design, or acquisition of
- 267 data, or analysis and interpretation of data; were involved in drafting the article or revising it
- critically for important intellectual content; approved the final version to be published; and
- agree to be accountable for all aspects of the work in ensuring that questions related to the
- accuracy or integrity of any part of the work are appropriately investigated and resolved.
- 271
- 272 **Competing interests**
- 273 We declare we have no competing interests.
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279



Uptake counts of microplastics (MP) across larval (a, b), pupal (c, d) and adult (e, f) Culex mosquito stages following single (a, c, e) and mixed (b, d, f) exposures to  $2\mu$ m and  $15\mu$ m beads. Means are ±SE (n=5 per experimental group).

115x144mm (300 x 300 DPI)

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100 µm

## Malpighian tubules

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100 µm