

Display of a Maize cDNA library on baculovirus infected insect cells

Article

Published Version

Creative Commons: Attribution 3.0 (CC-BY)

Open Access

Harel, H. Y. M., Fontaine, V., Chen, H., Jones, I. M. ORCID: <https://orcid.org/0000-0002-7738-2516> and Millner, P. A. (2008) Display of a Maize cDNA library on baculovirus infected insect cells. BMC Biotechnology, 8 (1). 64. ISSN 1472-6750 doi: <https://doi.org/10.1186/1472-6750-8-64> Available at <https://centaur.reading.ac.uk/9797/>

It is advisable to refer to the publisher's version if you intend to cite from the work. See [Guidance on citing](#).

To link to this article DOI: <http://dx.doi.org/10.1186/1472-6750-8-64>

Publisher: BioMed Central

All outputs in CentAUR are protected by Intellectual Property Rights law, including copyright law. Copyright and IPR is retained by the creators or other copyright holders. Terms and conditions for use of this material are defined in the [End User Agreement](#).

www.reading.ac.uk/centaur

CentAUR

Central Archive at the University of Reading

Reading's research outputs online



Methodology article

Open Access

Display of a Maize cDNA library on baculovirus infected insect cells

Helene Y Meller Harel*¹, Veronique Fontaine², Hongying Chen³,
Ian M Jones³ and Paul A Millner¹

Address: ¹Faculty of biological sciences, University of Leeds, Leeds, LS2 9JT, UK, ²UMR INRA/LISTL 1281, Stress Abiotiques et Différenciation des Végétaux cultivés 2, Chaussée Brunehaut, Estrées-Mons BP 50136, 80203 Péronne cedex, France and ³School of Biological Sciences, University of Reading, Whiteknights, Reading, Berks, RG6 6AJ, UK

Email: Helene Y Meller Harel* - bmbhymh@leeds.ac.uk; Veronique Fontaine - Veronique.Fontaine@mons.inra.fr; Hongying Chen - sas00hc@rdg.ac.uk; Ian M Jones - i.m.jones@rdg.ac.uk; Paul A Millner - p.a.millner@leeds.ac.uk

* Corresponding author

Published: 12 August 2008

Received: 22 April 2008

BMC Biotechnology 2008, 8:64 doi:10.1186/1472-6750-8-64

Accepted: 12 August 2008

This article is available from: <http://www.biomedcentral.com/1472-6750/8/64>

© 2008 Meller Harel et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

Background: Maize is a good model system for cereal crop genetics and development because of its rich genetic heritage and well-characterized morphology. The sequencing of its genome is well advanced, and new technologies for efficient proteomic analysis are needed. Baculovirus expression systems have been used for the last twenty years to express in insect cells a wide variety of eukaryotic proteins that require complex folding or extensive posttranslational modification. More recently, baculovirus display technologies based on the expression of foreign sequences on the surface of *Autographa californica* (AcMNPV) have been developed. We investigated the potential of a display methodology for a cDNA library of maize young seedlings.

Results: We constructed a full-length cDNA library of young maize etiolated seedlings in the transfer vector pAcTMVSVG. The library contained a total of 2.5×10^5 independent clones. Expression of two known maize proteins, calreticulin and auxin binding protein (ABPI), was shown by western blot analysis of protein extracts from insect cells infected with the cDNA library. Display of the two proteins in infected insect cells was shown by selective biopanning using magnetic cell sorting and demonstrated proof of concept that the baculovirus maize cDNA display library could be used to identify and isolate proteins.

Conclusion: The maize cDNA library constructed in this study relies on the novel technology of baculovirus display and is unique in currently published cDNA libraries.

Produced to demonstrate proof of principle, it opens the way for the development of a eukaryotic *in vivo* display tool which would be ideally suited for rapid screening of the maize proteome for binding partners, such as proteins involved in hormone regulation or defence.

Background

With its rich genetic heritage and well-characterized morphology, *Zea mays* (L. ssp. *mays*) is a very important crop and a model system for cereal genetics and development. Its moderately large genome (>2.5 Mb) comprises up to

80% of repeated retrotransposons [1]. It is in the process of being completely sequenced and is the subject of many recent studies [2-4]. In particular, the maize transcriptome provides a source of new functionally important genes and has been recently studied with various methods such

as comparisons between sense and antisense transcriptome [5], serial analysis of gene expression [6], and the construction of PCR-generated, normalized, or subtracted cDNA libraries [7-9].

Display technologies are powerful tools for proteomics studies since they allow direct coupling of proteins with their DNA encoding sequences. The prokaryotic phage display technology is the method of choice when studying non complex proteins but presents significant limitations in the case where folding, post-translational modification or oligomerization of the gene products are needed. Baculovirus surface display is a relatively recent technology, which allows correct expression and presentation of eukaryotic proteins [10]. It is based on the use of baculovirus *Autographa californica* multiple nuclear polyhydrosis virus (AcMNPV) as a vector for expression of a recombinant target sequence on the surface of the virions or the host *Spodoptera frugiperda* (Sf9) cells [11]. The major baculoviral envelope membrane protein, gp64, is usually used as the fusion partner for functional display of a foreign protein or peptide sequence on the viral surface. However, heterologous viral glycoproteins have been shown to be good alternatives to gp64. Such is the case of a truncated form of the vesicular stomatitis virus G glycoprotein (VSV-G) that allows an enhanced level of display as well as alternate presentation of epitopes [12]. Refining the display capabilities of AcMNPV has been the subject of many studies [10,13]. They have led to the development of tools for important applications, such as antigens display for production of antibodies [14], molecular screening and drug delivery into mammalian target cells [15], as well as baculovirus display of peptide libraries for selection of monoclonal antibody epitopes and to identify peptide antigen mimotopes [16,17]. To date however, the only study where baculovirus-based cDNA library were generated did not use baculovirus-infected cells as a display platform [18]. We describe here the construction and preliminary analysis of a baculovirus display cDNA library of maize young etiolated seedlings.

Results and discussion

Construction of a maize cDNA display library

Total RNA from young maize etiolated seedlings was reverse transcribed, converted to double stranded cDNA and cloned into the baculovirus transfer vector pAcTMVSVG. This vector, combining the signal peptide of the single major envelope glycoprotein gp64 from AcMNPV with the cytoplasmic and transmembrane domains of VSV-G vesicular stomatitis virus G glycoprotein, allows a high level of displayed proteins as well as alternate presentation of epitopes independent of gp64 [12]. The estimated size of the plasmid library was 2.5×10^5 independent clones, with a size distribution of cDNA insert ranging from 0.4 to 2 kb.

The resulting library of plasmids constructed in the transfer vector was incorporated into the baculovirus genome by homologous recombination with a modified viral DNA, BAC10:KO₁₆₂₉. This viral DNA cannot produce virus without undergoing recombination resulting in efficient transfer of the complexity of the vector library into the baculovirus background [19]. Following transfection of a mixture of linearised viral DNA with multiple aliquots of the transfer vector recombinant library into Sf9 insect cells, virus was harvested at 3 days post infection and the supernatant pooled to form a master virus library. The library was not grown further to avoid degeneration but was stored frozen at -80°C.

Expression of calreticulin and ABP1

A principal of the system is that any valid cDNA sequence from the maize genome would be expressed as a recombinant protein with its N-terminus fused to the gp64 signal sequence and C-terminus fused to the transmembrane and cytoplasmic domain of the VSV G glycoprotein (Figure 1). Therefore, in order to assess the functionality of the cDNA library, we studied the expression of two well-characterized maize proteins, calreticulin and ABP1.

Calreticulin is a protein abundant in the endoplasmic reticulum (ER) of many eukaryotes, and in particular of maize [20]. ABP1 is an auxin receptor found ubiquitously

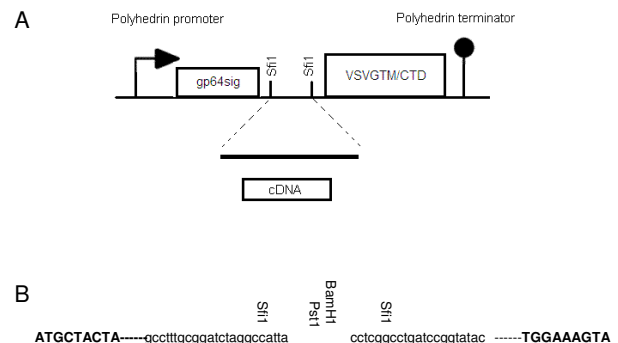


Figure 1

Construction of the display library. cDNA fragments were cloned in *SfiI* sites of pAcTMVSVG vector providing N-terminal fusion with the gp64 signal peptide and C-terminal fusion with the transmembrane (TM) and cytoplasmic terminal domains (CTD) of the VSV G protein. The chimeric genes were expressed under the control of polyhedrin promoter and terminator. **A**, Schematic representation of baculovirus vector pAcTMVSVG. **B**, Details of the cloning site with, in bold, the start and stop sequences from gp64 and VSV G respectively, in lower case, the sequences of the vector around the *SfiI* cloning site, in vertical-orientation, the restriction sites in the multiple cloning site.

in vascular plants and originally identified in maize (*Zea mays*) [21] as a soluble protein, a proportion of which is present on the outer surface of the cell [22].

The presence of maize calreticulin and ABP1 cDNAs in the maize seedlings cDNA library was shown directly by PCR on the plasmid library prepared, using primers specific to calreticulin and ABP1 sequences which revealed products of the expected sizes (670 bp and 490 bp respectively, results not shown).

The correct expression of calreticulin and ABP1 in insect cells infected with the baculovirus stock of the total cDNA library was studied by immunoblotting. On the blots probed with polyclonal antiserum raised against maize calreticulin [20] or maize ABP1 [23], polypeptides were detected in the infected extracts (Figure 2, lane 2 and Figure 3 lane 2) but not in the extract from non-infected cells (Figure 2, lane 1 and Figure 3, lane 2). The polypeptides migrated to a molecular weight corresponding to maize calreticulin (48 kDa) and to glycosylated ABP1 (22 kDa), respectively. The clear expression of glycosylated ABP1 in our system is consistent with previous work showing expression of an active ABP1 from its cDNA in a baculovirus system [24]. It was one of the first examples illustrating that insect cells were appropriate hosts for the expression of plant genes: ABP1 was correctly processed and glycosylated in the endo-membrane system of insect cells. In the case of calreticulin, expression in a baculovirus system of a plant calreticulin is shown here for the first time in this study, although human calreticulin has been co-expressed in a baculovirus system by others [25].

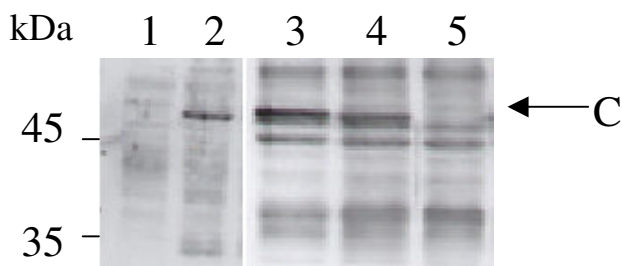


Figure 2
Expression and display of calreticulin in Sf9 cells infected by the cDNA library. Crude cell extracts (10 μ g of proteins) from non-infected Sf9 cells (lane 1), infected cells before panning (lane 2, 3), positive sorted cells after panning (lane 4) or negative sorted cells after panning (lane 5) were run on a SDS-PAGE (12%) and analysed by Western Blot against maize (lane 1, 2) or tobacco anti-calreticulin antiserum, (lane 3, 4, 5). Position of size marker is indicated on the left. Arrow points to calreticulin (C) band.

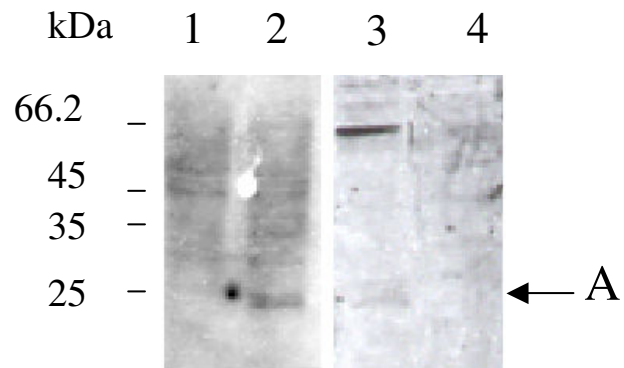


Figure 3
Expression and display of ABP1 in Sf9 cells infected by the cDNA library. Crude cell extracts (10 μ g of proteins) from non-infected Sf9 cells (lane 1), infected Sf9 cells before panning (lane 2), positive sorted cells after panning (lane 3), or negative sorted cells after panning (lane 4) were run on a SDS-PAGE (12%) and analysed by Western Blot against maize anti-abp1 antiserum. Position of size marker is indicated on the left. Arrow points to ABP1 (A) band.

Display of calreticulin and ABP1

Similarly to what was shown with eGFP expression using pActMVSVC vector [12], we expected the expressed proteins to complete the secretory pathway and be retained on the infected cell surface. In order to isolate such cells, we used the magnetic cell sorting (MACS) technology [26]. The indirect magnetic cell labelling was conducted using a primary antibody directed against the protein of interest and anti-immunoglobulin coated beads. MACS technology has been already used for selection of baculovirus-infected cells vs. non-infected ones [27]. In our case however, magnetic cell sorting is used for the biopanning of specific infected cells according to the expression of a surface protein, calreticulin or ABP1, interacting with a specific ligand, the primary antibody. The high conservation of calreticulin sequence in plants allowed us to use an antibody directed against tobacco calreticulin [28]. It showed good recognition of the maize recombinant calreticulin (Figure 2, lane 3) and was used to magnetically label cells displaying calreticulin. The maize ABP1 polyclonal antiserum described before was used to magnetically label cells displaying ABP1.

Virus from positively and negatively biopanned cells were amplified and used to infect fresh insect cells. Expression of the proteins was studied by immunoblotting, as previously described. Maize calreticulin is clearly revealed as a strong band in protein extracts from magnetically labelled cells (Figure 2, lane 4), but cannot be detected in protein extracts from non-labelled cells (Figure 2, lane 5), as expected. Maize ABP1 is also specifically detected,

although as a minor band, in labelled cell extracts but not in non-labelled extracts (Figure 3 lane 3 and 4). The weak signal obtained could result from a partial display of the recombinant protein or from a weak recognition by the first antibody because of unspecific interaction with the second antibody. The strong band visible in lane 4 could correspond to a cross-reaction between the first and the second antibody, as was noted in a study showing ABP1 immunoprecipitation [24].

We conclude therefore that calreticulin and ABP1 are expressed and displayed by insect cells infected by baculovirus containing the whole maize cDNA, demonstrating the proof of principle that our novel maize cDNA display library constructed using recombinant baculoviruses can be used to isolate proteins.

Conclusion

We have constructed a maize cDNA display library in the baculovirus transfer vector pAcTMVSVG, and have screened it by magnetic cell sorting for the expression and the display of two known proteins, calreticulin and ABP1.

Both proteins are moderately abundant in maize, their expression being inducible by various developmental stage or stimuli [20,22,29]. The method of detection by magnetic cell sorting was sensitive enough to reveal their presence on insect cells infected by the cDNA library. Based on the ease of identification, we would expect to detect proteins encoded by less abundant mRNAs as well, especially after normalization and/or subtraction of the amplified library [30].

The library described here is based on baculovirus-infected cells as a display platform, and not only as an expression system as it is the case in the only other published baculovirus-based cDNA expression library [18]. For this reason, it is unique in currently published cDNA library. In addition, it is also, to our knowledge, the first cDNA display library generated from *Zea mays*.

The presented study advances the development and use of cDNA display libraries expressed in baculovirus that could enhance gene discovery. In particular, over 2 million maize cDNA sequences are now available [2] which could be a source for discovery of new encoded proteins using this new eukaryotic *in vivo* display tool.

Fluorescence activated cell sorting (FACS) is the classical methodology used for selection of molecules in a baculovirus surface display, as in the case of displayed peptide sequences [14]. Here we report the use of MACS for an efficient biopanning of baculovirus-infected insect cell displaying a desired protein and therefore binding to a specific antibody. The same principle of methodology

could be very useful in addressing proteomic questions, such as rapid screening of the maize proteome for binding partners, for example proteins involved in hormone regulation or defence. This eukaryotic *in vivo* display tool would ultimately allow biopanning of a cDNA library displayed on the virus surface with a target ligand of interest, thus allowing rapid screening of a proteome for binding partners.

Methods

Library construction in the pACMNPVTM vector

Total RNA (1.5 µg) was extracted from etiolated 5 days old maize seedlings grown under dark conditions using the RNeasy Plant Mini Kit (Qiagen Ltd., UK). After DNase treatment (DNA-free™ kit from Ambion Applied Biosystem, UK), mRNA was isolated using the Fast Track2 kit (Invitrogen Ltd., UK). Double stranded cDNA was synthesised using the Smart™ cDNA library construction kit (Clontech UK): single-stranded cDNAs were obtained by reverse transcription, tagged by the SMART IV oligonucleotide, which allow template switching and synthesis of second strand by the reverse transcriptase. Subsequently, the double stranded cDNA were enriched by Long Distance PCR (LD PCR). The PCR amplification products were subjected to *Sfi*I digestion and size fractionation. cDNA *Sfi*I fragments of 0.2 to 4 kb were then ligated to pACVSVGTM vector linearised with *Sfi*I. Test ligations conducted with 20 ng of vector and 20–50 ng of cDNA were electroporated into *Escherichia coli*. Optimal ratio of cDNA to vector allowing size distribution of cDNA inserts from 0.4 to 2 kb was detected by colony PCR using the primers pACsfilfw: 5'-gcctttgcggatctaggccatta-3' and pACsfilrev:5'-gtataccggatcaggccgagg-3' (Invitrogen Ltd., UK) and Biotaq Red DNA polymerase (Bioline Ltd., UK). The selected ligations were transformed into SURE2 super-competent *Escherichia coli* (Stratagene, UK), or electrotransformed into *E. coli* DH5α according to the manufacturer's protocol (Bio-Rad, UK). The number of independent bacterial clones obtained were quantified to estimate the size of the library, and plasmid midi-preps (Qiagen Ltd., UK) were prepared and pooled together.

Transfection to Insect cells

Insect *S. frugiperda* (Sf9) cells were plated at 5×10^5 cells/well in a six-well plate and allowed to attach for at least 30 minutes. After one wash, the cells were resuspended in 1 ml of serum-free medium SF-900 II SFM (Invitrogen Ltd., UK). For each transfection, 0.1 µg of linearised BAC10:KO₁₆₂₉ DNA was mixed with 0.5 µg of cDNA plasmids library in a total volume of 12 µl. Lipofectin (Invitrogen Ltd., UK) (8 µl) was diluted with 4 µl of water and then mixed with the DNA mixture. Following an incubation for 15-min at room temperature, the DNA-lipofectin complexes were added to the cell monolayers and incubated overnight at 28°C. The cells were then washed with

fresh SF-900 II SFM medium and incubated for 4 days at 28°C. The culture was centrifuged to remove cells and the supernatant was kept at 4°C. The virus stock was titrated by plaque assays by absorbing serial dilutions of the viral preparation for 1 to 2 hours onto 2 ml of Sf9 cells 50% confluent in 6-well-plates. After incubation, viruses were removed and 2 ml of 1% agar (w/v) in SF-900 II SFM medium were added to each well and left to solidify. SF-900 II SFM medium (1 ml) was then added to each well and the plates were incubated at 28°C for 3 to 4 days. At the end of the incubation the top medium was removed and the cells were stained with Neutral red (1/13 dilution of 0.33% (v/v) stock solution) for 1 to 2 hours, and the plaques were counted after 3 hours.

Magnetic biopanning of the library

The maize cDNA library was biopanned using the magnetic sorting technology (Miltenyi Biotec Ltd., UK). Insect cells (10^8) were infected with the virus stock (MOI of 1 pfu/cell), harvested at 2 days post-infection, and resuspended in 1 ml of buffer PBS pH7.2 (137 mM NaCl, 2.7 mM KCl, 8 mM Na₂HPO₄, 2 mM KH₂PO₄) supplemented with 0.5% BSA (w/v) and 2 mM EDTA. Cells were then labelled for 5 minutes on ice with primary antibody previously incubated for 60 minutes at 28°C with non-infected cells. Cells were washed twice, resuspended in 1 ml buffer, and subsequently coated with anti-rabbit magnetic beads for 15 minutes at 4°C. After being washed and resuspended in 1 ml buffer, cells were passed through pre-separation filter and subjected to magnetic separation through a cell-separation MS column ((Miltenyi Biotec Ltd., UK) according to the manufacturer protocol. Magnetically labelled cells bound to the column were flushed out with 2 to 3 ml of SF-900 II SFM medium. Both the non-labelled cells coming in the flowthrough, and the labelled cells were incubated at 28°C for one day in SF900 II SFM serum-free medium. Virus stocks were then amplified for 4 days, and stored at -20°C in SF-900 II SFM medium with 2% serum (w/v) for further studies.

SDS-PAGE and Immunoblotting

Insect cells (10^6 /ml) were infected with virus stocks for 3 days (MOI of 1 pfu/cell) in 6 wells plates and protein extracts from cell pellets were collected by resuspending pellets in PBS pH 7.4 and heating for 5 minutes at 85°C in SDS sample buffer (100 mM Tris pH6.9, 4% SDS (w/v), 20% Glycerol (v/v), 0.1% Bromophenol Blue (v/v), 250 mM DTT). Protein contents were then resolved by 12% SDS-PAGE with a 14.4–116 kDa molecular weight marker (Fermentas, UK) and electrotransferred to nitrocellulose membrane using a Trans-Blot SD semi-dry electrophoretic transfer cell (Bio-Rad, UK). After being washed, the membrane was blocked with 5% milk (w/v) powder in PBST (PBS containing 0.1% Tween20), and incubated with antibodies raised against maize or tobacco proteins

diluted 10^{-4} in PBST at room temperature for 1 h. Polyclonal serum against maize proteins, calreticulin and ABP1, and tobacco calreticulin were kindly supplied by Prof. R. Napier (HRI, UK) and Dr. J. Denecke (University of Leeds, UK), respectively. After six washes with PBST, the membrane was then incubated with anti-rabbit IgG HRP-linked whole antibody (Amersham plc, UK) diluted 10^{-3} , washed, and detected by ECL detection (Amersham plc, UK), as recommended by the manufacturer.

Authors' contributions

HMH carried out most of the practical work of the study and the redaction of the manuscript. VF carried out the synthesis of double-strand cDNA from maize RNA, and their digestion. HC and IMJ conceived the technological approach of the study and guided the transfection and biopanning steps. IMJ and PAM helped to draft the manuscript. PAM initiated the concept of the study, participated in its design and coordination. All authors read and approved the final manuscript.

Acknowledgements

The research was funded by The Leverhulme trust. Advice for baculovirus work was kindly provided by Dr. Alan Cox and Mrs Elizabeth McIntosh in the laboratory of Prof. John Findlay (University of Leeds, UK). Useful feedbacks on the draft of the manuscript were kindly provided by Dr. Wayne Charlton and Dr. Aysha Divan (University of Leeds, UK).

References

- Messing J, Dooner HK: **Organization and variability of the maize genome.** *Curr Opin Plant Biol* 2006, **9**:157-63.
- Messing J, Bharti AK, Karlowski WM, Gundlach H, Kim HR, Yu Y, Wei F, Fuks G, Soderlund CA, Mayer KFX, Wing RA: **Sequence composition and genome organization of maize.** *Proc Natl Acad Sci* 2004, **101**:14349-14354.
- Rabinowicz PD, Bennetzen JL: **The maize genome as a model for efficient sequence analysis of large plant genomes.** *Curr Opin Plant Biol* 2006, **9**:149-156.
- The TIGR Maize Database** [<http://maize.tigr.org/>]
- Ma J, Morrow DJ, Fernandes J, Walbot V: **Comparative profiling of the sense and antisense transcriptome of maize lines.** *Genome Biol* 2006, **7**:R22.
- Poroyko V, Hejlek LG, Spollen WG, Springer GK, Nguyen HT, Sharp RE, Bohnert HJ: **The maize root transcriptome by serial analysis of gene expression.** *Plant Physiol* 2005, **138**:1700-1710.
- Breton C, Chaboud A, Matthys-Rochon E, Bates EEM, Cock JM, Fromm H, Dumas C: **PCR-generated cDNA library of transition-stage maize embryos: cloning and expression of calmodulin genes during early embryogenesis.** *Plant Mol Biol* 1995, **27**:105-113.
- Emberton J, Ma J, Yuan Y, SanMiguel P, Bennetzen JL: **Gene enrichment in maize with hypomethylated partial restriction (HMPR) libraries.** *Genome Res* 2005, **15**:1441-1446.
- Lê Q, Gutiérrez-Marcos JF, Costa LM, Meyer S, Dickinson HG, Lörz H, Kranz E, Scholten S: **Construction and screening of subtracted cDNA libraries from limited populations of plant cells: a comparative analysis of gene expression between maize egg cells and central cells.** *Plant J* 2005, **44**:167-178.
- Grabherr R, Ernst W, Oker-Blomb C, Jones I: **Developments in the use of baculoviruses for the surface display of complex eukaryotic proteins.** *Trends Biotechnol* 2001, **19**:231-236.
- Boublik Y, Di Bonito P, Jones IM: **Eukaryotic virus display: engineering the major surface glycoprotein of the *autographa californica* nuclear polyhedrosis virus (AcNPV) for the presentation of foreign proteins on the virus surface.** *Biotechnology* 1995, **13**:1079-1084.

12. Chapple SDJ, Jones IM: **Non-polar distribution of green fluorescent protein on the surface of *Autographa californica* nucleopolyhedrovirus using a heterologous membrane anchor.** *J Biotechnol* 2002, **95**:269-275.
13. Mäkelä AR, Oker-Blom C: **Baculovirus display: a multifunctional technology for gene delivery and eukaryotic library development.** *Adv Virus Res* 2006, **68**:91-112.
14. Lindley KM, Su JL, Hodges PK, Wisely GB, Bledsoe RK, Condreay JP, Winegar DA, Hutchins JT, Kost TA: **Production of monoclonal antibodies using recombinant baculovirus displaying gp64-fusion proteins.** *J Immunol Methods* 2000, **234**:123-135.
15. Makela A, Oker-Blom C: **The baculovirus display technology – an evolving instrument for molecular screening and drug delivery.** *Comb Chem High Throughput Screen* 2008, **11**:86-98.
16. Ernst W, Grabherr R, Wegner D, Borth N, Grassauer A, Kattinger H: **Baculovirus surface display: construction and screening of a eukaryotic epitope library.** *Nucleic Acid Res* 1998, **26**:1718-1723.
17. Crawford F, Jordan KR, Stadinski B, Wang Y, Huseby E, Marrack P, Slansky JE, Kappler JW: **Use of baculovirus MHC/peptide display libraries to characterize T-cell receptor ligands.** *Immunol Rev* 2006, **210**:156-170.
18. Granziero L, Nelboeck P, Bedoucha M, Lanzavecchia A, Reid HH: **Baculovirus cDNA libraries for expression cloning of genes encoding cell-surface antigens.** *J Virol Met* 1997, **53**:263-272.
19. Zhao Y, Chapman DA, Jones IM: **Improving baculovirus recombination.** *Nucleic Acids Res* 2003, **31**:E6-6.
20. Baluska F, Samaj J, Napier R, Volkman D: **Maize calreticulin localizes preferentially to plasmodesmata in root apex.** *Plant J* 1999, **19**:481-488.
21. Napier RM, David KM, Perrot-Rechenmann C: **A short history of auxin-binding proteins.** *Plant Mol Biol* 2002, **49**:339-348.
22. Diekmann W, Venis MA, Robinson DG: **Auxins Induce Clustering of the Auxin-Binding Protein at the Surface of Maize Coleoptile Protoplasts.** *Proc Natl Acad Sci* 1995, **92**:3425-3429.
23. Napier RM, Venis MA, Bolton MA, Richardson LI, Butcher GW: **Preparation and characterization of monoclonal and polyclonal antibodies to maize auxin-binding protein.** *Planta* 1988, **176**:519-526.
24. Macdonald H, Henderson J, Napier RM, Venis MA, Hawes C, Lazarus CM: **Authentic Processing and Targeting of Active Maize Auxin-Binding Protein in the Baculovirus Expression System.** *Plant Physiology* 1994, **105**:1049-1057.
25. Tate CG, Whiteley E, Betenbaugh MJ: **Molecular chaperones stimulate the functional expression of the cocaine-sensitive serotonin transporter.** *J Biol Chem* 1999, **274**:17551-17558.
26. Miltenyi S, Müller W, Weichel W, Radbruch A: **High gradient magnetic cell separation with MACS.** *Cytometry* 1990, **11**:231-238.
27. Philipps B, Forstner M, Mayr LM: **Baculovirus expression system for magnetic sorting of infected cells and enhanced titer determination.** *Biotechniques* 2004, **36**:80-83.
28. Denecke J, Carlsson LE, Vidal S, Höglund AS, Ek B, Van Zeijl MJ, Sinjorgo KMC, Palva ET: **The tobacco homolog of mammalian calreticulin is present in protein complexes in vivo.** *Plant Cell* 1995, **7**:391-406.
29. Heilmann I, Shin J, Huang J, Perera IY, Davies E: **Transient dissociation of polyribosomes and concurrent recruitment of calreticulin and calmodulin transcripts in gravistimulated maize pulvini.** *Plant Physiol* 2001, **127**:1193-1203.
30. Bonaldo MF, Lennon G, Soares MB: **Normalization and subtraction: two approaches to facilitate gene discovery.** *Genome Res* 1996, **6**:791-806.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

